

**Phytoplankton Growth and Diel Variations in Beam Attenuation
through Individual Cell Analysis**

by

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B.A., 1987, Carleton College

Submitted in partial fulfillment of the requirements for the degree of

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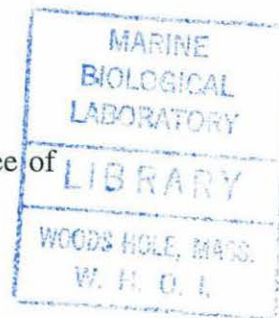
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Submitted to the Department of Biology in July 1995 in partial fulfillment of the requirements for the Degree of Doctor of Philosophy.

Abstract

A number of investigators have observed diel variations in the bulk water inherent optical property beam attenuation, with a minimum near dawn and a maximum near dusk, and have assumed them to be caused by the phytoplankton. In an attempt to understand these patterns, mean forward light scatter of different populations of phytoplankton, from flow cytometric analysis of individual cells, was determined and found to show similar diel patterns for populations of pico- and nanophytoplankton measured in the equatorial Pacific and the Sargasso Sea. The cell concentration patterns do not generally correspond to those of beam attenuation. Laboratory experiments, combined with theoretical calculations, were undertaken to attempt to account for the diel variations in beam attenuation observed at sea and to estimate *in situ* phytoplankton group-specific growth and loss rates.

To investigate how cell growth and division affect the optical properties of phytoplankton, cultures of the chlorophyte *Nannochloris* sp. were sampled over a diel cycle to measure cell size and concentration, light attenuation and absorption, flow cytometric light scattering (in forward and side directions), and carbon content. In addition, the refractive index was calculated using the anomalous diffraction approximation of Mie theory. At six different light levels ranging from 60 - 1500 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ (specific growth rates from ~ 0.2 to $\sim 0.7 \text{ d}^{-1}$), cell division was tightly phased to the light:dark cycle, occurring soon after dark. There were pronounced diel patterns, with a minimum near dawn and a maximum near dusk, in cell size and in cell-specific beam attenuation, absorption, flow cytometric light scatter and carbon. The diel variations in the attenuation cross section were primarily influenced by the changes in cell size due to growth and division, while changes in refractive index had only a small effect. Because eukaryotic cells in the size range of *Nannochloris* are major constituents of many phytoplankton communities, these results have important consequences for the interpretation of diel variations in optical properties observed in the ocean.

To interpret field data of diel variations in red beam attenuation, the relationships between cell optical properties determined in the laboratory were applied to three diel sampling experiments in the equatorial Pacific and four in the Sargasso Sea. In the equatorial Pacific in April and October 1992, the phytoplankton biomass was dominated by picophytoplankton (*Prochlorococcus* and *Synechococcus*), and by mixed populations of ultraphytoplankton (1-2 μm diameter) and nanophytoplankton (2-20 μm , mostly 2-3 μm). Flow cytometric measurements of

mean forward light scatter of each of these populations showed typical diel patterns which were similar to those of bulk beam attenuation due to particles, whereas cell concentration changes were not. Using a combination of empirical calibrations relating beam attenuation to flow cytometric measurements of pure cultures of phytoplankton in the laboratory, and Mie theory, the contributions of different groups of phytoplankton to the diel variations in beam attenuation were estimated. The results indicate that the phytoplankton assemblage measured by flow cytometry can account for essentially all of the diel variation in the beam attenuation signal. In most instances the nanophytoplankton were the largest contributor to total beam attenuation due to phytoplankton, but the ultraphytoplankton usually were more important in determining the diel variations in this property. *Prochlorococcus* were a smaller but appreciable contributor to beam attenuation changes, and *Synechococcus* were much less important.

A similar analysis was performed for diel sampling experiments in the Sargasso Sea in January 1992, July 1993, and May 1994. The bulk beam attenuation due to particles was strongly correlated with calculated beam attenuation due to phytoplankton. During the July 1993 diel sampling, when pico- and nano-phytoplankton populations were analyzed, in most instances the nanophytoplankton were the largest contributor to total beam attenuation due to phytoplankton, but *Prochlorococcus* were equally important at 70 m for some time points over the diel cycle. In the upper 40 m, *Prochlorococcus* were a smaller contributor to beam attenuation changes than the nanophytoplankton, and *Synechococcus* were even less important. These findings emphasize the need to characterize the composition of the phytoplankton community in order to use beam attenuation to monitor productivity.

Flow cytometric measurements of phytoplankton light scattering and cell concentration over the diel light cycle were used to estimate *in situ* phytoplankton group-specific growth and loss rates in the equatorial Pacific and the Sargasso Sea. Measurements of forward light scatter were converted to cell volume and carbon using laboratory and theoretically derived calibration factors for specific groups of phytoplankton, including *Prochlorococcus*, *Synechococcus*, ultraphytoplankton, nanophytoplankton, and coccolithophores. Assuming that division was phased, specific growth rates were estimated based on volume and carbon changes between minimum and maximum values over the day. Phytoplankton group-specific loss rates, and also separate day and night loss rates, were estimated from the calculated growth rates and measured cell concentrations over time. The method used to estimate growth rates works well for *Prochlorococcus* and appears to work for small eukaryotic phytoplankton, but leads to underestimates for *Synechococcus* and larger eukaryotes. Estimated growth rates for *Prochlorococcus* reached one division per day in the upper waters of the equatorial Pacific, but were about half that in the Sargasso Sea. For the eukaryotic phytoplankton, growth rates in the equatorial Pacific were near one division per day in the upper waters; in the Sargasso Sea, the growth rates approached that, but were more often lower. In general, phytoplankton growth rates were closely matched by *in situ* loss rates over the course of a day.

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Chapter 1

Introduction

There is a wide spectrum of approaches in studying phytoplankton growth and productivity in the ocean, including cell cycle markers, biochemical rate measurements, and cage and bottle incubations (reviewed by Furnas 1990). Currently, there is increasing interest in using optical measurements such as ocean color and beam attenuation, which can be made remotely or from a mooring, to provide convenient and efficient estimates of ocean biomass and primary productivity on large scales. The use of ocean color to estimate primary production, with bio-optical algorithms to interpret the remote measurements in terms of biogeochemical processes, is based on the fact that the color of the ocean is determined by the optical properties of the suspended particles, particularly the phytoplankton and their products (Lewis and Cullen 1991). Beam attenuation, which measures the combined effects of scattering and absorption when a beam of light travels through the water, can be partitioned into the contributions of pure seawater (which is known), dissolved organic material (which is negligible at the red wavelength of the SeaTech beam transmissometer), and particles. Thus an understanding of the variability in optical properties among the particles in the ocean can help us to better interpret these bulk measurements.

The absolute dependence of phytoplankton growth on light has a consequence that one of the most important scales of variation is over the daily cycle of light and

dark. Many researchers have observed diel variations in bio-optical properties in the ocean (Dickey et al. 1990; Hamilton et al. 1990; Stramska and Dickey 1992), and diel variations in beam attenuation have been used to estimate primary production (Siegel et al. 1989; Cullen et al. 1992) by assuming a constant carbon-specific beam attenuation and applying it to the diel measurements of beam attenuation. In order to more completely understand beam attenuation and relate it to productivity, it is necessary to understand the particles and processes that cause the observed patterns. In this thesis, I will use individual cell analysis from flow cytometry to interpret measurements of beam attenuation in terms of the contribution of different phytoplankton groups to the overall signal and to the diel variations.

A second aspect of this thesis is the interpretation of diel variations in flow cytometric light scatter of phytoplankton as *in situ* growth rates. From flow cytometric measurements of phytoplankton cells sampled at different times of the day (Olson et al. 1990) or over a diel cycle (Olson et al. 1991), it has been seen that the mean forward light scatter of a population of cells increases during the day and decreases at night. These observations have been made for populations of the picoplankton *Prochlorococcus* and *Synechococcus* and also for eukaryotic nanophytoplankton. It appears that these diel patterns in forward light scatter reflect cell growth during the day followed by division at night. In this thesis, I will use diel variations in flow cytometric measurements of cell light scatter for specific types of phytoplankton to estimate *in situ* phytoplankton group-specific growth rates and, with the concomitant cell concentration measurements, loss rates in the ocean.

The organization of the thesis is as follows. I first present the results of laboratory experiments on optical properties of cultures of the chlorophyte *Nannochloris* sp., which has a size and shape similar to open ocean nanophytoplankton (Chapter 2). In these diel sampling experiments, I determine cell size and concentration, light attenuation and absorption, flow cytometric light scattering, and carbon content of *Nannochloris* grown on a light:dark cycle at a range of light levels. I then present in Chapter 3 field observations from diel sampling experiments in the equatorial Pacific, where flow cytometric measurements of light scattering and cell concentration for different phytoplankton groups were made. Calibrations, both laboratory and theoretically derived, are applied to the field data in order to account for the diel variations in beam attenuation. Chapter 4 continues with a similar analysis for diel sampling experiments in the Sargasso Sea during three different months. In Chapter 5, I estimate *in situ* phytoplankton group-specific growth and loss rates in the equatorial Pacific and the Sargasso Sea, using a combination of field observations and laboratory calibrations. In Chapter 6, I present general conclusions of the thesis. In Appendix A, I describe an experiment to calibrate flow cytometric forward light scatter and cell volume over a range of cell sizes. Appendix B contains ancillary data from the field diel sampling experiments in Chapter 5.

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Chapter 2

Diel patterns in optical properties of the chlorophyte

***Nannochloris* sp. grown on light:dark cycles**

ABSTRACT

To investigate how cell growth and division affect the optical properties of phytoplankton, cultures of the chlorophyte *Nannochloris* sp. were sampled over a diel cycle to measure cell size and concentration, light beam attenuation and absorption, flow cytometric light scattering (in forward and side directions), and carbon content. In addition, the refractive index was calculated using the anomalous diffraction approximation of Mie theory. At six different light levels ranging from 60 - 1500 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ (specific growth rates from ~ 0.2 to $\sim 0.7 \text{ d}^{-1}$), cell division was tightly phased to the light:dark cycle, occurring soon after dark. There were pronounced diel patterns, with a minimum near dawn and a maximum near dusk, in cell size and in cell-specific attenuation, absorption, flow cytometric light scatter and carbon. The diel variations in the attenuation cross section were primarily influenced by the changes in cell size due to growth and division, while changes in refractive index had only a small effect. Because eukaryotic cells in the size range of *Nannochloris* are major constituents of many phytoplankton communities, the results presented here have important consequences for the interpretation of diel variations in optical properties observed in the ocean.

INTRODUCTION

There is increasing interest in using optical measurements such as ocean color and beam attenuation to provide convenient and efficient estimates of ocean biomass and primary productivity on large scales. The use of ocean color to estimate primary production, with bio-optical algorithms to interpret the remote measurements in terms of biogeochemical processes, is based on the fact that the color of the ocean is determined by the optical properties of the suspended particles, particularly the phytoplankton and their products (Lewis and Cullen 1991). Thus, an understanding of the variability in optical properties among the particles in the ocean can lead to a better interpretation of these bulk measurements, and thus better estimates of larger scale production.

Since phytoplankton growth is dependent on light, one of the most important scales of variation is over the daily cycle of light and dark. Many researchers have observed diel variations in bio-optical properties in the ocean (Dickey et al. 1990, Hamilton et al. 1990, Stramska and Dickey 1992), and diel variations in beam attenuation have been used to estimate primary production by utilizing a carbon-specific beam attenuation derived from the literature (Siegel et al. 1989, Cullen et al. 1992). In order to fully interpret such bulk measurements made at sea, the diel variations in optical properties of phytoplankton cells must be better understood. Stramski and Reynolds (1993) investigated the diel variations in optical properties of the diatom *Thalassiosira pseudonana* and found that variations in refractive index

were equal to or more important than variations in cell size in affecting changes in optical cross sections. The measured carbon-specific beam attenuation varied over the diel cycle (up to almost 30%). A recent study of *Synechococcus* indicated that, for a slowly growing culture, optical cross sections were minimum near dawn or mid-morning and maximum near dusk (Stramski et al. 1995). The carbon-specific beam attenuation showed substantial diel variability, changing by up to two-fold over the cycle and was different for *Synechococcus* ($2.48 \text{ m}^2 (\text{g C})^{-1}$) than for *T. pseudonana* ($3.81 \text{ m}^2 (\text{g C})^{-1}$). Stramski et al. (1995) suggested that there may be considerable variability in the carbon-specific beam attenuation which should be considered when estimating productivity from diel variations in beam attenuation.

In the experiments reported here on diel variations in phytoplankton optical properties, we chose the chlorophyte *Nannochloris* because it is of a size and shape similar to open ocean ultraphytoplankton (coccolid, 2-3 μm diameter) and it has division patterns typical of eukaryotic phytoplankton (non-diatoms): cell division occurs during the night (Chisholm 1981). We measured cell concentration, attenuation and absorption, cell volume, and carbon, and calculated optical cross sections and refractive index. We also measured forward and side light scatter of these cells using flow cytometry, a method which is used in the field to distinguish phytoplankton populations on the basis of their optical properties. Thus the results of this study can be readily applied to flow cytometric measurements of phytoplankton populations in the field (DuRand and Olson in press, Chapter 4).

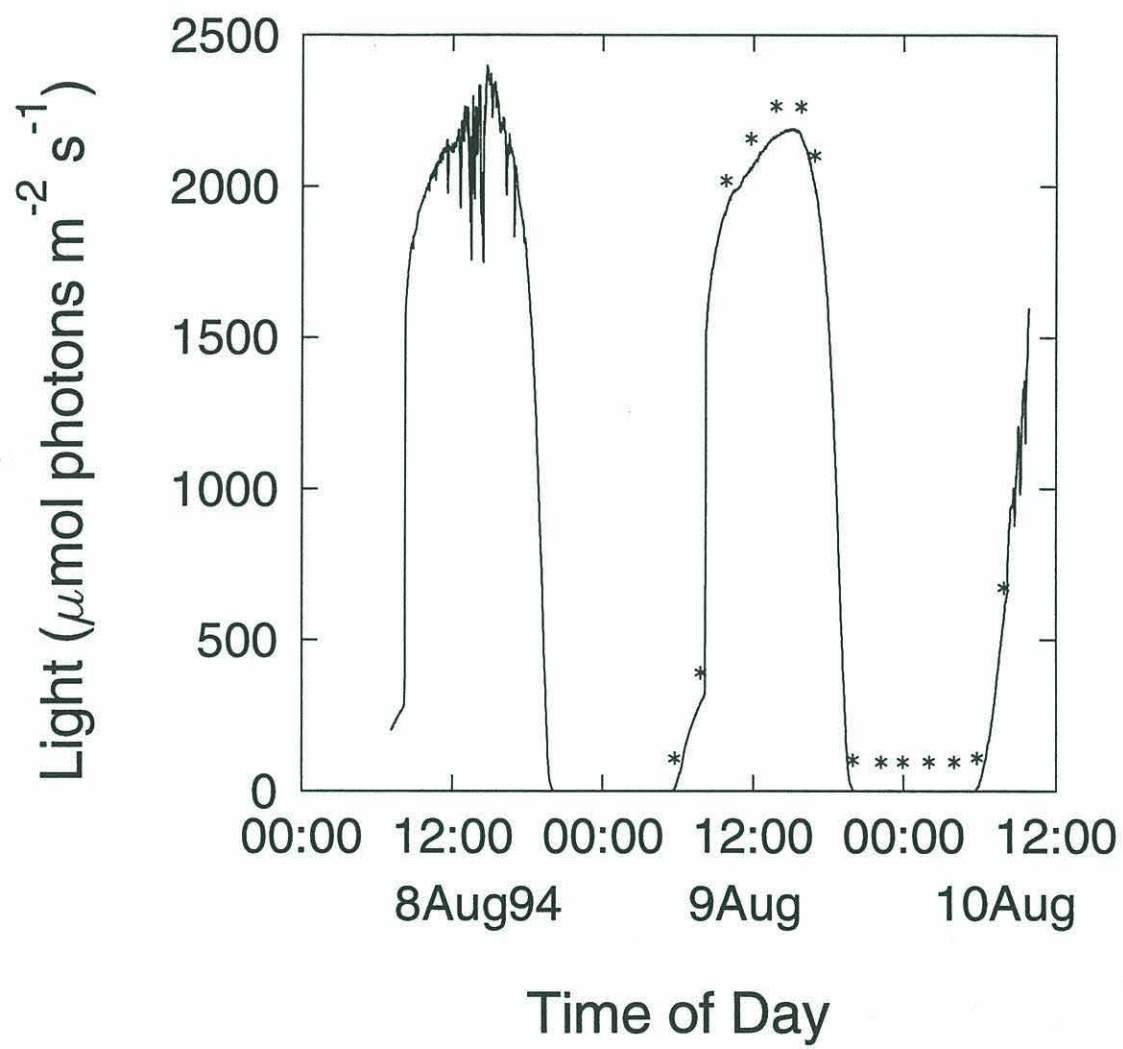
MATERIALS AND METHODS

Batch cultures - Three experiments were performed in which the chlorophyte *Nannochloris* sp. (2.5-3 μm diameter) was grown in f/2 medium in replicate cultures at 22° C (Table 1). The f/2 medium was autoclaved, then sterile filtered through a 0.22 μm Millipore filter (142 mm diameter). For two of the experiments the cultures were grown in 10-l carboys in an incubator on a 14:10 L:D cycle at 250 (Nan250) and 500 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ (Nan500) (Cool-White fluorescent lights, measured with a Biospherical Instruments Inc. QSL-100 4π sensor). The carboys were mixed with a magnetic stir bar and also bubbled with moisturized, filtered air. Samples were obtained through a sterile sampling port (forced out by air pressure). A third experiment (Nan1500) was performed outdoors under natural sunlight with an average integrated light level of about 1500 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ and a maximum of 2200 (14 h daylight on a mostly sunny day in August, Fig. 1). The light level was measured with a Biospherical Instruments Inc. QSR-240 4π sensor in the air. The cultures were in the shadow of the building from dawn (~0600 h) until ~0800 h, after which they were in full sunlight. The cells were grown in Fernbach flasks set in a clear plexiglass water bath with the temperature controlled by a recirculating water bath. The flasks were stirred throughout the experiment and samples were taken by transporting the flasks to a laminar flow hood and then pouring out an aliquot. The cultures were first grown in an incubator at 500 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ and then outdoors for one day before sampling began.

Table 1. Summary of laboratory experiments on *Nannochloris*.

Experiment	Light level	Type of experiment	Measurements
Nan250 July 92	250 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$	Replicate carboys in incubator	Cell size Concentration Attenuation Absorption Carbon Forward light scatter
Nan500 Aug. 92	500 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$	Replicate carboys in incubator	Cell size Concentration Attenuation Absorption Carbon Forward light scatter Side light scatter
Nan1500 Aug. 94	1500 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$	Replicate flasks in natural light	Cell size Concentration Attenuation Absorption Carbon Forward light scatter Side light scatter
Nan60 May 93	60 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$	Cyclostat in incubator	Cell size Concentration Forward light scatter
Nan120 May 93	120 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$	Cyclostat in incubator	Cell size Concentration Forward light scatter
Nan330 May 93	330 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ (up from 120 previous day)	Cyclostat in incubator	Cell size Concentration Forward light scatter

Figure 1. Light intensity ($\mu\text{mol photons m}^{-2} \text{s}^{-1}$) measured outdoors near the experimental setup for Nan1500 for the days the cultures were in natural light. Sampling times (9 Aug - 10 Aug) are marked with asterisks.



For each of the three experiments, beginning at dawn, the exponential phase cultures were sampled every two hours for 24 hours and a number of measurements were made (described below). All measurements were completed within two hours, except where noted. During the nighttime samplings, all samples were kept in dim light during analysis.

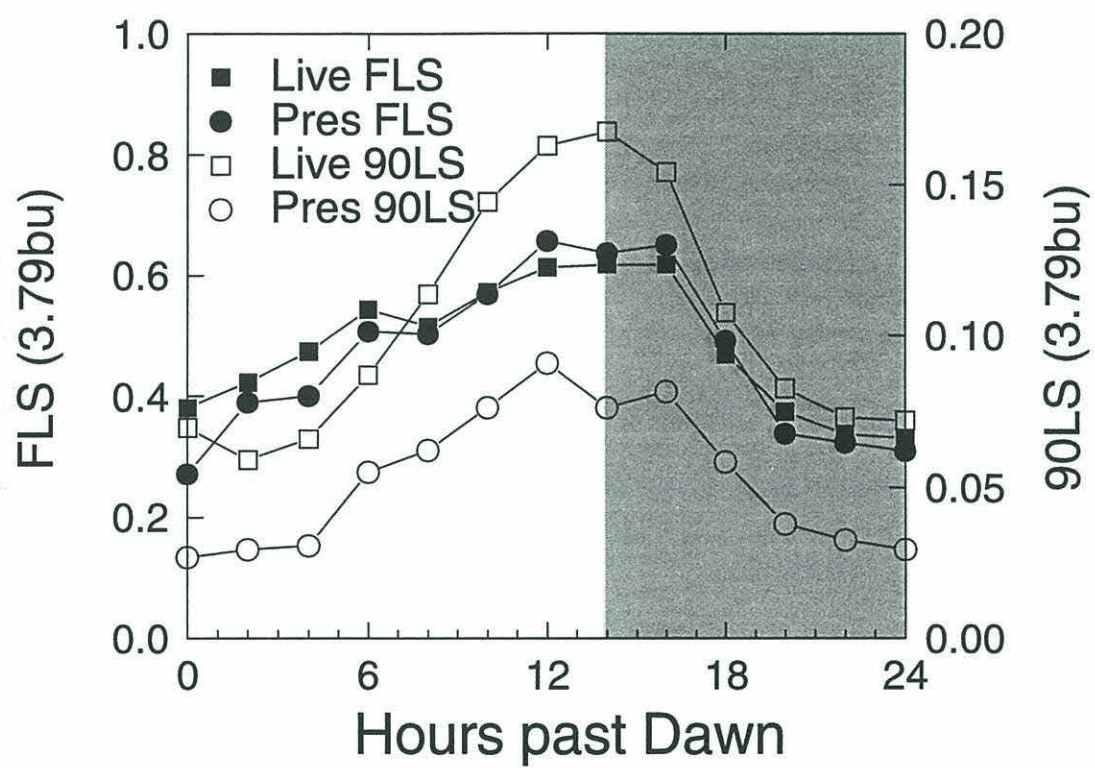
Cyclostat cultures - An additional set of experiments was performed using two cyclostats (nutrient-replete continuous cultures on a L:D cycle), in which samples were collected automatically every hour with a fraction collector (Table 1). The samples were refrigerated at 4° C for up to twelve hours and then analyzed on a Coulter Multisizer; 2-ml aliquots were preserved and frozen for later analysis on the flow cytometer (see below for description of analysis). Light levels for the cyclostats were 60 and 120 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ for the first 24-h sampling, then a second 24-h sampling was started after increasing the light level to 330 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ for the cyclostat that previously had been at 120 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$.

Flow cytometry - Triplicate samples were analyzed on an EPICS 753 flow cytometer modified to analyze 50 ml seawater samples at 5-10 ml min^{-1} (Olson et al. 1991, 1993). The culture was diluted 100 μl : 50 ml with 0.22 μm filtered seawater (FSW). Fluorescent microspheres (3.79 μm , Polysciences, Inc.) were added to each sample as an internal standard. The concentration of beads was calibrated on the flow cytometer in order to calculate cell concentrations (Olson et al. 1993). For each particle, forward light scatter (FLS, 3°-19°; see Appendix A), side scatter (90LS, 73°-107°), red (chlorophyll, 660-700 nm) fluorescence, and orange (phycoerythrin, 530-

630) fluorescence were collected. The resulting "listmode" files were analyzed using CytoPC software (D. Vaultot, Station Biologique, Roscoff, France) to obtain numbers of cells and beads analyzed and the mean values for each parameter collected. For Nan500 and Nan1500 the samples were analyzed live, for Nan250 and the cyclostat experiments (Nan60, Nan120, and Nan330) the samples were fixed with 0.1% glutaraldehyde and stored in liquid nitrogen until analysis (Vaultot et al. 1989, Olson et al. 1993). A comparison of live and frozen samples for Nan500 showed very little difference in forward light scatter, though side light scatter did change significantly and thus was not used for preserved samples (Fig. 2).

Beam attenuation - Beam attenuation was measured with a SeaTech transmissometer (665 nm light source, 25-cm pathlength) enclosed by a PVC sleeve with a sampling port. First, 1.6 liters FSW was added to the sleeve and a voltage reading was made. Then, culture was added, progressively, to total 8, 12, and 16 ml and a reading was made after each addition. For each measurement, the culture was added to the FSW, the mixture was stirred, and any bubbles were cleared from the transmissometer windows before taking the reading. The procedure was repeated twice for each culture. To analyze the data, voltages were converted to beam attenuation (c, m^{-1}), the preceding FSW blank reading was subtracted from each culture measurement, and the value was normalized to the concentration of cells in the transmissometer sleeve (the measured volume of the culture addition and the calculated concentration from flow cytometric measurements). The samples in the transmissometer sleeve were determined to be optically thin (since the sequential

Figure 2. Flow cytometric light scatter for *Nannochloris* grown at 500 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ (Nan500) analyzed live and after being preserved with glutaraldehyde and stored in liquid nitrogen. Forward light scatter (FLS) changed little, but side light scatter (90LS) showed significant changes and was thus the 90LS data were not used for preserved samples. Both FLS and 90LS are expressed relative to 3.79 μm beads.



addition series did not depart from linearity), thus multiple interactions were negligible (Bricaud et al. 1983).

Volume - Cell volume was measured using a Coulter Multisizer equipped with a 30 or 50 μm orifice. Samples were diluted in filtered seawater to keep the coincidence rate $< 5\%$. At each sampling point, duplicate or triplicate samples were mixed and measured. The samples were analyzed immediately after sampling, except for the cyclostat experiments (Nan60, Nan120, Nan330) when they were refrigerated for up to 12 hours before analysis. There was little difference between replicate test samples run on the Multisizer immediately after collection or after eleven hours of refrigeration (diameter was identical and cell concentration was only 1% different). For all samples the 256-channel data was saved and analyzed on a computer using software by M. Brzezinski.

Carbon - Replicate samples (25 ml) were filtered onto pre-combusted GF/F filters, frozen, and later dried overnight at 60°C and analyzed on a Perkin-Elmer 2400 CHN Analyzer. Replicate blanks consisting of a wet filter were made at each sampling point. Acetanilide was used as the standard. The blanks were subtracted from the samples to obtain the carbon content of the known volume (and number) of cells.

Absorption - Absorption was measured using the opal glass method (Shibata 1958) in a spectrophotometer. A piece of opal glass was attached as close to the detector as possible, and a 1-cm cuvette was placed directly against the opal glass. For Nan250 and Nan500, a Bausch and Lomb Spectronic spectrophotometer was

used (Model 88 for Nan250, Model 100 for Nan500) and a reading was made at two wavelengths, 488 nm to correspond to the flow cytometer laser light and 665 nm to correspond to the LED light source in the transmissometer. For Nan1500, a Shimadzu UV-260 spectrophotometer was used and each sample was scanned from 400-750 nm. For each experiment, sterile-filtered f/2 medium was used in the reference cuvette and subtracted as the blank.

Calculations of optical properties - The scattering coefficient (b) was calculated from the measured attenuation (c) and absorption (a) coefficients ($b = c - a$). The cross sections for attenuation (σ_c), absorption (σ_a), and scattering (σ_b) are the relevant coefficients normalized to the cell concentration (for example, $\sigma_c = c/(N/V)$, where N/V is the number of cells per volume). Cross sections are in units of length² per cell. The dimensionless efficiency factor for attenuation (Q_c , and similarly for absorption and scattering) was calculated: $Q_c = \sigma_c/(\pi r^2)$. Using the anomalous diffraction approximation of Mie theory (Van de Hulst 1957, as described in Morel and Bricaud 1986), the imaginary part of the refractive index n' (where $n' = \rho'/4[2\pi r/(\lambda n_w)]$, ρ' is the optical thickness due to absorption, λ = wavelength of light, and $n_w = 1.34$, the refractive index of seawater) was then calculated through iterations from

$$Q_a(\rho') = 1 + 2 \frac{\exp(-\rho')}{\rho'} + 2 \frac{\exp(-\rho') - 1}{\rho'^2}.$$

Once n' (and thus ρ') was determined, the real part of the refractive index was

calculated through iterations from

$$Q_c(\rho) = 2 - 4\exp(-\rho \tan \xi) \left[\frac{\cos \xi}{\rho} \sin(\rho - \xi) + \left(\frac{\cos \xi}{\rho} \right)^2 \cos(\rho - 2\xi) \right] + 4 \left(\frac{\cos \xi}{\rho} \right)^2 \cos 2\xi$$

where $\tan \xi = (1/2) \rho'/\rho$ and ρ is the phase lag of a ray which crosses the particle along its diameter (Morel and Bricaud 1986). Since there are multiple solutions for Q_c , the results were restricted to those reasonable for phytoplankton cells (the first solution with $n > 1.02$ was accepted; all calculated n values were between 1.04 and 1.07).

RESULTS AND DISCUSSION

Measurements of cell properties

For Nan250, Nan500, and Nan1500, cell division was tightly phased, with division starting two hours after darkness and continuing for about six hours (Fig. 3A). Nan1500 was performed outdoors in natural light, unlike the other experiments, thus the cells were exposed to a true dawn and dusk (Fig. 1). The timing of the cell division for this experiment was very similar to that of the others. The cells in the cyclostat experiments (Nan60, Nan120, and Nan330) also divided during the night (Fig. 3B). Specific growth rates determined from flow cytometric measurements of cell concentrations ranged from ~ 0.2 to $\sim 0.7 \text{ d}^{-1}$ (approximately one division per day; Fig. 4).

Cell volume of *Nannochloris* was minimum near both dawn samplings and maximum near dusk for each experiment (Fig. 5). Flow cytometric measurements of individual cell light scatter (forward, FLS, and side, 90LS) showed similar patterns (Fig. 6). Carbon per cell also increased during the day and decreased at night (Fig. 7). Thus the individual cells are photosynthesizing to add carbon and grow in size during the day and then dividing in the dark to produce smaller cells with less carbon per cell and lower scatter per cell.

The calculated values of refractive index (n) ranged from 1.04 to 1.066 (Fig. 8A), within the range expected for phytoplankton cells (1.02 - 1.07; Aas 1981 as referred to by Morel and Bricaud 1986). There appears to be a small diel pattern in refractive index. The slope of a linear regression fitted to the values of refractive index over time was significantly less than zero for all the experiments during the night, when cell division was occurring (t-test, $p < 0.05$ for each of the three experiments). The refractive index increased significantly during the day for Nan500 (t-test, $p < 0.0025$). The increase was not significant for Nan250 and Nan1500, perhaps because most of the increase occurred between the first two points for Nan1500 and because the first point for Nan250 was lost.

Refractive index most likely reflects the changing composition of the cells as they grow and divide, due to the different refractive indices of internal components and the cell water content (Aas 1981, as referred to by Morel and Bricaud 1986). The intracellular carbon concentration shows a pattern similar to that of refractive index (Fig. 8B). Both n and intracellular carbon concentration clearly decrease at

Figure 3. Cell concentrations changes, measured by flow cytometry, over the course of the experiments for *Nannochloris* at A) 250 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ (Nan250), 500 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ (Nan500), and 1500 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ (Nan1500) and B) the cyclostat experiments at 60 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ (Nan60), 120 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ (Nan120), and 330 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ (Nan330). Shading indicates the time period when the lights were out in the incubator (or nighttime for Nan1500). In A) each point represents the average value for the two carboys and error bars show standard errors. In B) the potential cell concentration is shown (measured cell concentration with the dilution rate of the cyclostat taken into account: potential cells $\text{ml}^{-1} = \text{measured cells ml}^{-1} + \text{hourly dilution rate} * \text{number of hours elapsed} * \text{measured cells ml}^{-1}$). Note that Nan120 and Nan330 were the same culture, with only the light level changed for the second day.

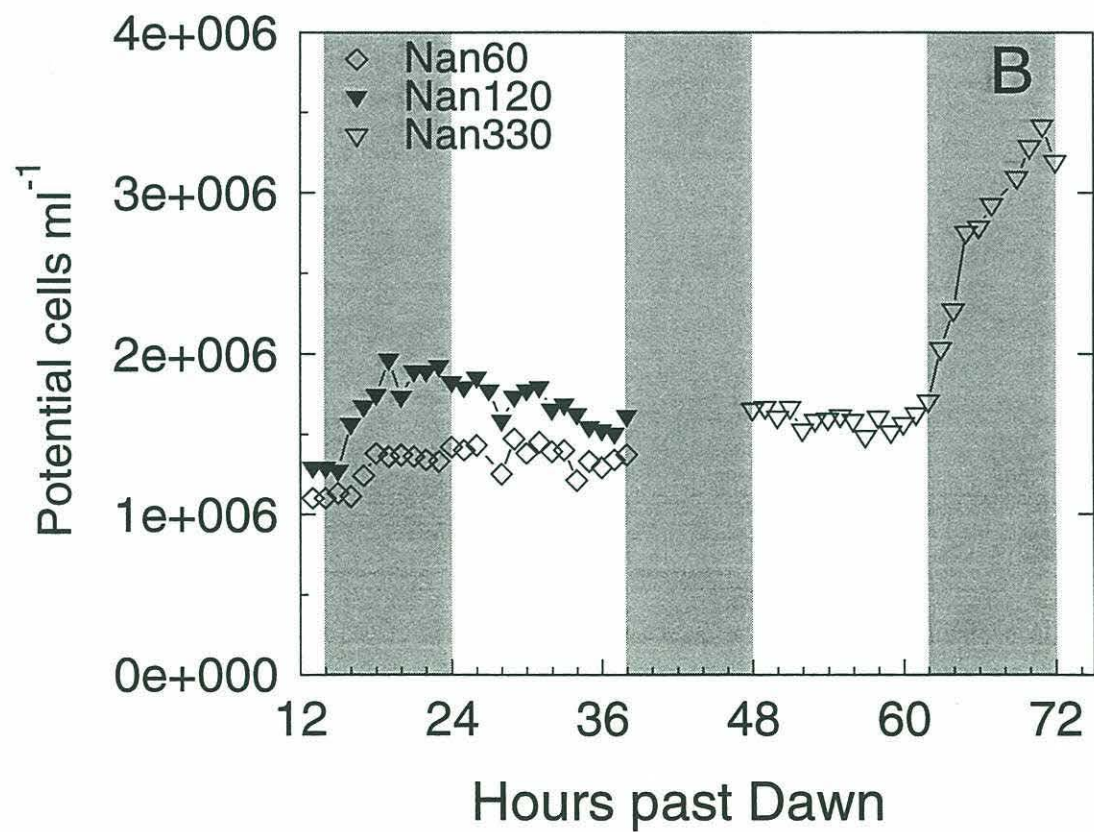
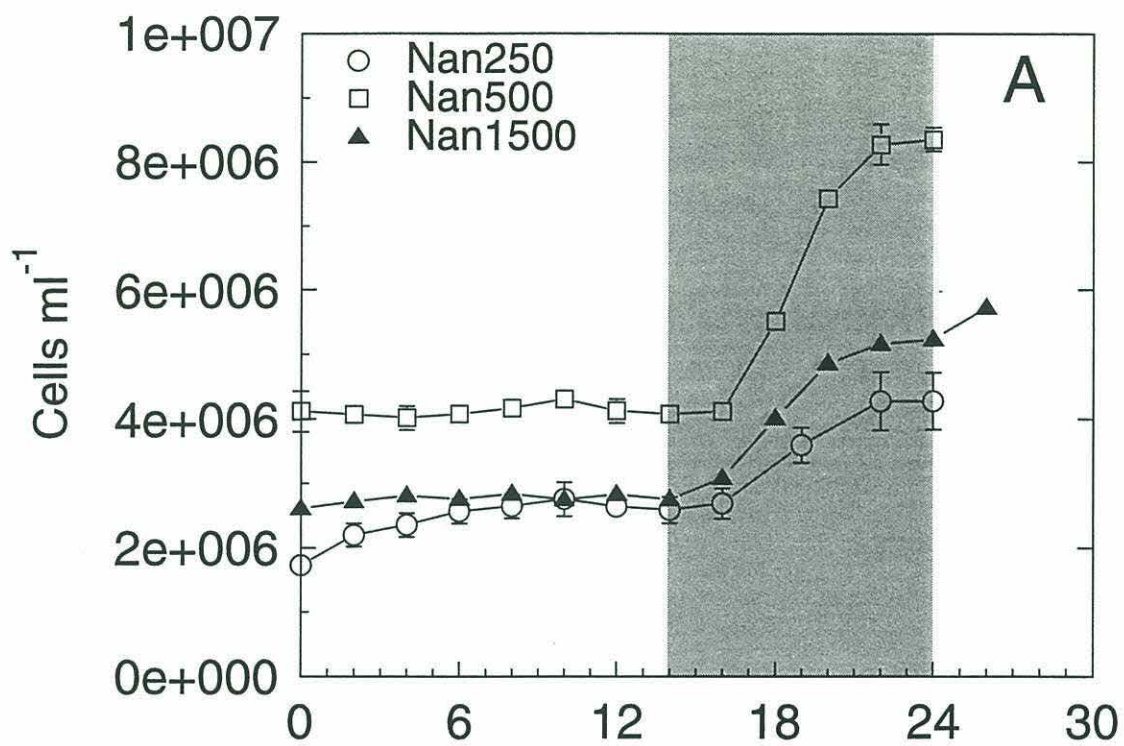


Figure 4. Specific growth rate (d^{-1}) determined from changes in cell concentration (see Fig. 3) at different light levels. For each experiment the cells were exposed to the light level indicated for fourteen hours out of every 24 hours.

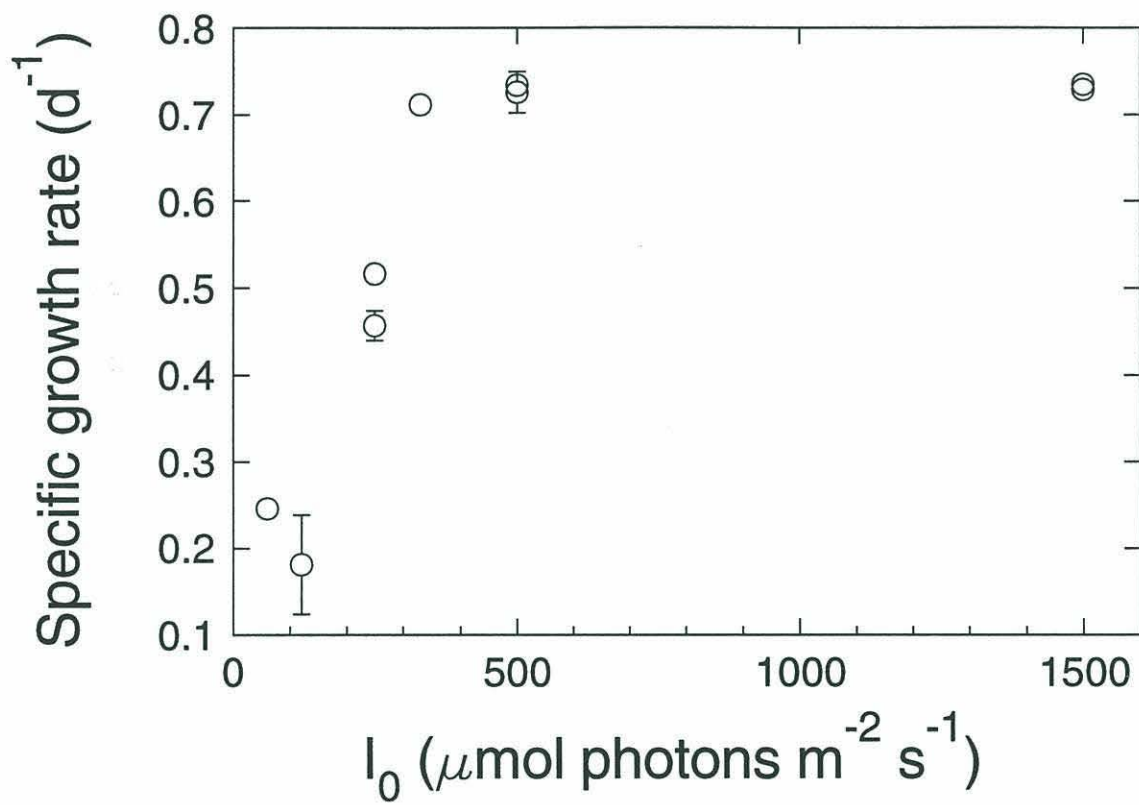
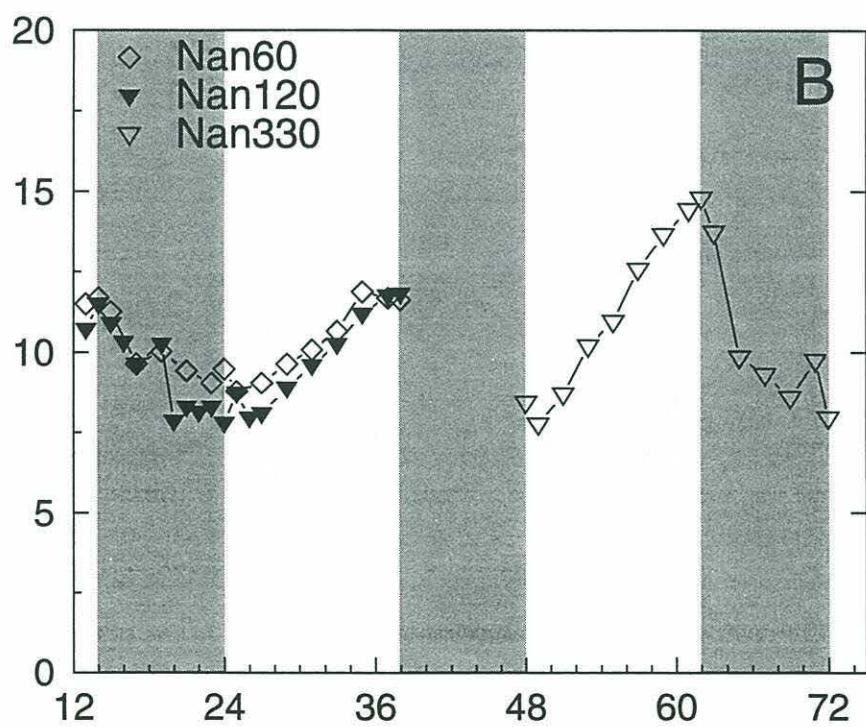
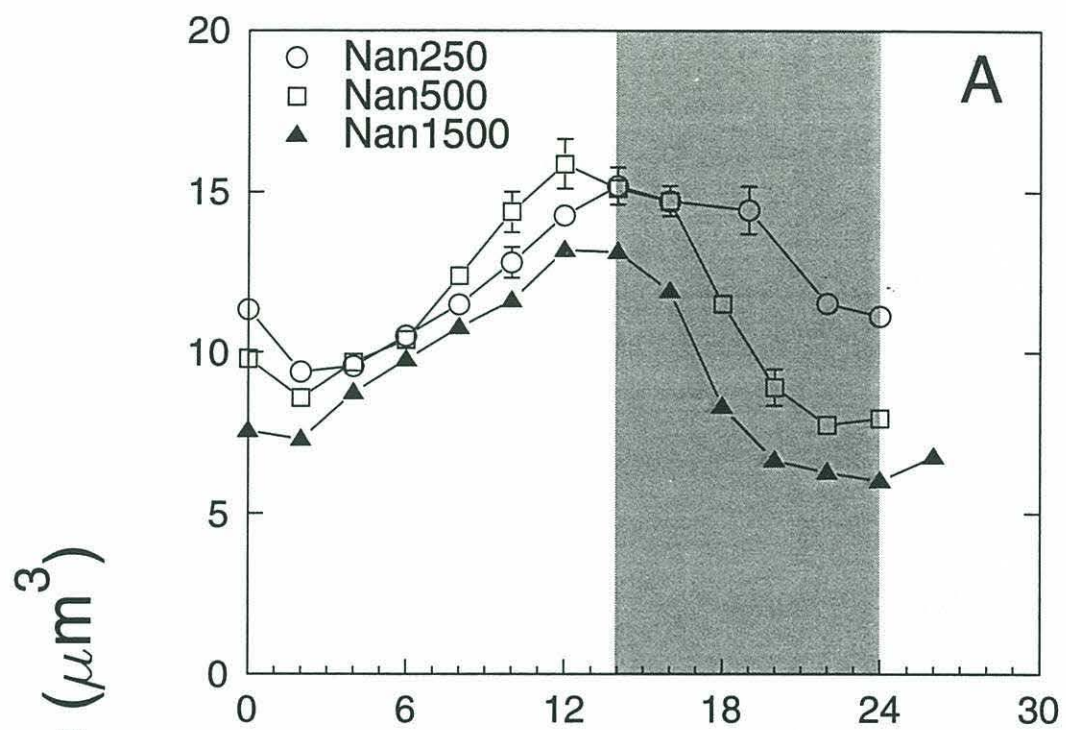


Figure 5. Time series measurements of cell volume from Coulter electronic particle sizer for A) Nan250, Nan500, and Nan1500 and B) the cyclostat experiments Nan60, Nan120, and Nan330. For A) average ($n = 2$) and standard error bars are shown.



Hours past Dawn

Figure 6. Time series measurements of forward light scatter (3° - 19°) for A) Nan250, Nan500, and Nan1500, and B) Nan60, Nan120, and Nan330. C) Side scatter (73° - 107°) from flow cytometric measurements for the same experiments as in panel A. Values are expressed relative to $3.79\ \mu\text{m}$ beads.

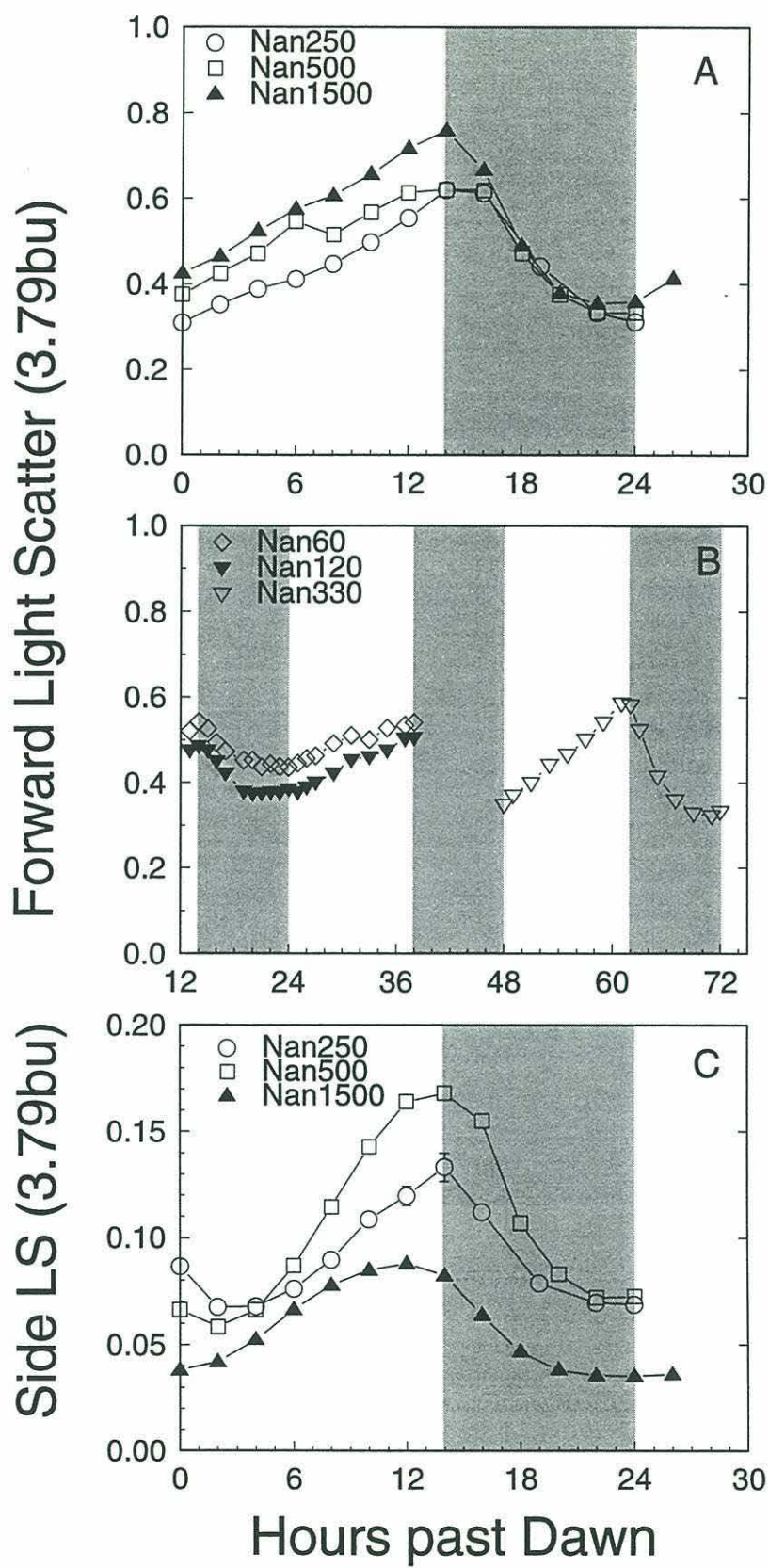


Figure 7. Time series measurements of fg carbon per cell for Nan250, Nan500, and Nan1500. Averages ($n = 2$ carboys) and standard error bars are shown.

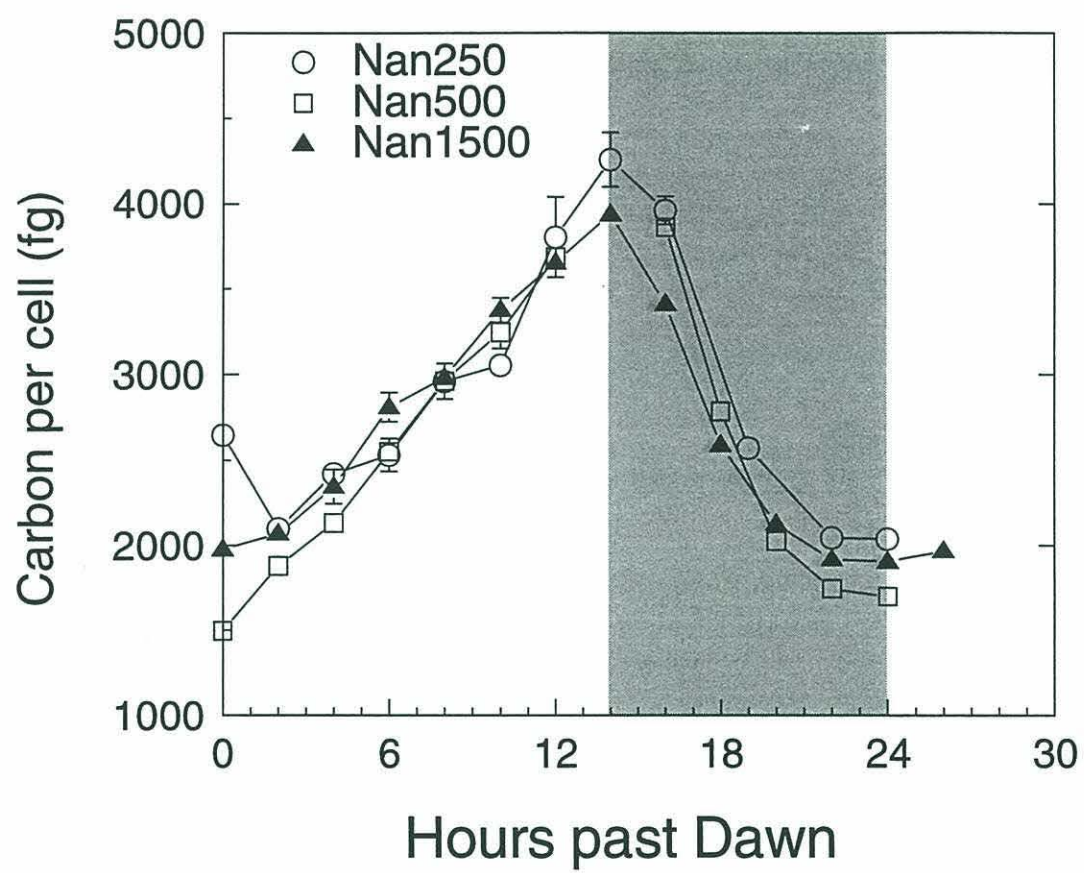
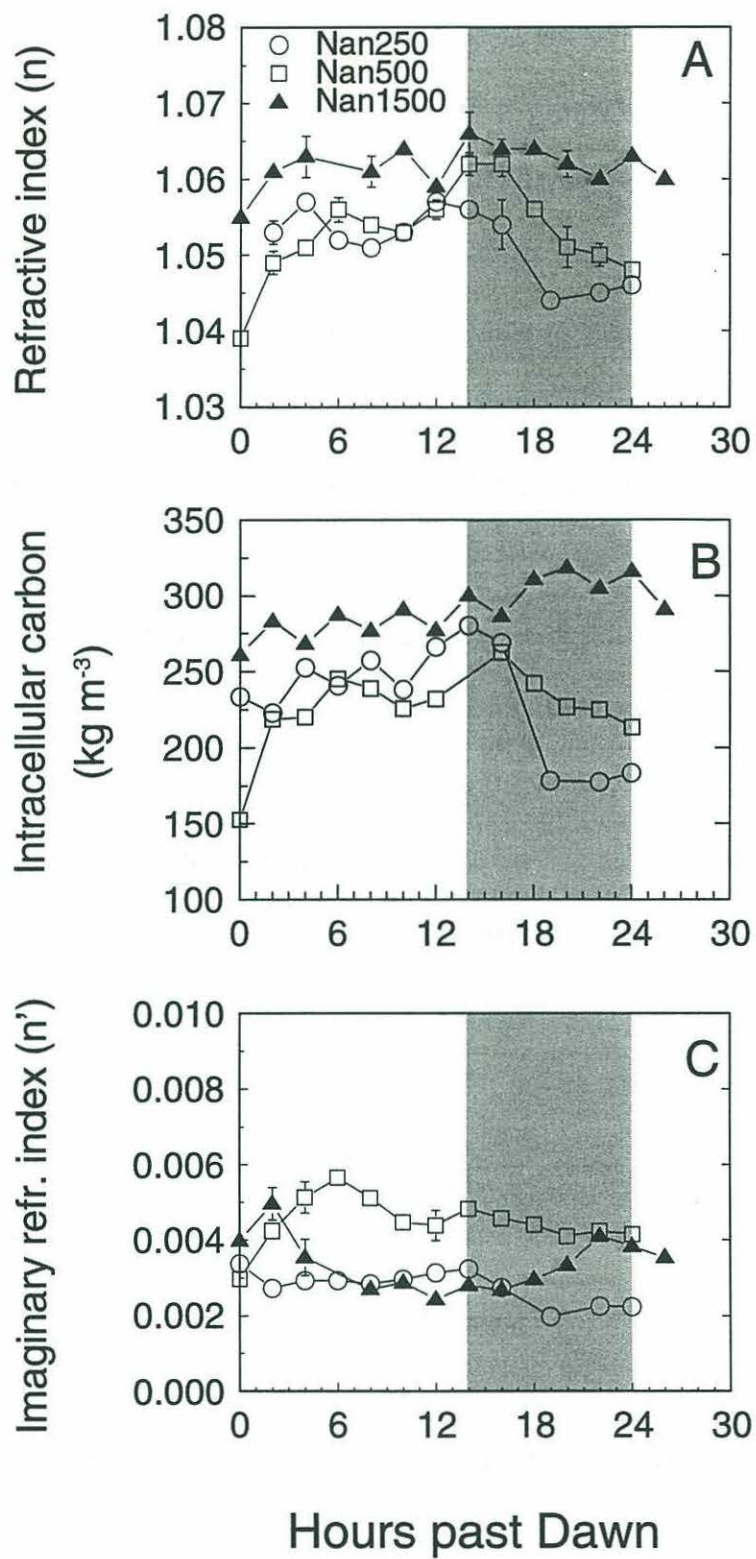


Figure 8. Time series of A) calculated real refractive index (n), B) intracellular carbon concentration (kg m^{-3}), and C) calculated imaginary refractive index (n') of *Nannochloris* for Nan250, Nan500, and Nan1500. All calculations were made at a wavelength of 665 nm.



night when the cells are dividing for Nan250 and Nan500. One explanation is that the cell water content could be higher (causing a lower refractive index) when the cells are dividing. Another possibility is that the decrease in refractive index reflects a loss of carbon due to cell respiration when the cells are dividing. Markager et al. (1992) observed a faster rate of dark respiration for the first two hours in the dark for a community of phytoplankton in a eutrophic lake. In studies of *Isochrysis galbana*, Carder et al. (1972) suggested that increases in refractive index seemed to be related to increases in the surface area to volume ratio of the cells, since the cell wall was expected to be one of the most refractive parts. However, if this were the case, we would expect an increase in n with cell division, instead of the observed decrease, since the newly divided (smaller) cells would have a higher surface area to volume ratio. It is interesting to note that the highest n (close to 1.06) and intracellular carbon concentration and the smallest changes in both properties during the diel cycle are for the natural light experiment (Nan1500). This could be due to the higher light level or the fact that there was a true dawn and dusk for that experiment. Additional experiments are required to further investigate these effects.

The calculated value of the imaginary part of the refractive index, generally ranging from 0.003 to 0.005 (Fig. 8C), was similar to that determined by others for phytoplankton cells ($n' \approx 0.004$ at 660 nm; Stramski and Reynolds 1993). For all three experiments, the changes seen are small. There was little pattern over the diel cycle for Nan250; Nan500 increased during the morning and then decreased during the afternoon, while Nan1500 tended to decrease during the day and increase at

night. Since the cultures for Nan1500 were only exposed to the higher natural light conditions for one day before the sampling began, they were not expected to be at steady state. Thus we are most likely seeing effects of photoacclimation during that experiment.

The attenuation cross section (σ_c) and the scattering cross section (σ_b) both increased during the day to reach a maximum at dusk and then decreased at night (Fig. 9A, B). These patterns are similar to those seen in cell volume (Fig. 5), light scatter per cell (Fig. 6) and carbon per cell (Fig. 7). The absorption cross section (σ_a) was always small compared to the attenuation cross section (Fig. 9C); it tended to increase during the day and decrease at night. The chlorophyll fluorescence per cell measured with the flow cytometer showed similar patterns to σ_a , with higher fluorescence for Nan60 and Nan120, the lowest light levels, compared to Nan250, Nan500, and Nan1500 (Fig. 10). The second day of the cyclostat experiment (when the Nan120 culture was exposed to high light to become Nan330) shows unexpected results, reaching the highest fluorescence value of all the experiments at its maximum.

Refractive index and intracellular carbon concentration are strongly correlated ($n = 36$, $r^2 = 0.85$; Fig. 11). The slope of the relationship (approximately $1.5e-4$) indicates that the refractive index of the *Nannochloris* cells increases by $1.5e-4$ for every one kg m^{-3} increase in the intracellular carbon concentration. This value is lower than that calculated by Morel and Ahn (1990) based on the contributions of the internal components of a cell ($2.3e-4$). It is also lower than that measured by

Figure 9. Time series of the attenuation cross section (σ_c , A), scattering cross section (σ_b , B), and absorption cross section (σ_a , C) at a wavelength of 665 nm for Nan250, Nan500, and Nan1500. The attenuation and absorption cross sections are from bulk measurements of attenuation and absorption normalized to the cell concentration. The scattering cross sections were obtained by subtraction ($\sigma_c - \sigma_a$).

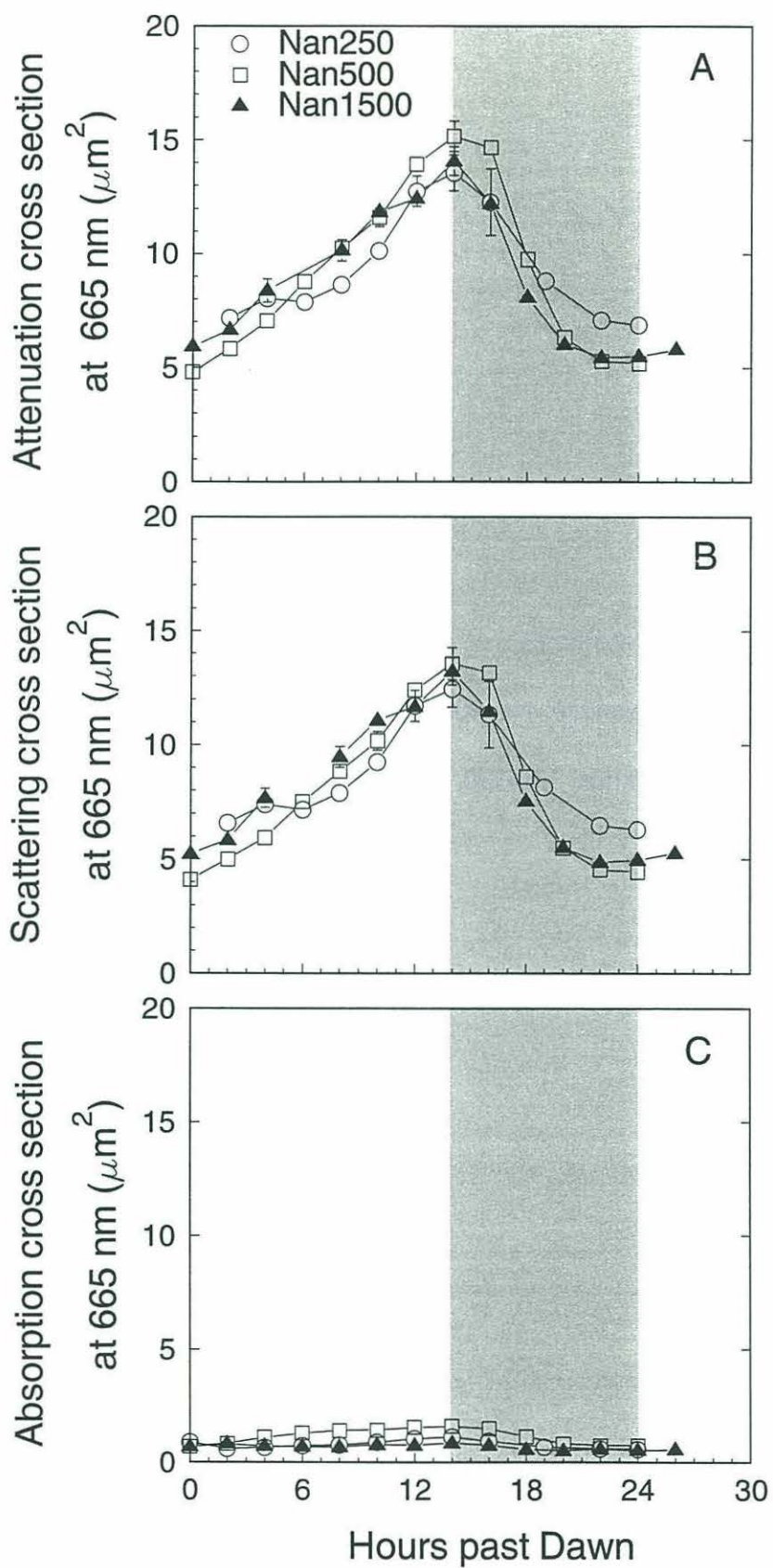


Figure 10. Time series measurements of chlorophyll fluorescence per cell from flow cytometric analysis for A) Nan250, Nan500, and Nan1500 and B) the cyclostat experiments Nan60, Nan120, and Nan330. Values are expressed relative to 3.79 μm beads.

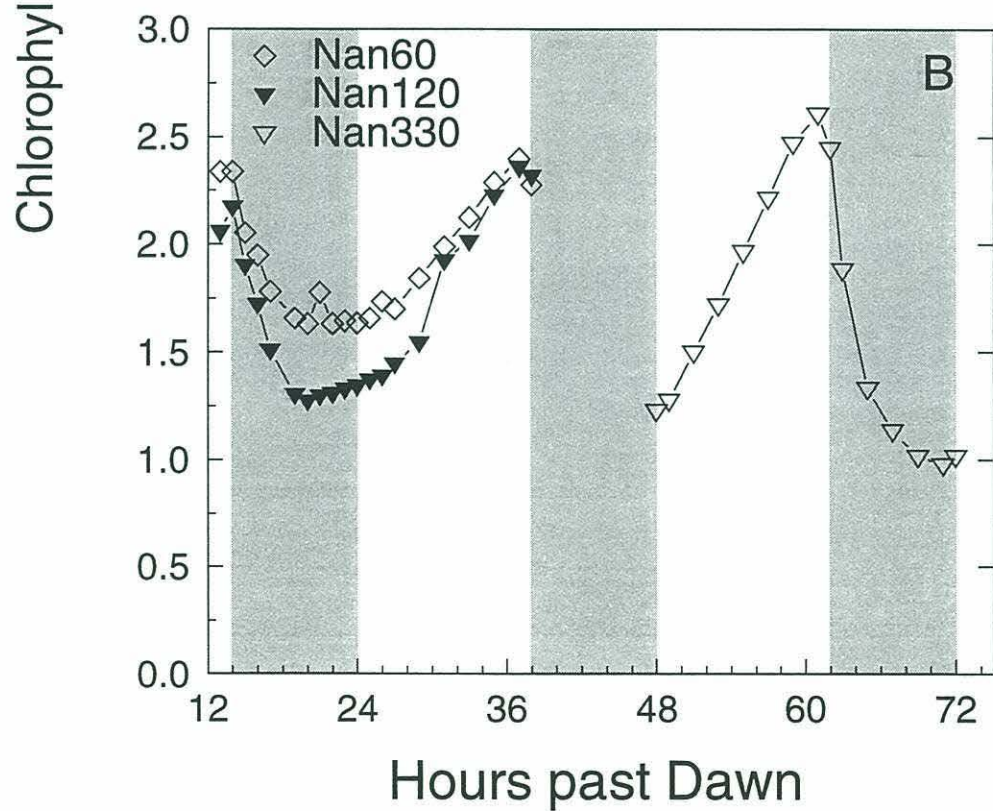
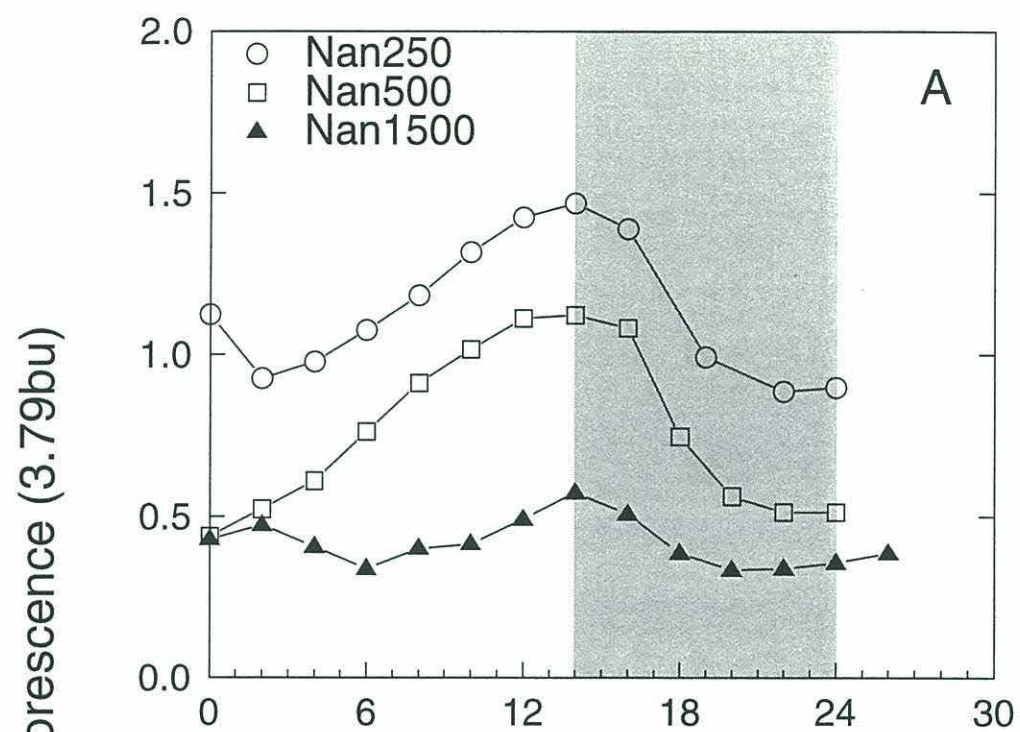
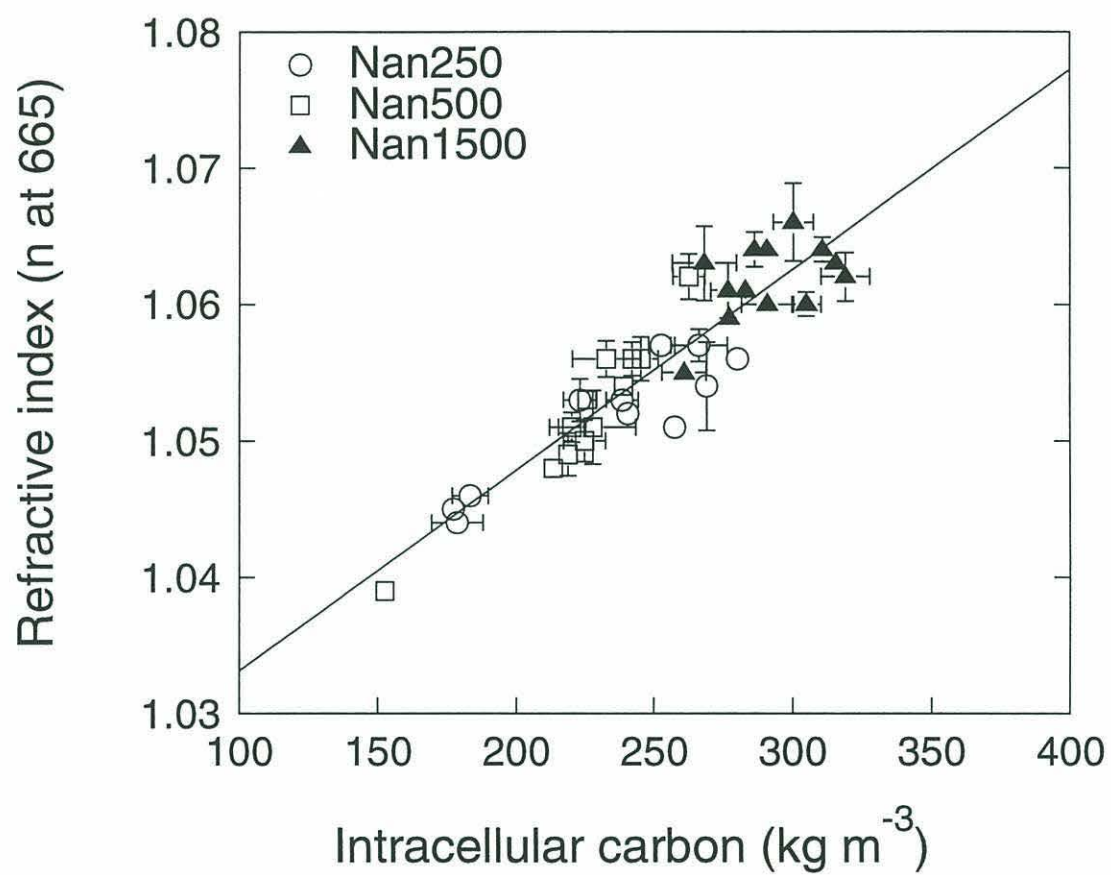


Figure 11. Relationship between refractive index (n at 665 nm) and intracellular carbon concentration (kg m^{-3}) for Nan250, Nan500, and Nan1500. The equation for the regression is $y = 1.47\text{e-}4 (x) + 1.02$, $r^2 = 0.85$.



Stramski and Reynolds (1993) for *Thalassiosira pseudonana* (1.7×10^{-4}), Stramski et al. (1995) for *Synechococcus* (1.7×10^{-4}), and Stramski and Morel (1990) for *Synechocystis* (2.4×10^{-4}). Thus, compared to previous cells tested, *Nannochloris* appears to have a smaller increase in refractive index for the same change in intracellular carbon concentration.

There is a strong correlation between the flow cytometric forward light scatter (at 488 nm) and the total scattering cross section (σ_b , at 665 nm) determined by subtraction of the measured σ_c and σ_a ($r^2 = 0.80$; Fig. 12). This is important since flow cytometric FLS can be measured on individual cells within mixed populations in natural samples. Flow cytometry provides an important link between optical properties which, though often measured in bulk, can be expressed on a per cell basis for laboratory studies, and measurements in the field which can only be made in bulk, such as beam attenuation (DuRand and Olson in press, Chapter 4).

Estimation of cell volume and specific growth rate

The relationship between cell size and forward light scatter changes depending on cell size. According to Mie theory, for picoplankton-sized cells, forward light scatter should vary with diameter to the sixth power (Morel and Bricaud 1986). For cells in the size range of *Nannochloris*, FLS should vary with diameter to the second or third power (i.e., with area or volume) depending on the assumed refractive index and angles of collection (results of Mie theory calculations performed using program of Ackleson and Spinrad 1988). Since we are interested in

Table 2. Results of linear regressions on volume (vol, in μm^3) and forward light scatter (FLS) or side light scatter (90LS), both expressed relative to 3.79 μm reference beads.

Experiment	Equation	n	r^2
Nan1500	$\text{Vol} = 18.6 * \text{FLS} - 0.70$	27	0.98
Nan1500	$\text{Vol} = 121 * 90\text{LS} + 2.29$	27	0.91
Nan60, 120, 330, 500	$\text{Vol} = 24.1 * \text{FLS} - 0.54$	77	0.80
Nan60, 120, 330, 500	$\text{Vol} = 68.9 * 90\text{LS} + 4.13$	26	0.91

small ranges of size (close to twofold), we can simplify the situation by assuming a linear relationship between size and FLS (given an approximately constant refractive index). For each *Nannochloris* experiment, a linear regression was obtained for the relationship between FLS and cell volume (Fig. 13). An analysis of covariance was performed and the hypothesis that all slopes (β) are equal was rejected ($p < 0.0005$). A multiple comparison testing of the slopes (Tukey test) revealed that they came from two separate populations, where $\beta_{120} = \beta_{60} = \beta_{500} = \beta_{330} \neq \beta_{1500}$ (we cannot conclude how β_{250} is related to the other population slopes). The implication is that Nan1500, the natural light experiment, is different from the others and we will consider it separately.

Relationships between FLS or 90LS and cell volume were determined separately for the two distinguishable groups of data: for Nan1500 and for Nan60, 120, 500, and 330 combined. Cell volume for both data sets was well estimated by either a linear regression on FLS or 90LS (Table 2), despite changes in refractive index seen over the day (Fig. 8A). Ackleson and co-workers used flow cytometric

Figure 12. Relationship between flow cytometric forward light scatter (at 488 nm, 3.79 μm bead units) and total scattering cross section (calculated from measured attenuation and absorption at 665 nm). Linear regression ($r^2 = 0.80$) and 95% confidence intervals are shown.

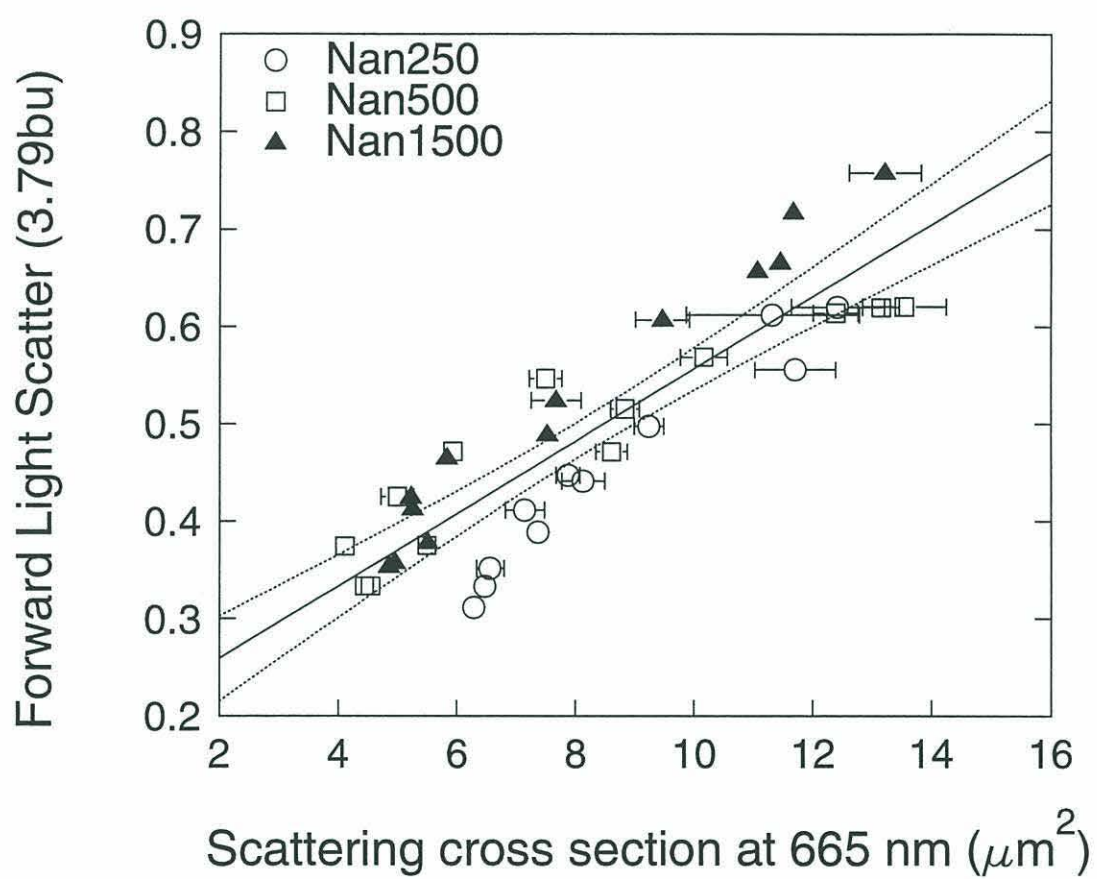
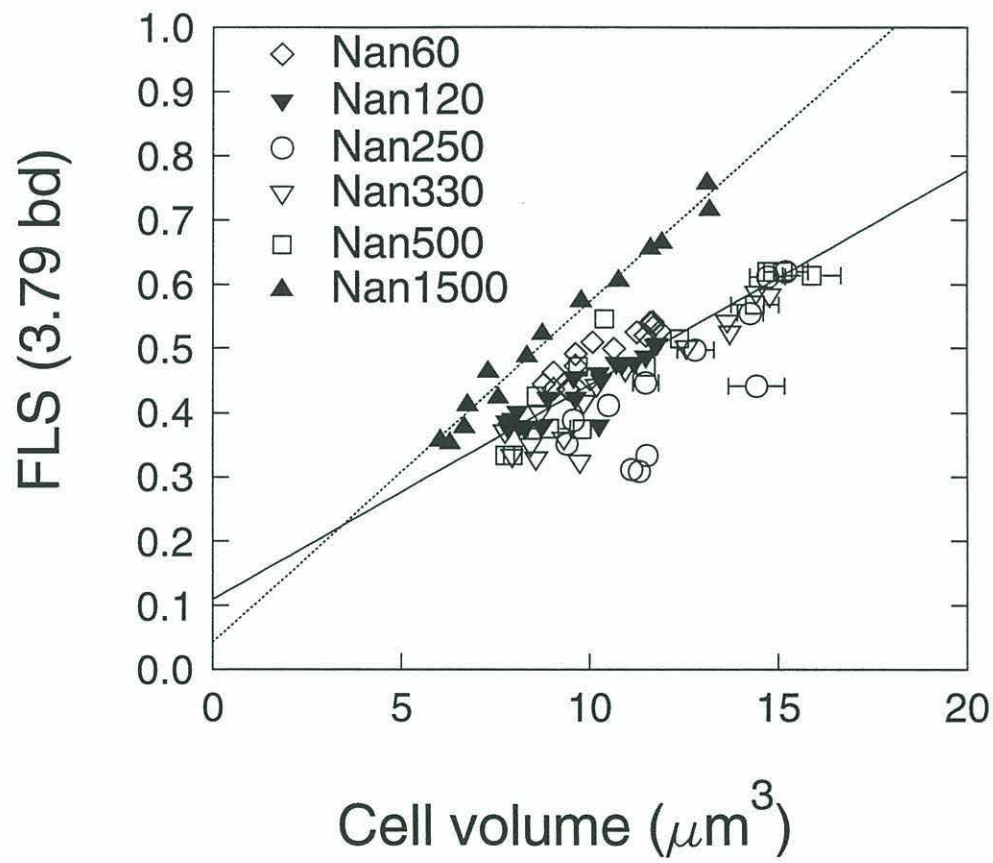


Figure 13. The relationship between forward light scatter (FLS in 3.79 μm bead units) and cell volume (μm^3) for Nan60, Nan120, Nan250, Nan330, Nan500, and Nan1500. The dotted line is a linear regression on the Nan1500 data only. The solid line is a linear regression on the combined data for Nan60, Nan120, Nan330, and Nan500.



measurements of FLS and side scatter to estimate cell volume and refractive index using Mie theory for laboratory cultures (Ackleson and Spinrad 1988) and also in the field (Ackleson et al. 1988). For their laboratory culture work, the range in the estimated refractive index was up to nearly 0.01, which is similar to the range calculated for the *Nannochloris* experiments reported here. Thus it appears that such a method may not be able to distinguish the small changes in refractive index which may occur over the day. This may not be an important drawback, however, if the small refractive index changes do not affect the total scattering or attenuation as much as changes in cell size do.

Specific growth rates can be estimated from the diel measurements in cell volume if cell division is phased. If, for example, the cells are dividing once per day, then the average volume would be expected to double during the day and then halve upon division at night (in general, $\mu = \ln (\text{MaxVolume}/\text{MinVolume})$). If the refractive index is not changing significantly, then cell light scattering should also give a good estimate of the growth rate for cells of this size, for which forward light scatter is proportional to volume (near d^3). For these experiments, the forward light scatter measurements give a better estimate of the growth rate (calculated from cell concentrations) than do cell volume or side scatter (Table 3).

Table 3. Results of linear regressions (with zero intercept) where X = specific growth rate calculated from cell concentration ($\mu = \ln(N_1/N_0)/(t_1-t_0)$; where N is the cell concentration, t is time, and the subscripts indicate values at two times (1 is after division and 0 is before division; the time interval is one day)) and Y = specific growth rate calculated from forward light scatter, side light scatter, or cell volume ($\mu = \ln(\text{Max}/\text{Min})$; with a time interval of one day). $N = 9$, $p < 0.001$ for all regressions except Volume Daytime increase where $p < 0.10$.

Data for Y	Equation	r^2
FLS (Daytime increase)	$y = 0.740x$	0.82
FLS (Nighttime decrease)	$y = 0.934x$	0.88
90LS (Daytime increase)	$y = 1.23x$	0.78
90LS (Nighttime decrease)	$y = 1.186x$	0.72
Volume (Daytime increase)	$y = 0.871x$	0.25
Volume (Nighttime decrease)	$y = 0.947x$	0.82

Carbon-specific beam attenuation

The carbon-specific beam attenuation (c_c^* , Fig. 14) varies by up to 25% over the course of the measurements. C_c^* for both Nan500 and Nan1500 tends to increase during the day and decrease rapidly concurrent with cell division at the beginning of the night; c_c^* for Nan250 shows no distinct pattern. This may be important in relation to estimates of carbon production from diel variations in beam attenuation at sea (Siegel et al. 1989, Cullen et al. 1992, Walsh et al. in press). In these cases, a constant c_c^* was applied to represent all phytoplankton at all times of the day. If, indeed, c_c^* changes during the day then these conclusions may not be valid. The increase from dawn to dusk is up to 25% for *Nannochloris* (reported here), and about 30% for slowly-growing *Synechococcus* (Stramski et al. 1995), but with no consistent pattern from dawn to dusk for *Thalassiosira pseudonana* (though up to 30% variation

Figure 14. Time series of the carbon-specific beam attenuation (c_c^* in $\text{m}^2 (\text{gC})^{-1}$) for Nan250, Nan500, and Nan1500.

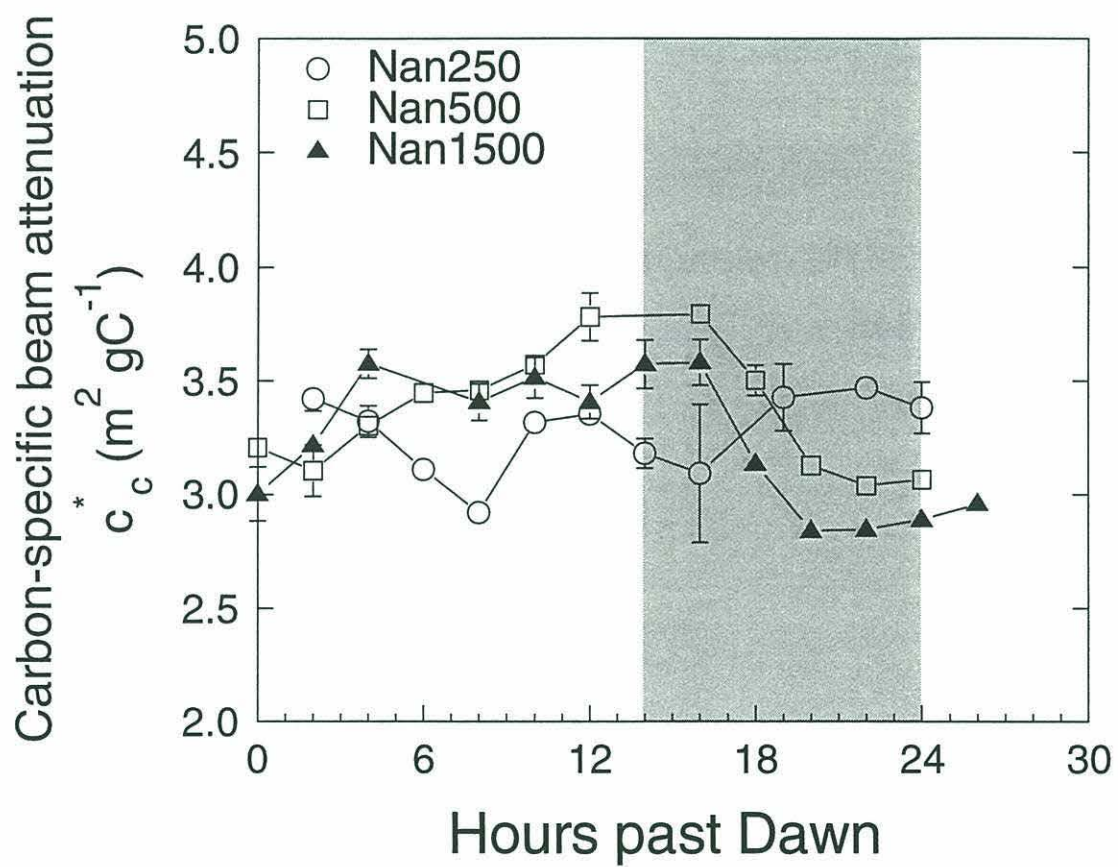
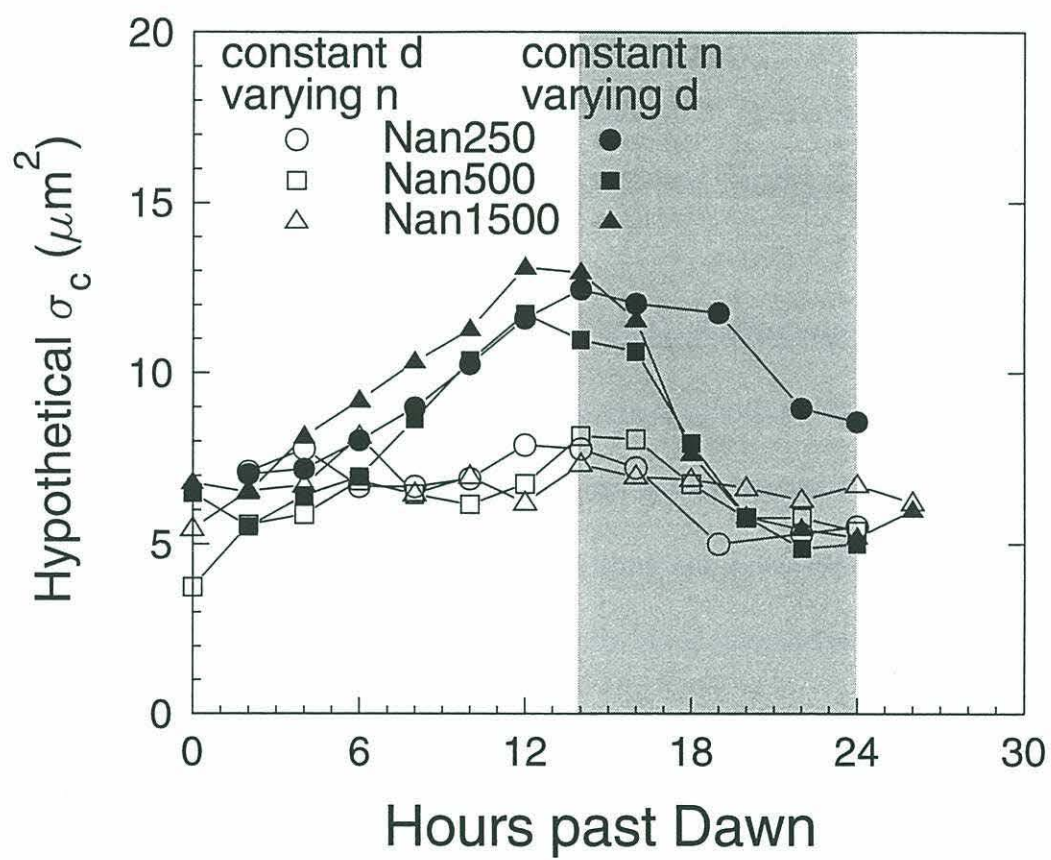


Figure 15. Results of the sensitivity analysis of the attenuation cross section (σ_c) to varying refractive index, n , at a constant diameter, d (open symbols) and varying diameter at a constant refractive index (filled symbols).



over a three-day sampling; Stramski and Reynolds 1993). Our average c_c^* for *Nannochloris* of $3.38 \text{ m}^2 (\text{g C})^{-1}$ ($n = 49$, $sd = 0.34$) is similar to that of Stramski and Reynolds for *T. pseudonana*, 3.81. Stramski et al. found *Synechococcus* to have a lower value of $2.48 \text{ m}^2 (\text{g C})^{-1}$. We also found a lower c_c^* for *Synechococcus*, 1.52 ($n = 8$, $sd = 0.26$, for four strains, unpublished data). It therefore appears that there can be significant differences in the carbon-specific beam attenuation both with time of day and among different species. Thus it may be possible to improve estimates of production by using a different c_c^* for dawn and for dusk (the minimum and maximum of beam attenuation in the field) and for different species (if the composition of the phytoplankton community is known; DuRand and Olson in press).

Contributions of diameter and refractive index to σ_c

We performed a sensitivity analysis to determine the contributions of diameter and refractive index to the attenuation cross section (σ_c). Using the data from each carboy in each diel experiment, the mean cell diameter was held constant at the minimum value near dawn (two hours after dawn), and Q_c and σ_c were calculated to determine the effect of changing refractive index on σ_c (n' , which had only a small effect, was assumed to be zero for these calculations). Similarly, the refractive index was held constant (at its value at two hours after dawn) and Q_c and σ_c were calculated to determine the effect of changing diameter on σ_c . The observed variations in diameter through the diel cycle had a much greater effect on σ_c than did those of refractive index (Fig. 15). To quantify this effect, we performed a linear

regression between the measured attenuation cross section (data in Fig. 9A) and the hypothetical attenuation cross section (Fig. 15). We found that 79% of the variance in measured σ_c is explained by the hypothetical σ_c when only diameter is allowed to vary. Thus the diel variations in σ_c for *Nannochloris* are primarily influenced by changes in cell size and not in refractive index. In contrast to this, studies on *Thalassiosira pseudonana* and *Synechococcus*, have shown that changes in cell size were equally or less important than changes in refractive index in determining σ_c , depending on the time of day (Stramski and Reynolds 1993, Stramski et al. 1995).

CONCLUSIONS

At six different light levels ranging from 60 - 1500 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ (specific growth rates from ~ 0.2 to $\sim 0.7 \text{ d}^{-1}$), cell division of *Nannochloris* was tightly phased to the light:dark cycle, occurring soon after dark. There were pronounced diel patterns, with a minimum near dawn and a maximum near dusk, in cell size and in cell-specific attenuation, absorption, flow cytometric light scatter and carbon. The diel variations in the attenuation cross section were primarily influenced by the changes in cell size due to growth and division, while changes in refractive index had only a small effect. Because eukaryotic cells in the size range of *Nannochloris* are major constituents of many phytoplankton communities, the results presented here have important consequences for the interpretation of diel variations in optical properties observed in the ocean. Since properties measurable by flow

cytometry were determined, the relationships calculated in this work can be applied to field measurements made using flow cytometry to distinguish particular phytoplankton populations. It should thus be possible to account for variations in bulk optical properties such as beam attenuation (Chapters 3 and 4). In addition, since the variations observed represent growth processes of the cells, the relationships determined in this work can be used to estimate *in situ* growth rates of different phytoplankton groups (Chapter 5).

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Chapter 3

Contributions of phytoplankton light scattering and cell concentration changes to diel variations in beam attenuation in the equatorial Pacific from flow cytometric measurements of pico-, ultra-, and nanoplankton

This chapter has been accepted for publication in Deep-Sea Research with
R. J. Olson as co-author.

ABSTRACT

The phytoplankton biomass in the equatorial Pacific is dominated by picophytoplankton (*Prochlorococcus*, $< 1\ \mu\text{m}$, and *Synechococcus*, $\sim 1\ \mu\text{m}$ diameter), and by mixed populations of ultraphytoplankton ($1\text{--}2\ \mu\text{m}$ diameter) and nanophytoplankton ($2\text{--}20\ \mu\text{m}$, mostly $2\text{--}3\ \mu\text{m}$). The mean forward light scatter of each of these populations, measured on individual cells by flow cytometry, increased during the day and decreased during the night. This pattern was similar to that of bulk red beam attenuation due to particles. In contrast, cell concentration changes did not correspond to the patterns in beam attenuation.

Using a combination of empirical calibrations relating beam attenuation to flow cytometric measurements of pure cultures of phytoplankton in the laboratory, and Mie theory, we estimated the contributions of different groups of phytoplankton to the diel variations in beam attenuation observed. Our results indicate that the phytoplankton assemblage measured by flow cytometry can account for essentially all of the diel variation in the beam attenuation signal. In most instances the nanophytoplankton were the largest contributor to total beam attenuation due to phytoplankton, but the ultraphytoplankton usually were more important in determining the diel variations in this property. *Prochlorococcus* were a smaller but appreciable contributor to beam attenuation changes, and *Synechococcus* were much less important. These findings emphasize the need to characterize the composition of the phytoplankton community in order to use beam attenuation as a tool for monitoring productivity.

INTRODUCTION

Diel variations in red beam attenuation have been observed in many parts of the world's oceans (SIEGEL *et al.*, 1989; HAMILTON *et al.*, 1990; GARDNER *et al.*, 1990, 1995). Since beam attenuation has been shown to be correlated with suspended particle mass, analysis of this diel pattern has been proposed as a method for estimating primary production (SIEGEL *et al.*, 1989). However, the relationship between particle mass and beam attenuation is not a constant, but depends on the size distribution and optical properties of the constituent particles (BAKER and LAVELLE, 1984). In order to better interpret diel variations in the bulk water inherent optical property of beam attenuation, we used flow cytometry to measure the light scattering and cell concentrations of different groups within the phytoplankton, whose dynamics have been presumed to cause the variations.

The beam attenuation coefficient (c) is composed of the scattering coefficient (b) and the absorption coefficient (a), where $c(\lambda) = a(\lambda) + b(\lambda)$. Theoretical and experimental studies on a variety of phytoplankton species have indicated that attenuation is dominated by scattering, and only a small portion of the signal is due to absorption (BRICAUD *et al.*, 1988). Beam attenuation can be partitioned into the contributions of pure seawater (c_w), dissolved organic material (yellow substance, c_y), and particles ($c_{\text{particles}}$): $c = c_w + c_y + c_{\text{particles}}$. C_w is a constant and c_y is negligible in the open ocean at the red wavelength (660 nm) of the transmissometer light source (BARTZ *et al.*, 1978), thus $c_{\text{particles}}$ determines the variations in c . In this work, $c_{\text{particles}}$

will be further subdivided into the contributions of phytoplankton (c_{phyto}) as distinguished using flow cytometry, and of non-phytoplankton ($c_{\text{non-phyto}}$). C_{phyto} consists of the contributions of different groups of phytoplankton: *Prochlorococcus* ($< 1 \mu\text{m}$ diameter, c_{pro}), *Synechococcus* ($\sim 1 \mu\text{m}$, c_{syn}), ultraphytoplankton ($1\text{-}2 \mu\text{m}$ cells, c_{ult}), and nanophytoplankton (mostly $2\text{-}3 \mu\text{m}$ cells, but including cells up to $20 \mu\text{m}$, c_{nan}).

Previous investigators observed diel variations in beam attenuation with a minimum near sunrise and a maximum near sunset. From observations in the North Pacific, SIEGEL *et al.* (1989) suggested that these patterns could be accounted for by a balance between photosynthetic production of ultraplankton (causing an increase in particles) and grazing by microzooplankton (causing a decrease in particles). During spring bloom conditions in the North Atlantic, diel variations in beam attenuation were measured in the mixed layer, and the daytime increases in beam attenuation were found to be primarily due to increases in particulate matter concentration (GARDNER *et al.*, 1993). STRAMSKA and DICKEY'S time series measurements of water column bio-optical properties (1992) in the North Atlantic also suggested a relationship between phytoplankton production and beam c .

In an attempt to elucidate this relationship for conditions encountered in the equatorial Pacific, we have related direct measurements of changes in the light scattered by the phytoplankton to changes in beam c over the diel cycle from sampling experiments in April and October 1992. Our observations reveal diel variations in mean forward light scatter for populations of eukaryotic and prokaryotic

phytoplankton that are minimum near dawn and maximum near dusk, the same pattern as that seen for bulk beam attenuation (GARDNER *et al.*, 1995). In contrast, phytoplankton cell concentrations in these samples do not have the same diel pattern; indeed, they usually have the opposite pattern. Using laboratory calibrations on phytoplankton combined with theoretical calculations, we estimated the contributions that each phytoplankton population made to the diel variations in beam attenuation. The implications of these findings for the use of diel variations in beam c for estimating productivity will be discussed.

MATERIALS AND METHODS

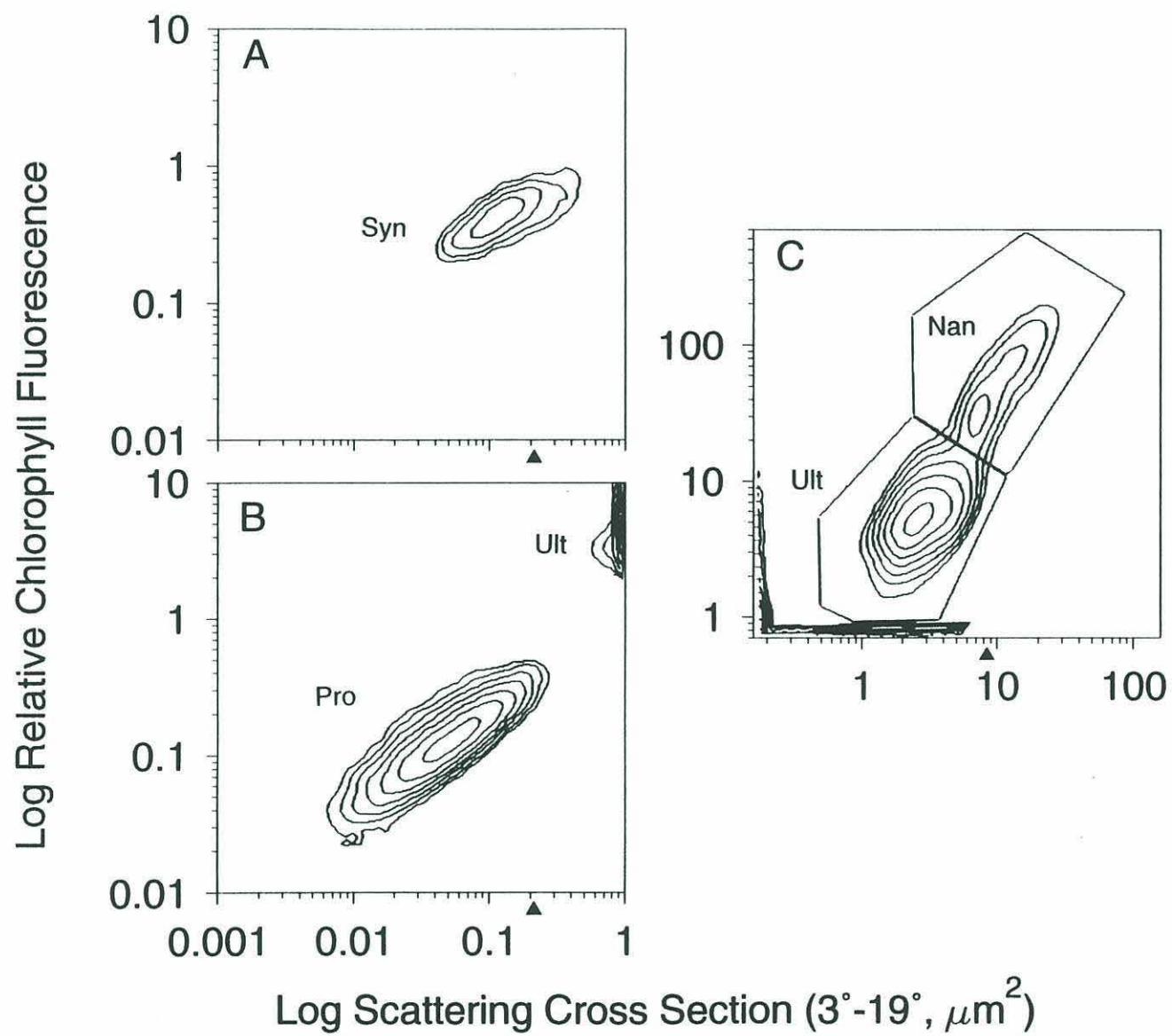
Field Studies

Intensive diel sampling was performed on five occasions during the two JGOFS Time Series cruises in the equatorial Pacific (cruises 8 and 12 of R.V. *Thomas Thompson* to 0°, 140° W). Of these, we will discuss only three: the first diel study on cruise TT8, 1 April 1992 (TT8D1), and the first two diel studies on cruise TT12, 5 and 11 October 1992 (TT12D1 and TT12D2). Portions of the data from the other two were lost due to equipment failure. In each case, CTD casts were made every three hours for 24 hours and samples for flow cytometric analysis were taken with Niskin bottles at 0, 15, 30, 45, 60, 75, 90, and 100 m. A limited number of daily dawn and dusk casts during each cruise were also sampled for flow cytometry.

The eukaryotic phytoplankton were analyzed immediately on board the ship using an EPICS V flow cytometer modified to analyze 50 ml seawater samples at 5-10 ml min⁻¹ (OLSON *et al.*, 1991, 1993). Samples were kept at room temperature in the dark until analysis; the surface samples were analyzed first and all depths were completed within two hours of collection (see below for comparison of test samples). Fluorescent microspheres (3.79 µm, Polysciences) were added to each sample as an internal standard. Eukaryotic phytoplankton from 1-20 µm in diameter were counted and, for each particle, forward light scatter (FLS, 3°-19° at 488 nm), side scatter, red (chlorophyll, 660-700 nm) fluorescence, and orange (phycoerythrin, 530-630) fluorescence were collected. Phytoplankton populations were discriminated on plots of forward light scatter vs. chlorophyll fluorescence (Fig. 1) using CytoPC software (D. Vaultot, Station Biologique, Roscoff, France). During each cruise the phytoplankton groups were sorted on the flow cytometer and examined microscopically. Typically, the most abundant population of eukaryotic phytoplankton consisted of 1-2 µm-diameter coccoid cells (referred to here as the ultraphytoplankton). The population referred to here as the nanophytoplankton primarily consisted of 2-3 µm diameter coccoid cells, but also included cells as large as 20 µm. In addition, there were a small number of phytoplankton cells which were off-scale on chlorophyll fluorescence (but not on FLS) at the flow cytometer settings we used. These were not included in the nanophytoplankton category, but were considered separately (referred to as "off-scale"). Analysis of test samples (one each from 0, 15, and 30 m during TT8) kept for greater than two hours in the dark at

Figure 1. Typical flow cytometric signatures of prokaryotic picoplankton (A and B) and eukaryotic phytoplankton (C) populations (from a surface sample from TT12D2) after gating to clearly show the separate populations. Contour plots show relative chlorophyll fluorescence (red, 660-700 nm) vs. scattering cross section (3° - 19° in μm^2 , calculated from FLS). Each contour level represents a doubling in cell number.

A) *Synechococcus* (Syn), were first distinguished by gating on their high phycoerythrin fluorescence. B) *Prochlorococcus* (Pro) population after removing *Synechococcus* by gating. C) Polygons outline the populations of the ultraphytoplankton (Ult) and the nanophytoplankton (Nan). A) and B) are from analysis of the sample run with a high-sensitivity configuration of the flow cytometer; the scattering cross section of $0.57 \mu\text{m}$ beads is marked by a triangle. C) is from the less sensitive high flow rate configuration of the flow cytometer used to analyze the less abundant eukaryotic phytoplankton (scattering cross section of $3.79 \mu\text{m}$ beads marked by a triangle).



room temperature showed an average 4% decrease from initials for ultraphytoplankton FLS and a 1% increase for nanophytoplankton FLS.

The picoplankton *Synechococcus* and *Prochlorococcus* were analyzed on shore from samples preserved with 0.125% glutaraldehyde and stored in liquid nitrogen (VAULOT *et al.*, 1989, OLSON *et al.*, 1993), using a high sensitivity configuration of an EPICS 753 flow cytometer (OLSON *et al.*, 1993), and 0.57 μm beads (Polysciences) as an internal standard. *Synechococcus* were distinguished by their high phycoerythrin fluorescence and *Prochlorococcus* by their low FLS and chlorophyll fluorescence (Fig. 1).

Laboratory Studies

Three laboratory experiments (shore-based) were performed in which the chlorophyte *Nannochloris* sp. (2.5-3 μm diameter) was grown in f/2 medium in replicate carboys at 22° C. For two of the experiments the cultures were grown in an incubator on a 14:10 L:D cycle, at 250 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ for one experiment and at 500 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ for the other (Cool-White fluorescent lights). A third experiment was performed under natural sunlight with an average integrated light level of 1500 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ (14 h daylight on a mostly sunny day in August). The cultures were sampled at mid-exponential phase every two hours for 24 hours. Triplicate samples were analyzed immediately on the flow cytometer, as at sea except that the culture was diluted 100 μl : 50 ml 0.22 μm filtered seawater (FSW), to obtain cell FLS and concentration. Beam attenuation was measured with a

SeaTech transmissometer (25-cm pathlength; $\lambda = 665$ nm) enclosed by a PVC sleeve with a sampling port, by adding progressively 8, 12, and 16 ml of culture to 1600 ml FSW in the transmissometer. For each measurement, the culture was added to the FSW, the mixture was stirred, and any bubbles were cleared from the transmissometer windows before taking the reading. The procedure was repeated twice for each culture. Replicate samples (25 ml) were filtered onto pre-combusted GF/F filters, frozen, and later dried and analyzed on a Perkin-Elmer 2400 CHN Analyzer. Absorption was measured using the opal glass method (SHIBATA, 1958) in a spectrophotometer (Bausch and Lomb Spectronic 88) to confirm that absorption was small relative to scattering (see Introduction).

A similar experiment was performed on *Synechococcus*, with 4 strains (WH8012, 8015, 8018, and 8103; from J. Waterbury, WHOI) grown at $75 \mu\text{mol photons m}^{-2} \text{ s}^{-1}$ on a 14:10 L:D cycle and sampled at "dawn" and "dusk." For the beam attenuation measurements, 20-35 ml of the culture was added to 1600 ml FSW and mixed in a flask and then poured into the transmissometer "sleeve." Replicate measurements were made for each strain. Measurements of carbon content were also made.

Conversion of forward light scatter to scattering cross section

Our measurements of forward light scatter (3° - 19°) are expressed relative to light scattered by standard beads. To compare our results to theory, we need to express them in more meaningful units. We therefore calculated the scattering cross

section (μm^2) of the cells by reference to the calculated scattering cross section (3° - 19°) of the beads, for which the size, refractive index, and absorption are known (BOHREN and HUFFMAN, 1983; ACKLESON and SPINRAD, 1988):

$$\frac{\text{FLS cells}}{\text{FLS beads}} * \text{scat. cross section beads } (\mu\text{m}^2) = \text{scat. cross section of cells } (\mu\text{m}^2).$$

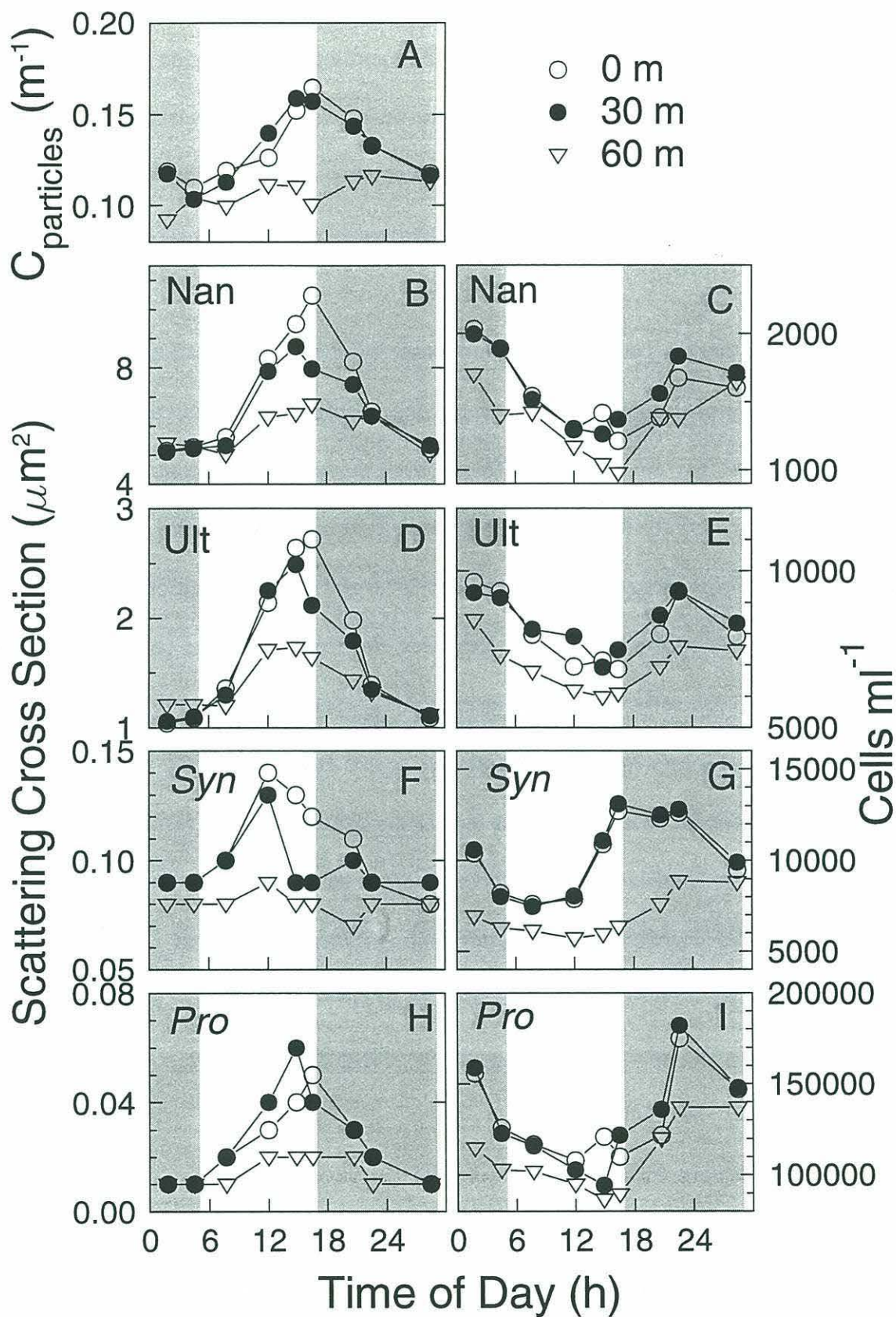
Mie calculations (program of ACKLESON and SPINRAD, 1988) of scattering over the collection angles of the flow cytometer were performed using the diameter of the beads ($3.79 \mu\text{m}$), the refractive index of polystyrene (1.59 relative to air), the concentration of beads (calibrated on the flow cytometer), and the absorption coefficient of the "cellular" material ($0.12 \mu\text{m}^{-1}$, measured on a suspension of beads using the opal glass technique on a Shimadzu UV-260 spectrophotometer; SHIBATA, 1958). Similar calculations were performed for the $0.57 \mu\text{m}$ beads (used in the picoplankton samples), except that the absorption was assumed to be zero (absorption has a negligible effect at this size). The angles of collection of the flow cytometer FLS were estimated to be 3° - 19° , by measuring the angles of the light intercepted by the FLS detector, and by comparing the ratio of measured FLS of different sized beads to the ratio of theoretical determinations of scattering cross sections of the same beads.

RESULTS AND DISCUSSION

Diel Patterns in Phytoplankton and Beam Attenuation

During each of the three diel time series we sampled, the mean scattering cross section (calculated from flow cytometric forward light scatter) of nanophytoplankton, ultraphytoplankton, and *Prochlorococcus* cells was consistently at a minimum near dawn and maximum near dusk (Fig. 2B-E, H, I). The *Synechococcus* mean scattering cross section was also at a minimum near dawn, but its maximum was near noon rather than dusk (Fig. 2F, G). These patterns were visible down to 100 m for TT8D1 and TT12D1 and to 75 m for TT12D2; the amplitude of the diel variation tended to decrease with depth. Cell concentrations generally decreased during the day and increased at night (or increased at the beginning of the night and then leveled off or decreased before dawn) down to at least 60 m for all diel samplings, except for *Synechococcus*, for which cell concentrations increased from near noon until dusk and then decreased during the night hours. The patterns in beam attenuation due to particles ($c_{\text{particles}}$, GARDNER *et al.*, 1995) were similar to those of FLS, with a minimum near dawn and a maximum near dusk (Fig. 2A). This pattern was seen down to 100 m during TT8D1, down to 60 m during TT12D1, and down to 45 m during TT12D2. As with FLS, the amplitude of the diel change in beam attenuation was smaller at depth than in the upper water column.

Figure 2. Time series data for the selected depths 0 m (○), 30 m (●), and 60 m (▽) from TT12D2. A) Beam attenuation due to particles ($c_{\text{particles}}$ in m^{-1} , data from W. Gardner). Forward light scattering cross section (μm^2), and cell concentration (cells ml^{-1}) for nanophytoplankton (B and C), ultraphytoplankton (D and E), *Synechococcus* (F and G), and *Prochlorococcus* (H and I). Note that the y-axis for each plot is scaled independently. Patterns for TT8D1 and TT12D1 were similar to those shown here, though the cell concentrations differed. At dawn in the near surface depths, nanophytoplankton concentrations were 1500 ml^{-1} (TT8D1) and 1800 ml^{-1} (TT12D1 and D2). Ultraphytoplankton concentrations were 4000 ml^{-1} (TT8D1), 7500 ml^{-1} (TT12D1), and 9500 ml^{-1} (TT12D2). *Synechococcus* concentrations were 9000 ml^{-1} (TT8D1), 6000 ml^{-1} (TT12D1), and 8000 ml^{-1} (TT12D2). *Prochlorococcus* concentrations were $2 \times 10^5 \text{ ml}^{-1}$ (TT8D1), $1 \times 10^5 \text{ ml}^{-1}$ (TT12D1), and $1.3 \times 10^5 \text{ ml}^{-1}$ (TT12D2).



In addition to the diel sampling periods, we analyzed a few "pairs" of data from consecutive dawn/dusk profiles (two for TT8 and four for TT12; data not shown). In all of these pairs, at every depth that bulk beam c was lower at dawn than at dusk, the FLS for ultra- and nanophytoplankton showed the same pattern. The cell concentration for both ultra- and nanophytoplankton were generally higher at dawn than at dusk.

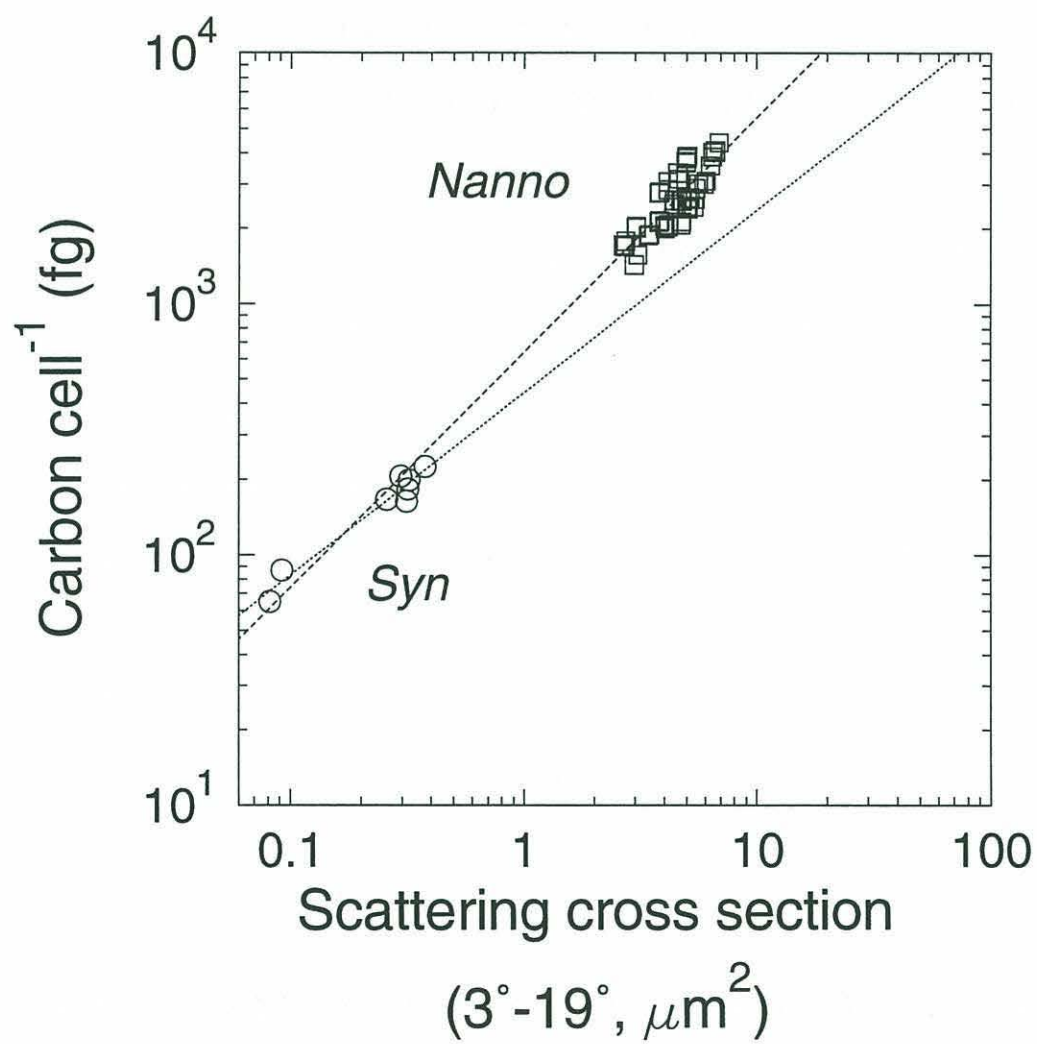
It is clear that the patterns in the bulk property of beam attenuation are similar to the patterns in mean light scatter per cell of different populations of phytoplankton, and they are generally different from those in cell concentration. The increase in light scatter during the day can be explained by growth of the phytoplankton cells as they photosynthesize and fix carbon. As a result of the increase in light scatter, the total beam c increases, despite the fact that cell concentrations are generally decreasing during the day (presumably due to grazing). The maximum in cell light scatter almost always occurs at the same time as the minimum in cell concentration, near dusk (except for *Synechococcus*, for which they occur at noon). The implication is that the cells begin to divide at this time, so that the average cell size decreases, with a corresponding increase in the cell concentration (newly divided cells). Diel patterns in DNA per cell for *Prochlorococcus* in the same samples support this interpretation (VAULOT *et al.*, 1995; BINDER *et al.*, submitted).

Changes in light scatter can be due to changes in refractive index or shape as well as cell size, although these have usually been considered less important than changes in size (JERLOV, 1976; GORDON *et al.*, 1984). ACKLESON *et al.* (1993)

reported variations in refractive index for phytoplankton cultures at high light intensity which were to some extent carbon independent. However, in laboratory experiments with the diatom *Thalassiosira pseudonana* (STRAMSKI and REYNOLDS, 1993) and with *Synechococcus* (STRAMSKI *et al.*, 1995) strong correlations between refractive index and intracellular carbon concentration were found. In our laboratory experiments on *Nannochloris* and *Synechococcus*, the cell scattering cross section was strongly correlated with cell carbon, though the relationship may be different for the two species (Fig. 3). During the same cruises we discuss, GARDNER *et al.* (1995) found that beam *c* was well correlated with particle mass. Thus we interpret the daytime increases in scatter we observed as due primarily to cell growth (carbon fixation).

A number of factors can contribute to the decrease in beam *c* during the night; these include cell division, respiration, mixing, cell aggregation, and grazing. According to the anomalous diffraction approximation of Mie scattering theory (VAN DE HULST, 1957; MOREL and BRICAUD, 1986), the redistribution of cell material into smaller packages during cell division should cause a decrease in *c* for cells smaller than about 5 μm . For example, *c* would decrease by 12% if a 2 μm diameter cell divided into two cells, each with half the original volume. Loss of cell material through respiration would be expected to reduce beam *c* by an amount that depends on the growth conditions but which is not easy to quantify. Nocturnal deepening of the mixed layer has been suggested as a means of removing particles from the upper, productive water; such a mechanism was clearly documented during the Time Series cruises (GARDNER *et al.*, 1995).

Figure 3. Relationship between carbon per cell and forward scattering cross section (3° - 19° , measured with the flow cytometer,) for laboratory cultures of *Nannochloris* and *Synechococcus* measured over the light:dark cycle. Data for *Nannochloris* are from diel experiments at two different light levels (250 and $500 \mu\text{mol photons m}^{-2} \text{s}^{-1}$). Data for *Synechococcus* are from "dawn" and "dusk" samplings of four different strains. A linear regression for each cell type is shown.



Since particles larger than about 20 μm are typically too rare in the open ocean to be effectively sampled by the transmissometer (PAK *et al.*, 1988; GARDNER *et al.*, 1993), any process which repackages the phytoplankton biomass into large (and therefore inadequately sampled) parcels will cause beam c to decrease. Walsh and Gardner (pers. comm.) measured aggregates in the North Atlantic and saw a diel variation in aggregate abundance, which they suggested could be a factor in the decrease in beam c at night. Grazing by zooplankton that are not effectively sampled by the transmissometer would decrease beam c by the same mechanism.

Calculation of beam attenuation due to phytoplankton

To obtain estimates of beam c from field measurements of scattering cross sections and cell concentrations of phytoplankton populations, we began with an empirical calibration using *Nannochloris* and *Synechococcus* cultures. Attenuation cross section (beam c normalized to cell concentration) was regressed on scattering cross section (linear regression of the log transformed data, $r^2 = 0.97$; Fig. 4). This empirical relationship was compared to Mie theory (an iterative calculation using program from ACKLESON and SPINRAD, 1988) assuming a constant refractive index of 1.05 and absorption of cellular material (a_{cm}) of either 0.1 or 0.2 μm^{-1} and using a range of cell diameters from 0.5-8 μm (Fig. 5). Changing the refractive index to 1.07 changes the relationship between the attenuation and scattering cross sections only slightly (not shown). Though a linear regression of our empirical data is reasonably close to Mie theory calculations over the range of cell sizes from

Synechococcus to *Nannochloris*, extending the same regression to much smaller cells (such as *Prochlorococcus*) is not appropriate. We do not yet have an axenic culture of *Prochlorococcus*, so we have not empirically confirmed the relationship between attenuation and scattering cross sections for these cells. For *Prochlorococcus*, we obtained a relationship analogous to that used for the other populations by using theoretically-derived attenuation cross sections ($n = 1.05$, $a_{cm} = 0.2 \text{ } \mu\text{m}^{-1}$) obtained over the range of *Prochlorococcus* scattering cross sections observed in the field.

The appropriate calibration was applied to the field data of mean scattering cross section for different phytoplankton populations to obtain beam c per cell ml^{-1} (i.e., attenuation cross section) for each population. This was multiplied by the cell concentration to obtain an estimate of beam attenuation due to each specific group of phytoplankton. The diel patterns in beam c due to each of the populations are qualitatively quite similar to that of $c_{\text{particles}}$ (Fig. 6) and to those of each other, indicating that all the phytoplankton groups are contributing to attenuation in a similar manner. At first glance the patterns in c of the groups appear to follow those of scattering cross section (see Fig. 2B, D, F, H), which was the original observation from flow cytometric FLS measurements. However, upon closer examination it is apparent that the amplitude of the diel excursions in c are damped relative to those in mean scattering. This is the result of the opposing trends in scattering and cell concentration. At 60 m in this diel sampling, for example, the diel pattern in c is no longer visible, even though both scatter and cell concentration patterns are still evident. Apparently cell growth at this depth is limited by light and only balances

Figure 4. Relationship between attenuation cross section (measured with a transmissometer) and forward scattering cross section for cultures of *Nannochloris* and *Synechococcus* measured over the light:dark cycle. Data are from the same laboratory experiments as in Fig. 3, plus values from the natural light ($1500 \mu\text{mol photons m}^{-2} \text{s}^{-1}$) experiment on *Nannochloris*.

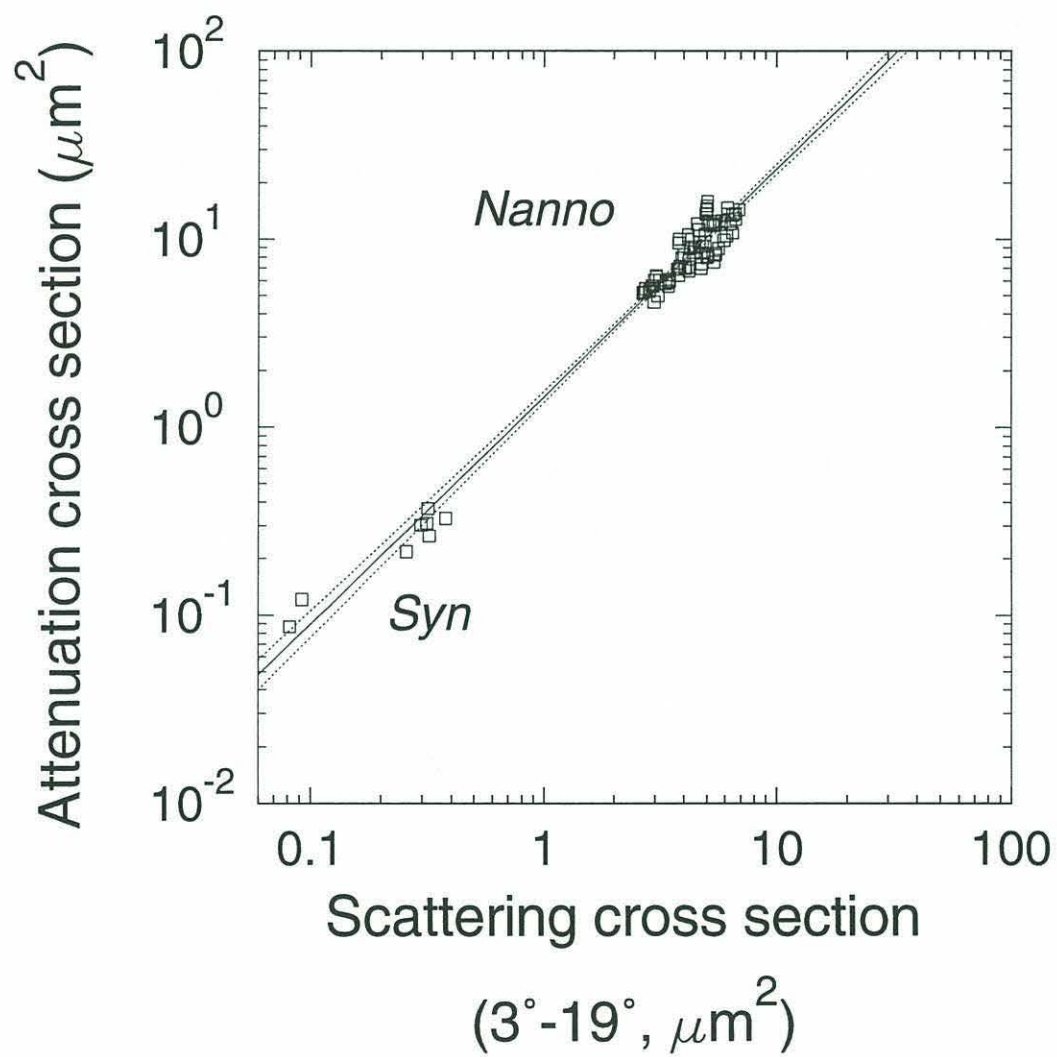


Figure 5. Comparison between empirical and theoretical relationships between the attenuation cross section (at the wavelength of the transmissometer, 665 nm) and the scattering cross section (3° - 19° ; at the wavelength of the flow cytometer, 488 nm). The dotted line shows the linear regression calculated from empirical data on *Synechococcus* and *Nannochloris* cells (shown in gray circles; data as in Fig. 4). The solid line shows the results of Mie theory calculations over a range of particle diameters (0.5 to 8 μm), assuming a constant refractive index of 1.05 and a constant absorption of cellular material (a_{cm}) of $0.1 \mu\text{m}^{-1}$ (and assuming the same a_{cm} for 488 and 665 nm). The dashed line shows similar calculations, except the a_{cm} is constant at $0.2 \mu\text{m}^{-1}$. The letters superimposed on the theory line mark the ranges of mean scattering cross section for specific types of cells observed in the field (P = *Prochlorococcus*, S = *Synechococcus*, U = ultraphytoplankton, and N = nanophytoplankton).

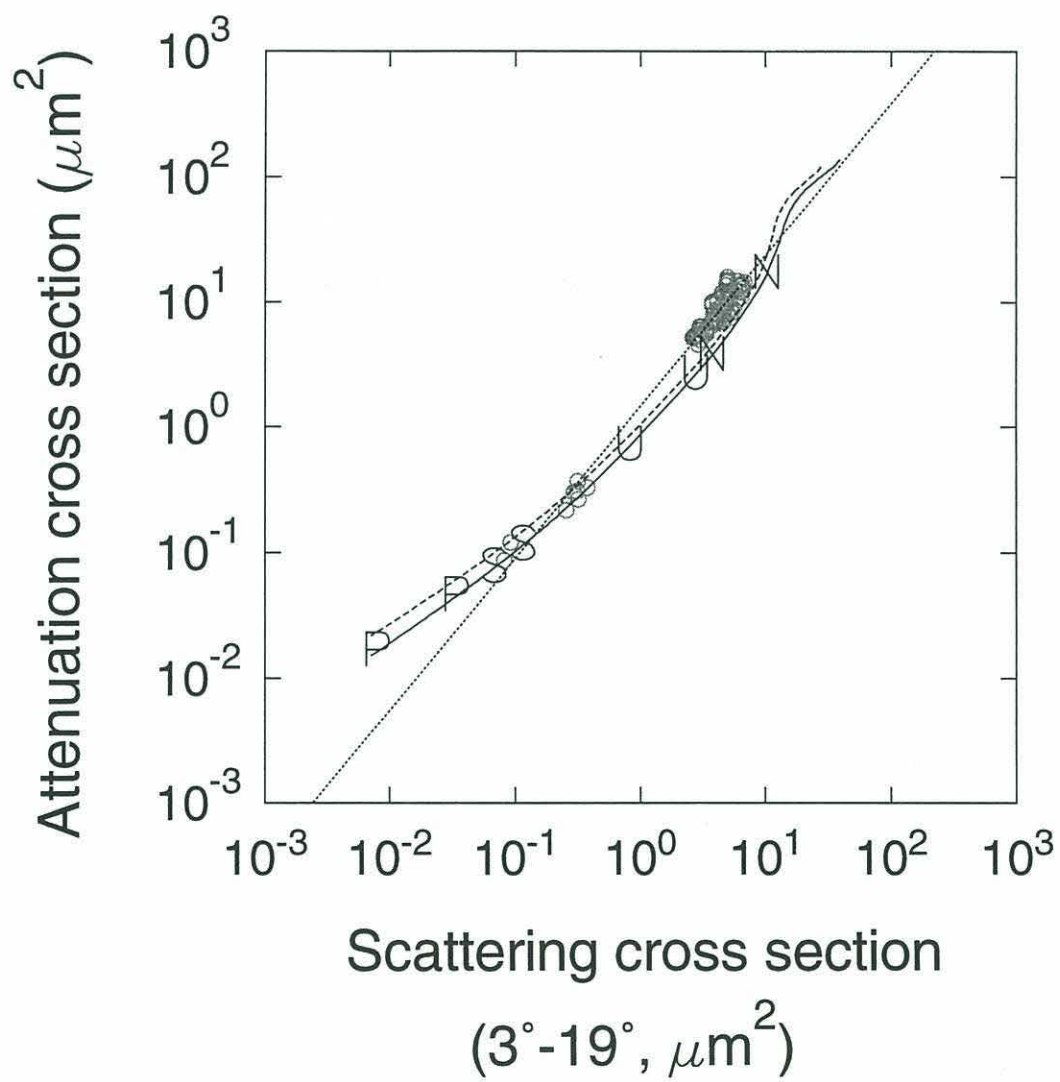
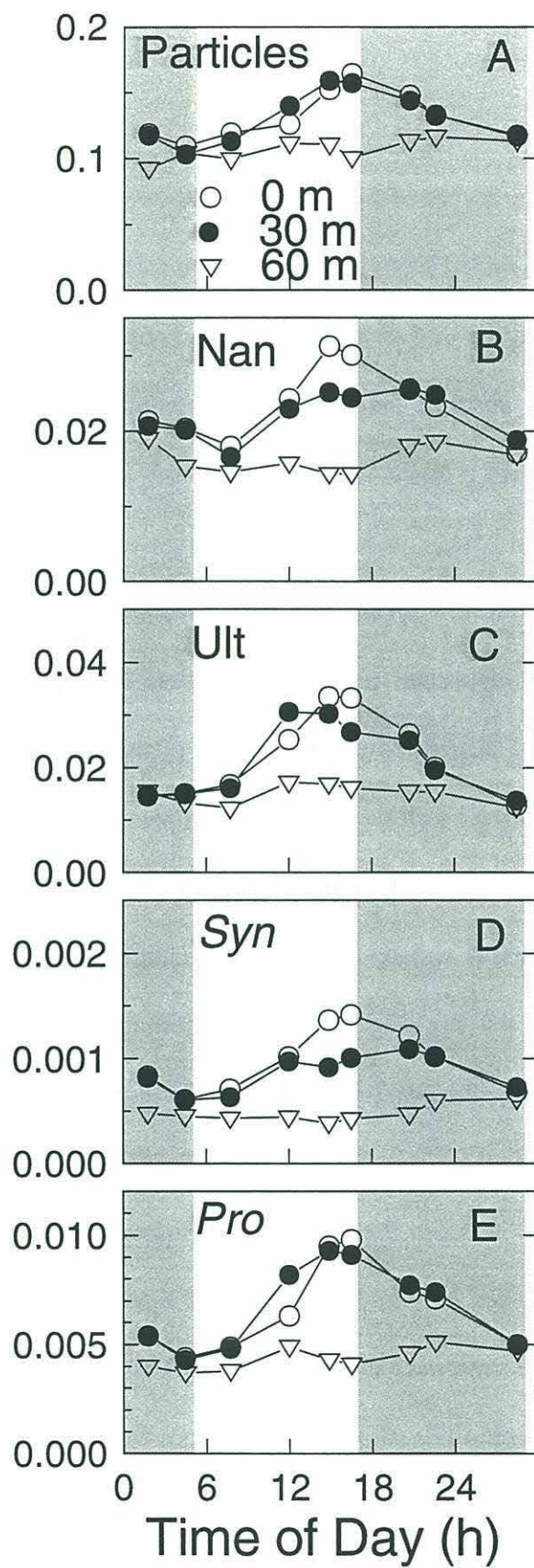


Figure 6. Time series of bulk beam attenuation due to particles (A) and calculated beam attenuation due to nanophytoplankton (B), ultraphytoplankton (C), *Synechococcus* (D), and *Prochlorococcus* (E) for TT12D2. Note that each plot is scaled independently in order to show the trends. Selected depths of 0 m (\circ), 30 m (\bullet), and 60 m (∇) are plotted here. Results from TT8D1 and TT12D1 were similar to those shown here.

Beam attenuation, $c \text{ (m}^{-1}\text{)}$



grazing losses during the day (in contrast to shallower depths where the increase in cell material during the day exceeds grazing). The increase in cell concentration at this depth during the night is caused not only by cell division, but by mixing of cells from above as a result of a deepened mixed layer (GARDNER *et al.*, 1995). We see evidence of this deepened mixed layer in the homogenization of all the phytoplankton properties at the end of the night (Fig. 2, 6).

Examination of the absolute magnitudes of the contributions of each phytoplankton group to beam c reveal several noteworthy features (Fig. 7). The most important is that most of c_{phyto} was due to the nano- and ultraphytoplankton, with a smaller proportion due to *Prochlorococcus*; *Synechococcus* was always less than 5% of the total. The "off-scale" phytoplankton were low in number but had large scattering, thus $c_{\text{off-scale}}$ accounted for about 10% of the total c_{phyto} (data not shown). Although the nanoplankton were in most samples the largest contributor to beam c , ultraphytoplankton often had greater changes in c during the diel cycle. In TT12D2, for example, this was apparently due to a greater relative increase in scatter of the smaller cells during the day (Fig. 2B, D), and perhaps also to a greater loss rate of the larger cells during the day (Fig. 2C, E). These data also show that there were differences in phytoplankton community composition (as expressed in terms of contribution to attenuation) between the two cruises: ultra-, and to a lesser degree nanophytoplankton, were more important during TT12 than TT8, while *Prochlorococcus* was more important during TT8 than TT12, especially at depth; similar trends in cell concentrations were also seen from flow cytometric analyses of daily samples from the two cruises (BINDER *et al.*, submitted).

Comparison of $c_{particles}$ and c_{phyto}

Since the pattern in c for each phytoplankton group matched that of $c_{particles}$, we wished to see if the changes in $c_{particles}$ could be quantitatively accounted for by the changes in c_{phyto} ($c_{phyto} = c_{Pro} + c_{Syn} + c_{ult} + c_{nan} + c_{off-scale}$). To do this, we examined the changes between successive samples in $c_{particles}$ ($\Delta c_{particles}$) and c_{phyto} (Δc_{phyto}). Thus, positive values were obtained when c 's were increasing and negative values when decreasing. $\Delta c_{particles}$ and Δc_{phyto} were strongly correlated ($r^2 = 0.67$) for all three diel experiments combined (Fig. 8). The intercept of the linear regression is negligible, and the slope (1.07) is not significantly different from 1 (t-test, $p < 0.05$). More detailed examination of the data in Fig. 8 reveals no trend with depth (data not shown). These results are important in that they suggest that the changes in $c_{particles}$ can be completely accounted for by the changes in c_{phyto} . They also imply that our flow cytometer is measuring nearly all of the particles contributing to the diel pattern in beam c . This is expected since the sampling volume of the transmissometer (44 ml) is close to that typically analyzed with our flow cytometer (50 ml).

Non-phytoplankton beam attenuation

Once we have estimated the beam attenuation due to phytoplankton, then we can by difference also estimate the non-phytoplankton contribution ($c_{non-phyto}$) which could include bacteria, microzooplankton, and detritus. $C_{non-phyto}$ is approximately 50% of the beam attenuation signal near the surface (Fig. 9); its contribution increases with depth as both the concentration and diel variations of

Figure 7. Time series of calculated beam attenuation due to each group of phytoplankton (c_{nan} (■), c_{ult} (▽), c_{syn} (○), and c_{pro} (▲)) plotted together to show their relative contributions. Selected depths of 0, 30, and 60 m are plotted for each of the diel samplings (TT8D1, TT12D1, and TT12D2).

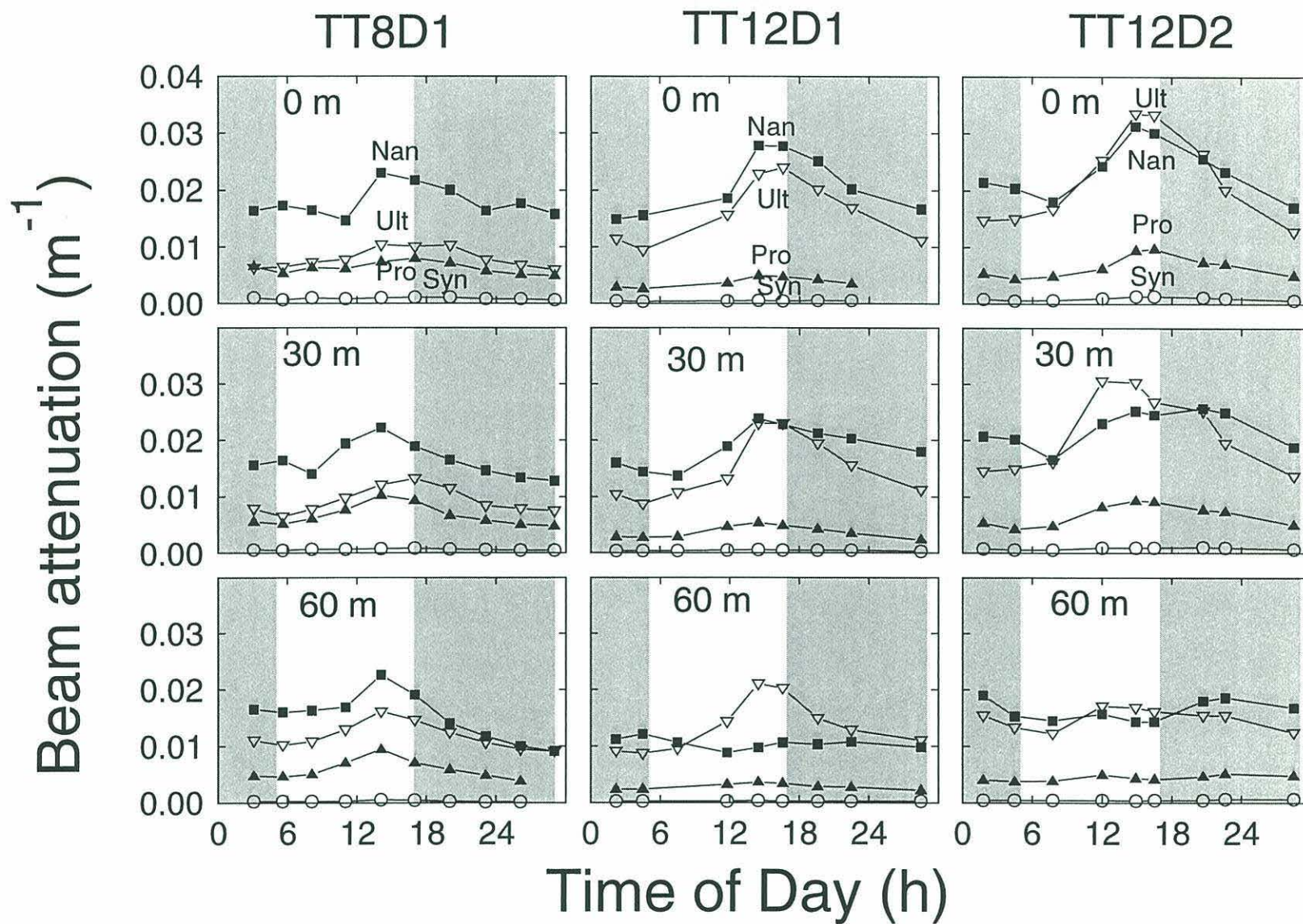


Figure 8. Relationship between $\Delta c_{\text{particles}}$ and Δc_{phyto} for TT8D1 (0-100 m), TT12D1 (0-100 m), and TT12D2 (0-75 m). Solid line is linear regression ($r^2 = 0.67$, $n = 149$).

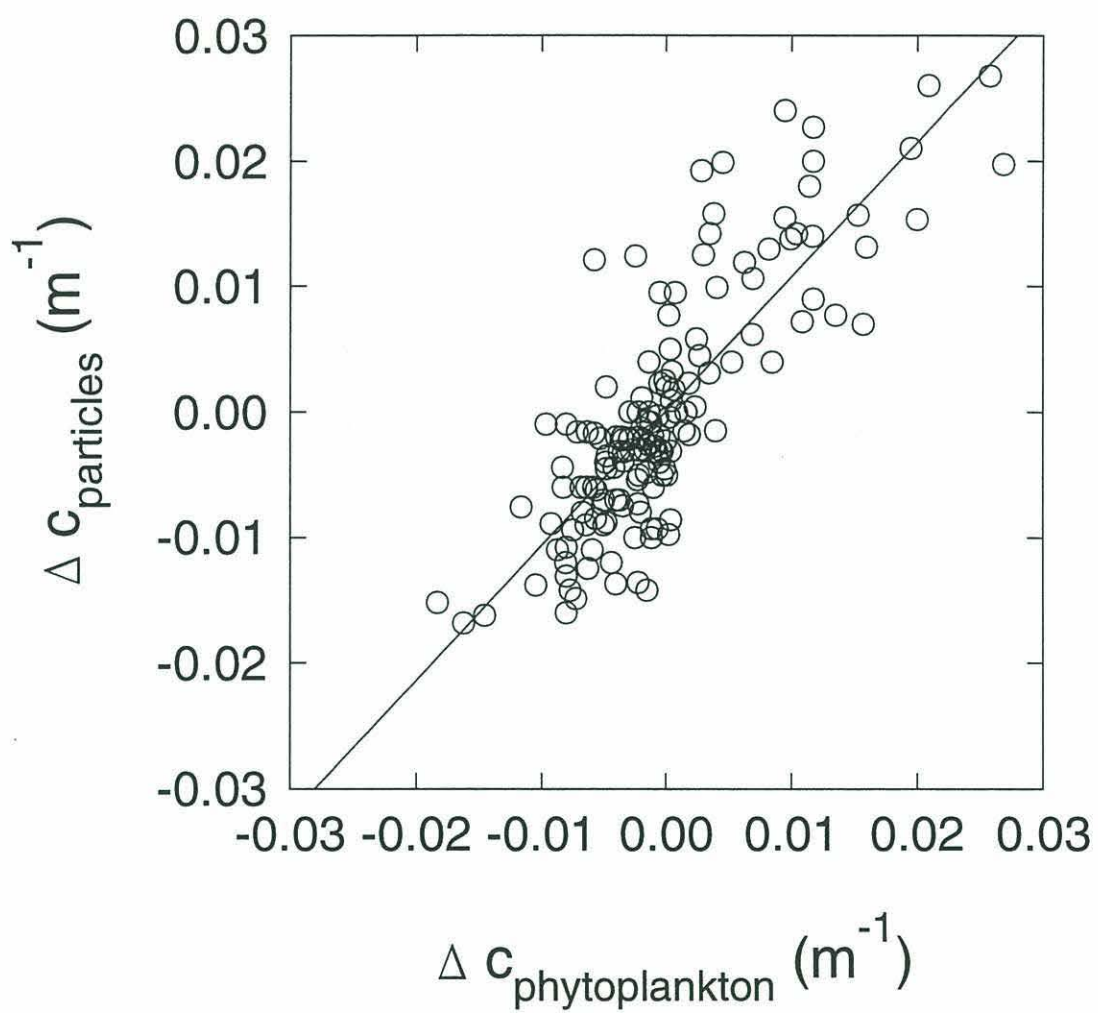
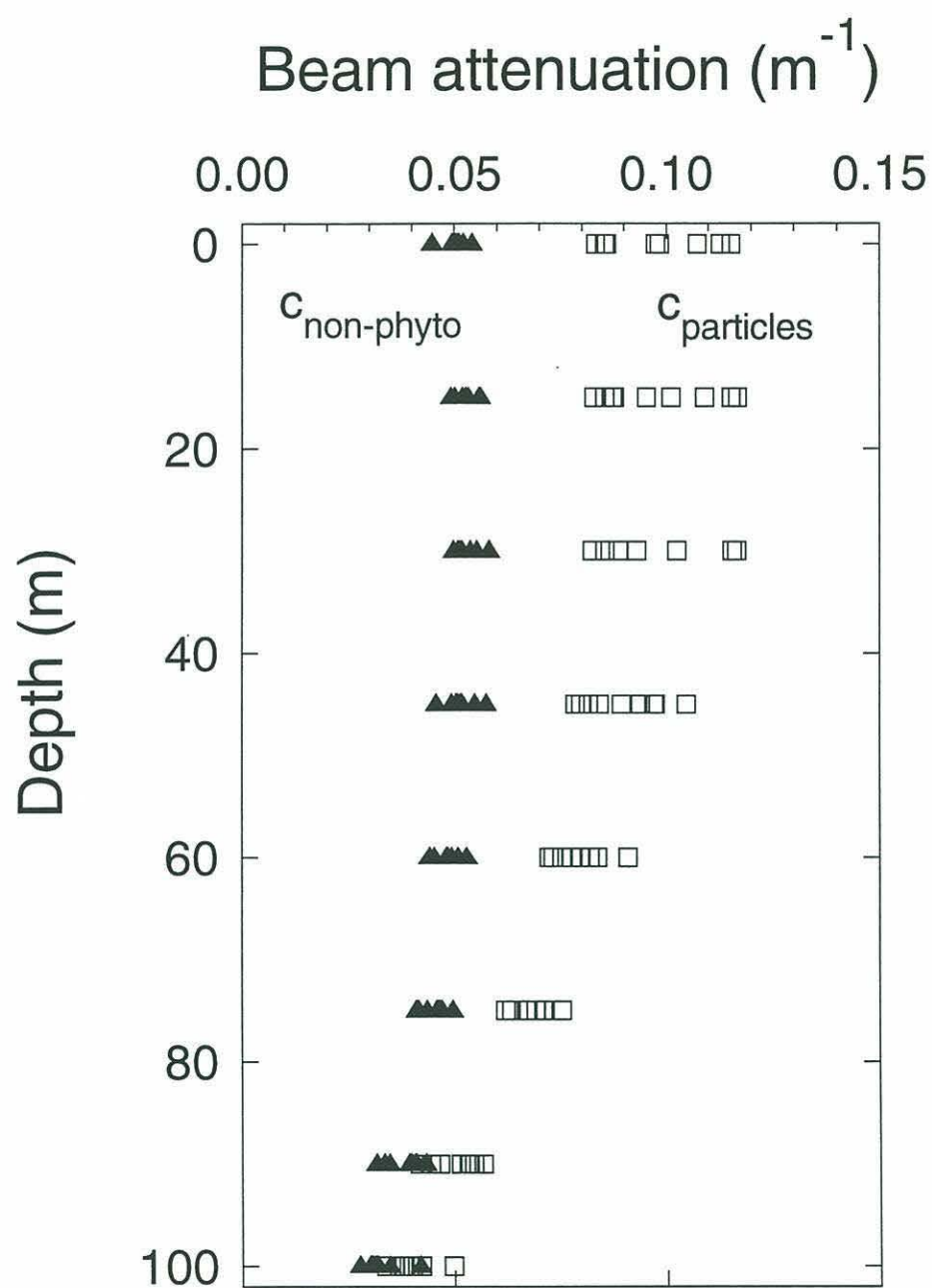


Figure 9. Depth profile for TT12D1 showing $c_{\text{particles}}$ (\square) and $c_{\text{non-phyto}}$ (\blacktriangle , the difference between $c_{\text{particles}}$ and c_{phyto}). At each depth, the time series of data are plotted. Data from TT8D1 and TT12D2 showed similar though noisier trends.



the phytoplankton scatter signal decrease. The variation in $c_{\text{non-phyto}}$ over time (the presumed cause of the spread in the data at each depth) is quite small compared to the variation in $c_{\text{particles}}$ as expected. An estimate of the bacterial contribution to $c_{\text{non-phyto}}$ can be made from bacterial abundance and volume data (DUCKLOW *et al.*, 1995). They determined bacterial abundance (from acridine orange direct counts) from a near noon Go-Flo rosette cast. We compared this data to our *Prochlorococcus* concentrations from the near noon CTD/Niskin rosette cast during each diel sampling and found that bacteria were 3-5 times more abundant in the upper 60 m during TT8D1, 5-8 times more during TT12D1, and 7-9 times more during TT12D2. Since the estimated bacterial cell volume ($0.03\text{-}0.04\ \mu\text{m}^3$, DUCKLOW *et al.*, 1995) is 3-9 times smaller than the estimated volume of *Prochlorococcus* (based on a diameter of $0.6\text{-}0.8\ \mu\text{m}$), the bacterial contribution to $c_{\text{non-phyto}}$ would be expected to be similar to that of *Prochlorococcus* (assuming the same relationship between size and attenuation for the two groups), though it could be three times greater or three times smaller. *Prochlorococcus* contribution to c_p in the upper 60 m at the near noon sampling was 9-12% during TT8D1 and 4-6% during TT12D1 and TT12D2. Note that these estimates of bacterial contribution to c_p may be high since the bacterial abundances from acridine orange counts probably included *Prochlorococcus* (which were also detected by flow cytometry) as well as heterotrophic bacteria.

Interpretation of diel patterns in c as production

Some investigators have used diel variations in beam attenuation to estimate community production, by relating the daily change in c to suspended particulate material and assuming that a given proportion (25%-40%) of it is particulate organic carbon (SIEGEL *et al.*, 1989; CULLEN *et al.*, 1992; WALSH *et al.*, in press). The effect is to use a constant conversion factor between beam c and carbon (c_c^* , the carbon specific beam attenuation) to estimate carbon production from the diel variations in c . SIEGEL *et al.* and CULLEN *et al.* used a c_c^* of $3.92 \text{ m}^2(\text{gC})^{-1}$, and obtained reasonable estimates of carbon production compared to carbon uptake production determinations. For the JGOFS Time Series cruises in the equatorial Pacific, WALSH *et al.* used a c_c^* of 5.5 for TT8 and 3.9 for TT12; the resulting estimates were approximately one-third of the ^{14}C -derived production estimates of BARBER *et al.* (1994). WALSH *et al.* explain this discrepancy as being the result of mixing, grazing, aggregation, and respiration which can affect the beam attenuation measurements but not the bottle incubations. However, if the c_c^* differs among species or over the course of the day (as indicated by STRAMSKI and REYNOLDS, 1993, and STRAMSKI *et al.*, 1995), analysis using a constant c_c^* may not be valid. STRAMSKI and REYNOLDS measured an average c_c^* of 3.81 for *Thalassiosira pseudonana*, which was relatively constant over the day:night cycle. For *Synechococcus*, STRAMSKI *et al.* measured an average c_c^* of 2.48, though it tended to be higher at dusk than at dawn. Our laboratory calibrations with *Nannochloris* and *Synechococcus* also indicate inter-species differences in c_c^* (see Fig. 3; for

Nannochloris, mean $c_c^* = 3.38$, sd = 0.34, n = 49; for *Synechococcus*, mean $c_c^* = 1.52$, sd = 0.26, n = 8). Using a lower estimate of c_c^* (such as that determined for *Nannochloris*) leads to a higher estimate of production than WALSH *et al.* obtained, but still considerably lower than estimates from carbon uptake. Applying a variable c_c^* which is higher at dusk than at dawn, reduces the estimate even more. However, applying a c_c^* typical for *Synechococcus* to the portion of the change in beam c for which the picoplankton were responsible does lead to slightly higher estimates of production (though still not as high as carbon uptake determinations). These considerations illustrate the necessity of knowing the phytoplankton community composition for interpreting diel variations in beam c. The beam c approach appears to underestimate production; one reason for this is illustrated by the 60 m data in Fig. 2. The bulk beam c pattern changes very little from dawn to dusk, yet variations in scattering cross section, presumably related to carbon growth, of all the different phytoplankton groups are evident. These increases during the day are, in effect, cancelled out by the decreasing cell concentrations (presumably due to grazing) to produce a nearly constant beam c due to the phytoplankton groups during the day (Fig. 6), and thus an unchanging $c_{\text{particles}}$. This implies that grazed particles are removed from the beam attenuation signal (as suggested by CULLEN *et al.*, 1992) and supports the contention that using diel variations in beam c to estimate production includes the effects of grazing (WALSH *et al.*, 1995).

CONCLUSIONS

We have shown that, over three intensive diel samplings and several pairs of dawn-dusk samplings during two cruises in the equatorial Pacific, diel variations in the bulk water optical property of beam attenuation (minimum near dawn and maximum near dusk) correspond to diel variations in the forward light scattering of phytoplankton populations, and not to cell concentration, which often exhibits the inverse diel pattern. These diel variations in light scattering of phytoplankton appear to be due primarily to cell growth as the cells photosynthesize and add carbon during the day, and then divide at night. The decrease in the bulk beam attenuation during the night is due to a combination of repackaging of cell material via cell division, aggregation, and grazing, as well as losses of material due to respiration and, mixing. By using laboratory and theoretically derived calibrations to analyze diel variations in phytoplankton populations at sea, we were able to account for essentially all of the diel variation in $c_{\text{particles}}$. The majority of the c_{phyto} signal was from eukaryotic phytoplankton (the nano- and ultra- phytoplankton), with a smaller portion from the most abundant cell, *Prochlorococcus*, and a negligible amount from *Synechococcus* in these waters. In most cases, the largest contributor to the diel variation in c_{phyto} was the ultraphytoplankton group. These findings emphasize the need to characterize the composition of the phytoplankton community in order to use beam attenuation as a tool for monitoring productivity.

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Chapter 4

Contributions of phytoplankton light scattering and cell concentration changes to diel variations in beam attenuation in the Sargasso Sea

ABSTRACT

The mean forward light scatter of a mixed population of nanophytoplankton and the picophytoplankton, *Prochlorococcus* and *Synechococcus*, measured on individual cells by flow cytometry, increased during the day and decreased during the night in the Sargasso Sea. This pattern was similar to that of bulk beam attenuation due to particles. In contrast, cell concentrations changes did not generally correspond to the patterns in beam attenuation.

Using a combination of empirical calibrations relating beam attenuation to flow cytometric measurements of pure cultures of phytoplankton in the laboratory, and Mie theory, we estimated the contributions of phytoplankton to the diel variations in beam attenuation observed during diel samplings in January 1992, July 1993, and May 1994. The bulk beam attenuation due to particles was strongly correlated with calculated beam attenuation due to phytoplankton. During the July 1993 diel sampling, when pico- and nanophytoplankton populations were analyzed, in most instances the nanophytoplankton were the largest contributor to total beam attenuation due to phytoplankton, but *Prochlorococcus* were equally important at 70 m for some time points over the diel cycle. In the upper 40 m, *Prochlorococcus* were a smaller contributor to beam attenuation changes than the nanophytoplankton, and *Synechococcus* were even less important. These findings emphasize the need to characterize the composition of the phytoplankton community in order to use beam attenuation as a tool for monitoring productivity.

INTRODUCTION

Diel variations in beam attenuation, with a minimum near dawn and a maximum near dusk, have been used to estimate primary productivity in the North Pacific (Siegel et al. 1989) and the equatorial Pacific (Cullen et al. 1992). In order to elucidate the relationship between phytoplankton and beam attenuation, we have related direct measurements of changes in the light scattered by the phytoplankton to changes in beam attenuation over the diel cycle in the Sargasso Sea in January 1992, July 1993, and May 1994. We observed diel variations in mean forward light scatter for populations of eukaryotic and prokaryotic phytoplankton that are minimum near dawn and maximum near dusk, the same patterns seen for bulk beam attenuation. In contrast, phytoplankton cell concentrations generally do not have the same diel patterns; instead, they usually have the opposite pattern. Using laboratory calibrations on phytoplankton combined with theoretical calculations, we estimated the contribution that each phytoplankton population made to the diel variations in beam attenuation in the Sargasso Sea. The complete explanation of the motivation behind the work and the calibrations used can be found in DuRand and Olson (in press [Chapter 3]), which uses the same method to analyze data from diel cycles in the equatorial Pacific.

MATERIALS AND METHODS

Field Studies

Diel samplings were performed on four occasions during three cruises in the Sargasso Sea. During R.V. *Endeavor* 232 in January 1992, two diel samplings were performed (EnD1 for 48-h at 27°N 68°W and EnD2 for 28-h at 35°N 68°W). During R.V. *Columbus Iselin* 93-06 (July 1993), one 40-h sampling was performed (Iselin at 35°N 69°W). During R.V. *Cape Hatteras* 0694 (May 1994), one 48-h sampling was performed (Hat at 35°N 71°W). For each sampling, a holey-sock drogue set at 25 m was launched and tracked in order to follow the same water mass during the sampling. CTD-transmissometer casts were made every two hours and samples for flow cytometric analyses were taken with Niskin bottles at four to six depths.

The eukaryotic phytoplankton were analyzed on board the ship within two hours of collection, using an EPICS V flow cytometer modified to analyze 50 ml seawater samples at 5-10 ml min⁻¹ (Olson et al. 1991, 1993). Samples were kept at room temperature in the dark until analysis. Fluorescent microspheres (3.79 µm, Polysciences) were added to each sample as an internal standard. Eukaryotic phytoplankton from 1-20 µm in diameter were counted and, for each particle, forward light scatter (FLS, 3°-19° at 488 nm), side scatter, red (chlorophyll, 660-700 nm) fluorescence, and orange (phycoerythrin, 530-630) fluorescence were collected. Phytoplankton populations were discriminated on plots of forward light scatter vs.

chlorophyll fluorescence using CytoPC software (D. Vaultot, Station Biologique, Roscoff, France). The population referred to here as the nanophytoplankton primarily consisted of 2-5 μm diameter coccoid cells, but also included cells as large as 20 μm .

For the Iselin diel sampling, the picoplankton *Synechococcus* and *Prochlorococcus* were also analyzed on board the ship within two hours of collection using a high sensitivity configuration of an EPICS 753 flow cytometer (Olson et al., 1993), and 0.57 μm beads (Polysciences) as an internal standard. *Synechococcus* were distinguished by their high phycoerythrin fluorescence and *Prochlorococcus* by their low FLS and chlorophyll fluorescence.

Laboratory Studies

Laboratory experiments were performed on the chlorophyte *Nannochloris* sp. grown on a 14 h light:10 h dark cycle and sampled every two hours for 24 hours. These experiments are described fully elsewhere (Chapter 2). Briefly, diel sampling experiments were performed at 250 (Nan250) and 500 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ (Nan500) in batch cultures in an incubator and at 1500 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ (Nan1500) outdoors under natural sunlight. Briefly, measurements were made of mean forward light scatter and cell concentration (flow cytometric analysis), beam attenuation (SeaTech 25-cm transmissometer), absorption (opal glass method, Shibata 1958, in a spectrophotometer), cell size (Coulter Multisizer), and carbon content (Perkin-Elmer 2400 CHN Analyzer).

A similar experiment was performed on *Synechococcus*, with 4 strains (WH8012, 8015, 8018, and 8103) grown at $75 \mu\text{mol photons m}^{-2} \text{ s}^{-1}$ on a 14:10 L:D cycle and sampled at "dawn" and "dusk." Measurements were made of FLS and cell concentration, beam attenuation, cell size, and carbon content.

Conversion of forward light scatter to scattering cross section

Our measurements of forward light scatter (3° - 19°) are expressed relative to light scattered by standard beads. To compare our results to theory, we need to express them in more meaningful units. We therefore calculated the scattering cross section (σ_b in μm^2) of the cells by reference to the calculated scattering cross section (3° - 19°) of the beads, for which the size, refractive index, and absorption are known (Bohren and Huffman, 1983; Ackleson and Spinrad, 1988). This procedure is fully described in DuRand and Olson (in press).

RESULTS AND DISCUSSION

Diel Patterns in Phytoplankton and Beam Attenuation

The mean scattering cross section (calculated from flow cytometric forward light scatter) of nanophytoplankton, *Synechococcus*, and *Prochlorococcus* cells was consistently at a minimum near dawn and maximum near dusk at all depths sampled during EnD1, Hat, and Iselin and at 0 and 20 m during EnD2 (Figs. 1B-3B, 4B, D, F). The patterns in beam attenuation due to particles ($c_{\text{particles}}$) were similar to those

Figure 1. Time series data for each depth sampled during a 48-hour diel sampling experiment in January 1992 at 27°N 68°W (R.V. *Endeavor*, EnD1). A) Beam attenuation due to particles ($c_{\text{particles}}$, m^{-1}), B) Forward light scattering cross section (σ_b , μm^2), and C) cell concentration (cells ml^{-1}) for nanophytoplankton.

EnD1

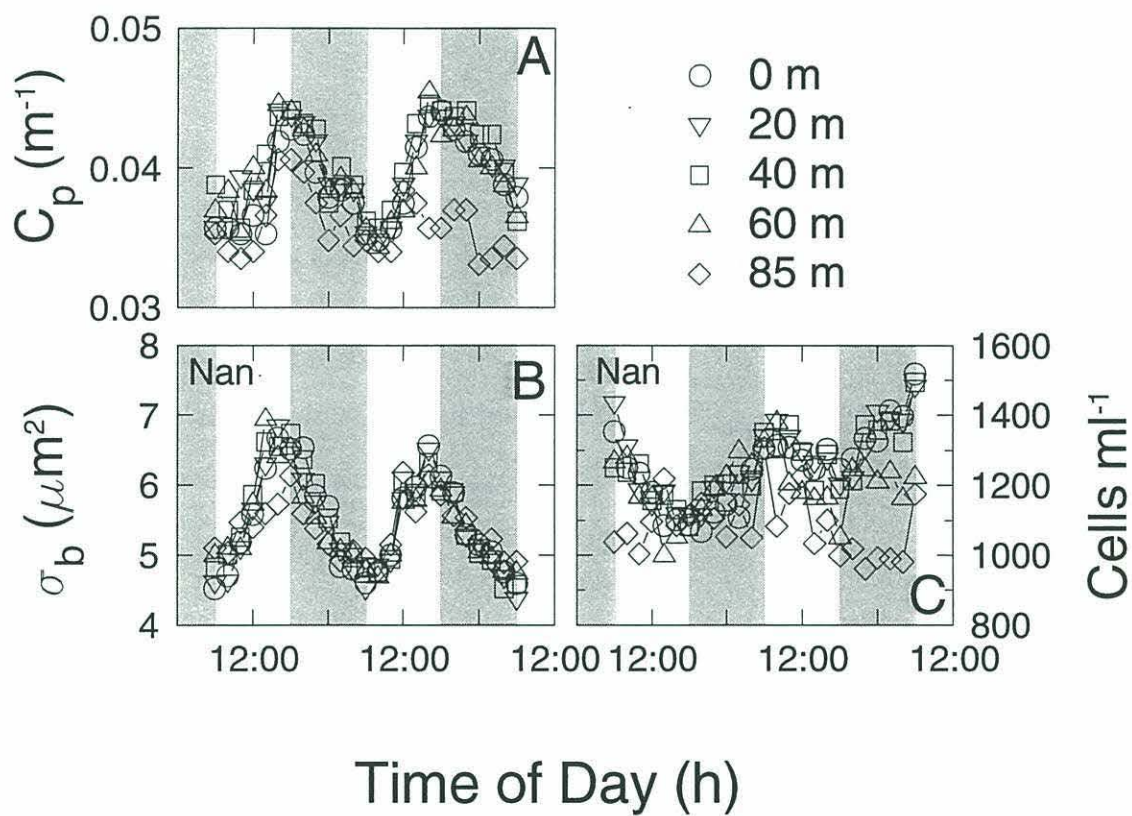


Figure 2. Time series data for depths to 100 m during a 28-hour diel sampling experiment in January 1992 at 35°N 68°W (R.V. *Endeavor*, EnD2). A) Beam attenuation due to particles ($c_{\text{particles}}$, m^{-1}), B) Forward light scattering cross section (σ_b , μm^2), and C) cell concentration (cells ml^{-1}) for nanophytoplankton.

EnD2

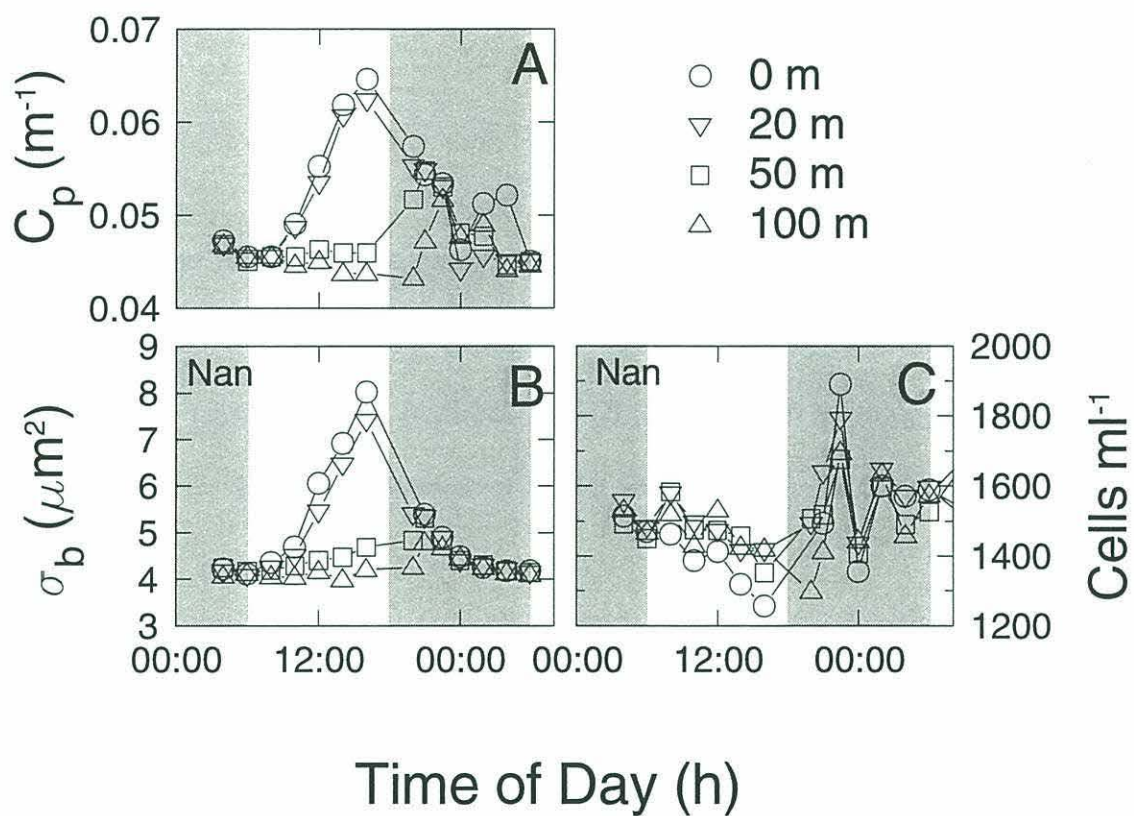


Figure 3. Time series data for all depths sampled during a 48-hour diel sampling experiment in May 1994 at 35°N 71°W (R.V. *Cape Hatteras*, Hat). A) Beam attenuation due to particles ($c_{\text{particles}}$, m^{-1}), B) Forward light scattering cross section (σ_b , μm^2), and C) cell concentration (cells ml^{-1}) for nanophytoplankton.

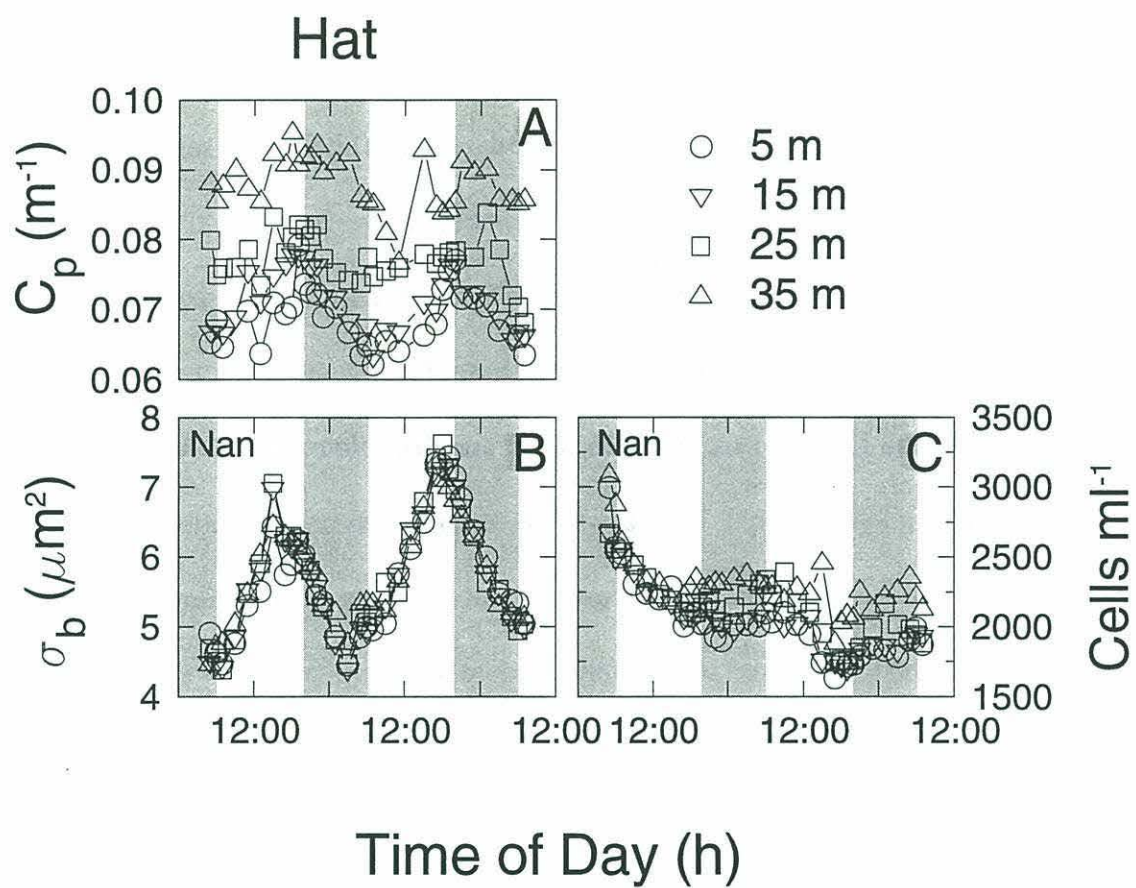
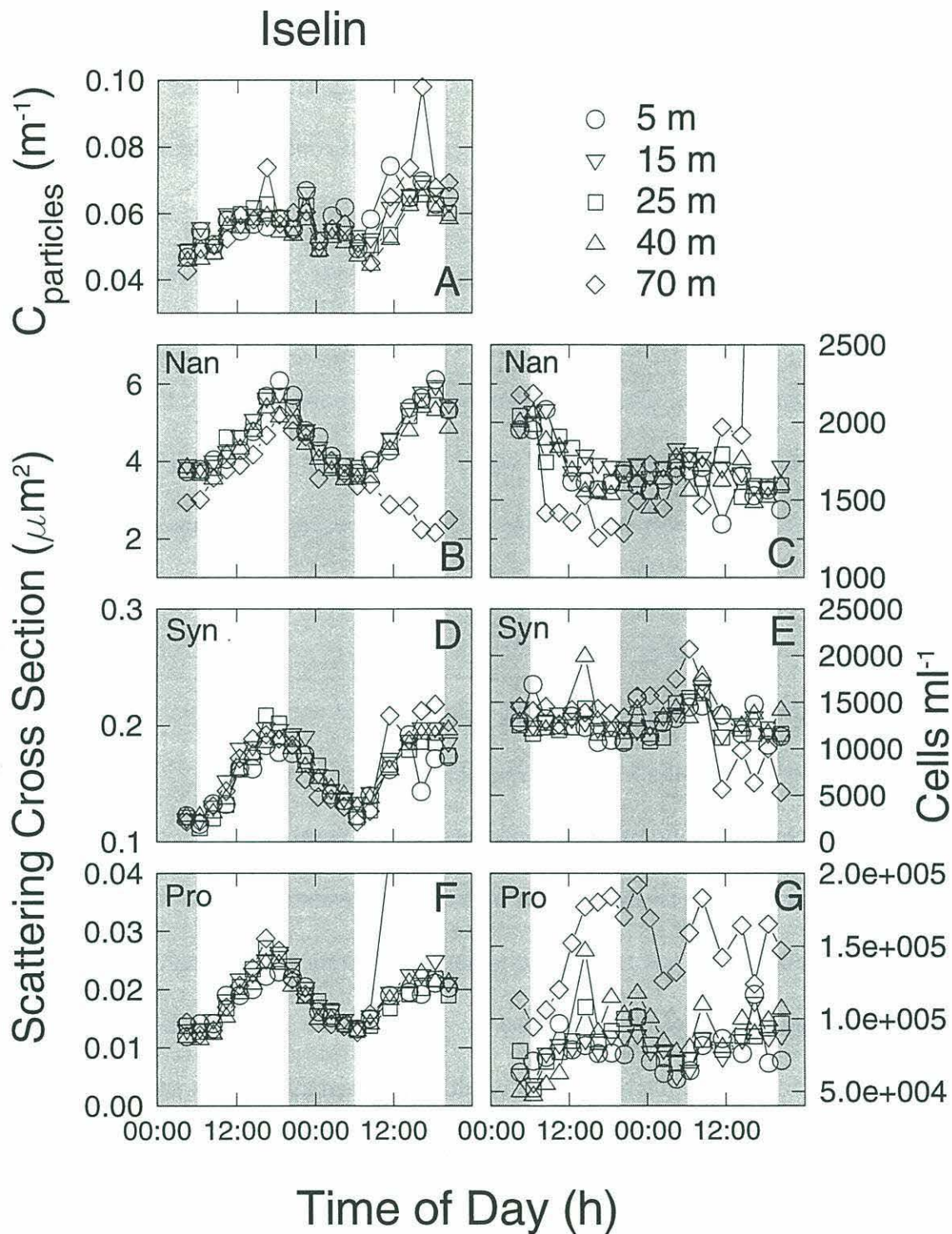


Figure 4. Time series data for all depths sampled during a 40-hour diel sampling experiment in July 1993 at 35°N 69°W (R.V. *Columbus Iselin*, Iselin). A) Beam attenuation due to particles ($c_{\text{particles}}$, m^{-1}). Forward light scattering cross section (μm^2), and cell concentration (cells ml^{-1}) for nanophytoplankton (B and C), *Synechococcus* (D and E), and *Prochlorococcus* (F and G). Note that the y-axis for each plot is scaled independently. The nanophytoplankton cell concentrations and the *Prochlorococcus* scattering cross sections are both off-scale at 70 m for the last few data points in the time series, presumably because we were no longer sampling the same water mass at that depth (the drogue was set at 25 m).



of FLS, with a minimum near dawn and a maximum near dusk (Figs. 1A-4A). Cell concentrations for the nanophytoplankton generally decreased during the day and increased during the night (Figs. 1C-3C, 4C, E, G). During the Iselin diel sampling, for which picoplankton were analyzed, *Synechococcus* cell concentrations were steady during the first day, increased during the night, and decreased the second day. *Prochlorococcus* cell concentrations exhibited a pattern never before observed in numerous diel sampling experiments in the Sargasso Sea and the equatorial Pacific (DuRand and Olson in press, Chapter 5, Appendix B; Dusenberry 1995); they increased during the day and decreased at night. However, Dusenberry did observe similar trends during laboratory experiments on *Prochlorococcus* in a mixostat. At the highest mixing rates (and thus the highest growth rates) tested, the cell concentrations increased during the day, either just before or concurrent with the increase in FLS during the day. Dusenberry suggested that nutrient limitation in these mixostats prevented cells from dividing until the light period.

It is clear that the patterns in the bulk property of beam attenuation are similar to the patterns in mean light scatter of different populations of phytoplankton, and they are generally different from those in cell concentration. The increase in light scatter during the day can be explained by growth of the phytoplankton cells as they photosynthesize and fix carbon. As a result of the increase in light scatter, the total beam c increases, despite the fact that cell concentrations are generally decreasing during the day (presumably due to grazing). The maximum in cell light scatter almost always occurs at the same time as the minimum in cell concentration,

near dusk. The implication is that the cells begin to divide at this time, so that the average cell size decreases, with a corresponding increase in the cell concentration (newly divided cells). A number of factors can contribute to the decrease in beam c during the night; these include cell division, respiration, mixing, cell aggregation, and grazing. These are discussed in detail in DuRand and Olson (in press [Chapter 3]).

Calculation of beam attenuation due to phytoplankton

To obtain estimates of beam c from field measurements of scattering cross sections and cell concentrations of phytoplankton populations, we began with an empirical calibration using *Nannochloris* and *Synechococcus* cultures. The calibrations used are fully described in DuRand and Olson (in press). Briefly, we determined a relationship between the attenuation cross section (σ_c , beam c normalized to cell concentration) and scattering cross section (σ_b , from flow cytometric FLS) from *Nannochloris* and *Synechococcus* cultures and found that it compared well to calculations based on Mie scattering theory. For *Prochlorococcus*, we obtained a relationship analogous to that used for the other populations by using theoretically-derived attenuation cross sections.

The appropriate calibration was applied to the field data of mean scattering cross section (σ_b) for different phytoplankton populations to obtain beam c per cell ml^{-1} (i.e., σ_c , attenuation cross section) for each population. This was multiplied by the cell concentration to obtain an estimate of beam attenuation (c , m^{-1}) due to each specific group of phytoplankton. The diel patterns in beam c due to the phytoplankton are qualitatively quite similar to that of $c_{\text{particles}}$ (Figs. 5-8).

Figure 5. Time series of bulk beam attenuation due to particles (A) and calculated beam attenuation due to nanophytoplankton (B) during a 48-hour diel sampling experiment in January 1992 at 27°N 68°W (R.V. *Endeavor*, EnD1).

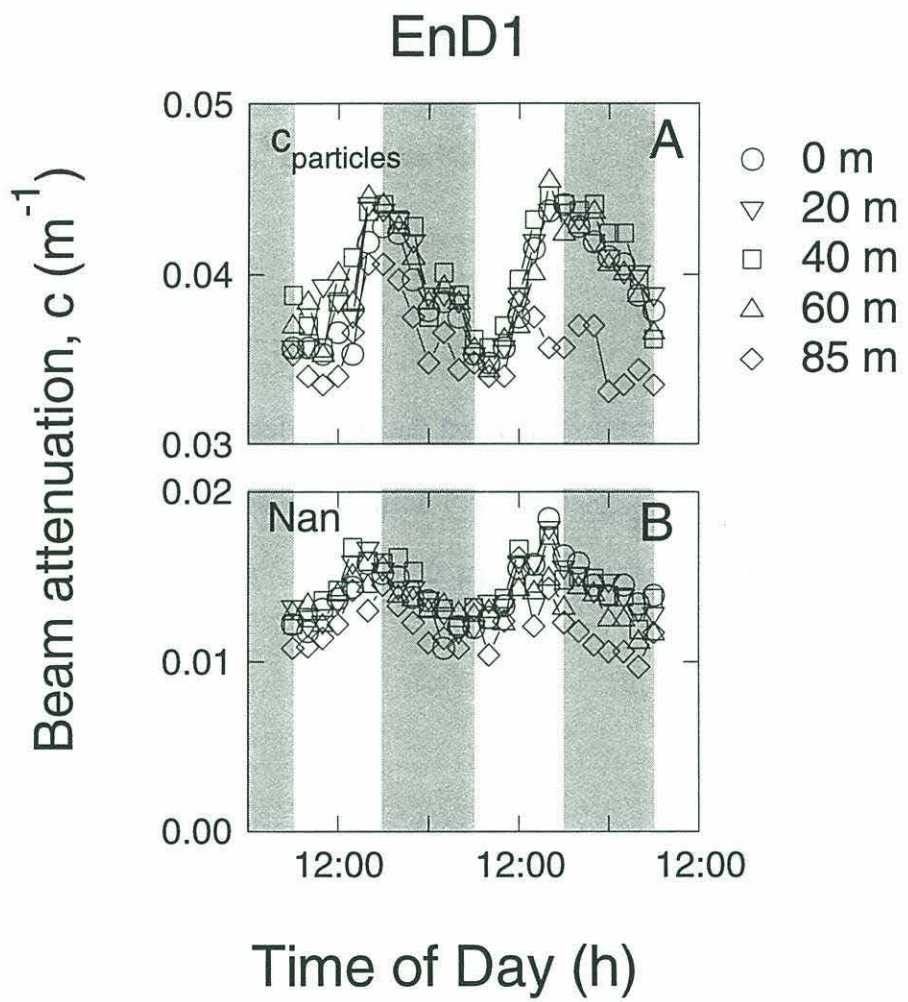


Figure 6. Time series of bulk beam attenuation due to particles (A) and calculated beam attenuation due to nanophytoplankton (B) during a 28-hour diel sampling experiment in January 1992 at 35°N 68°W (R.V. *Endeavor*, EnD2).

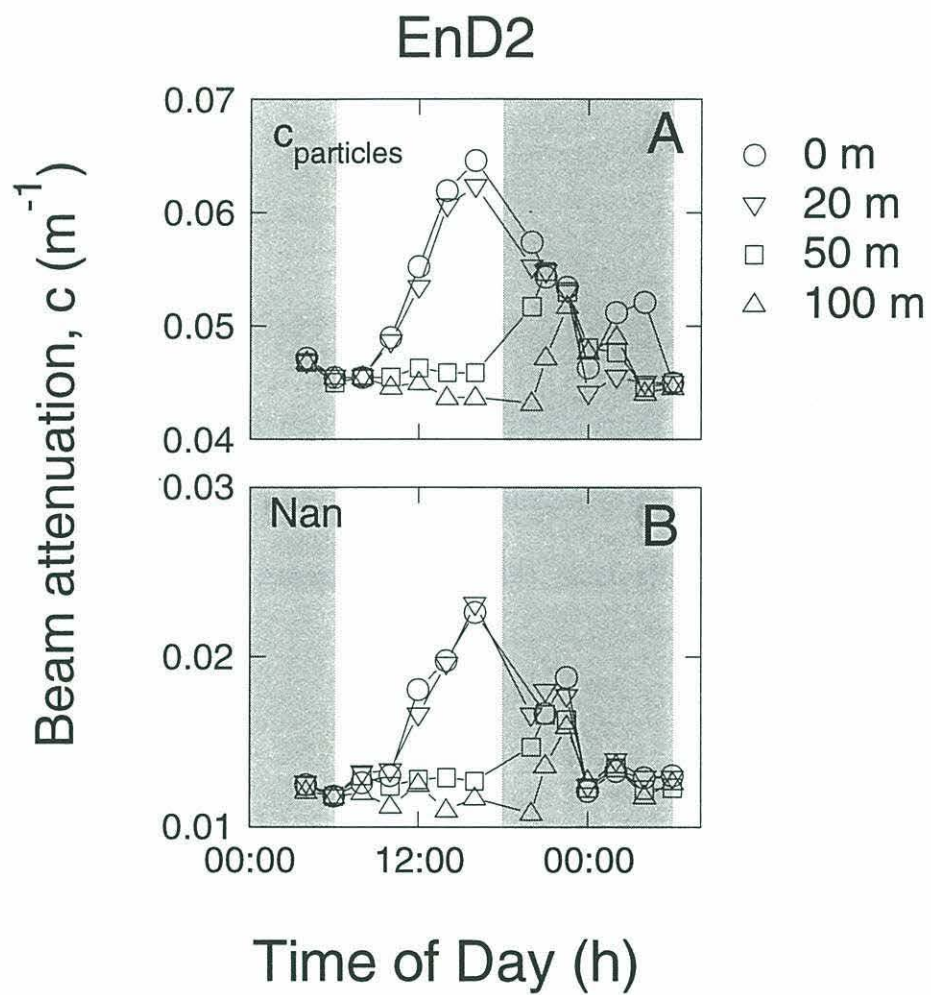


Figure 7. Time series of bulk beam attenuation due to particles (A) and calculated beam attenuation due to nanophytoplankton (B) during a 48-hour diel sampling experiment in May 1994 at 35°N 71°W (R.V. *Cape Hatteras*, Hat).

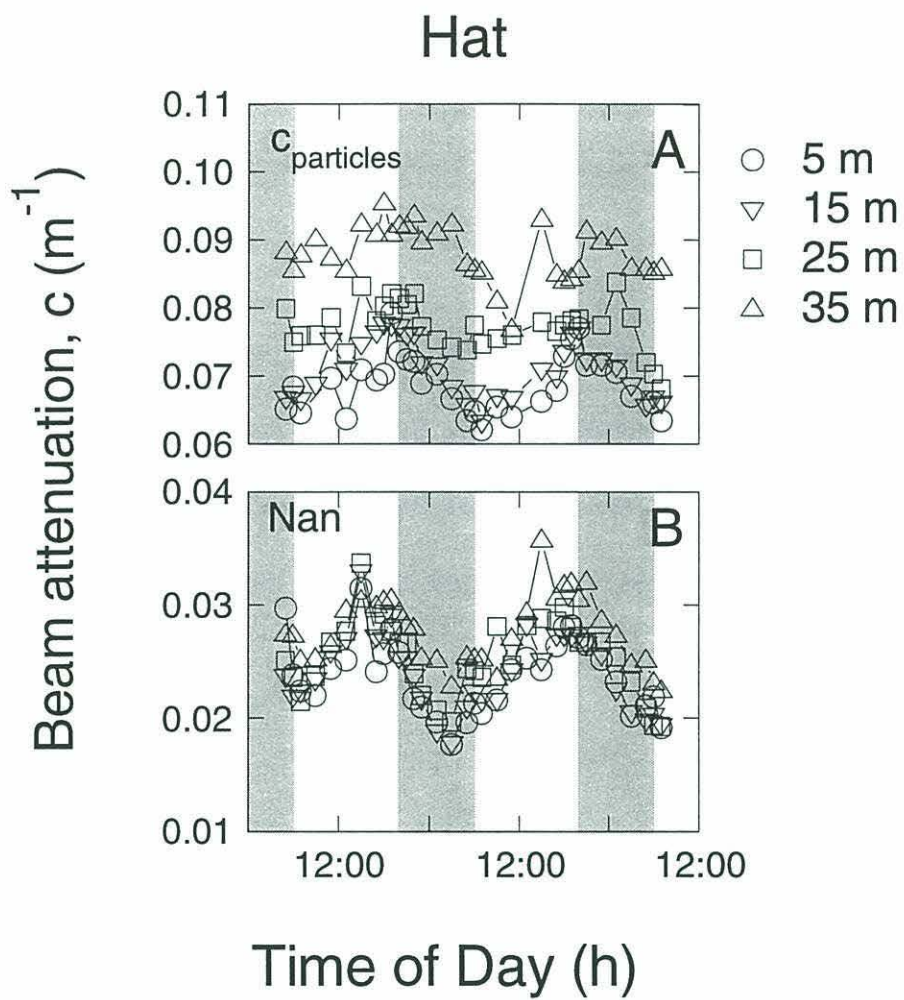
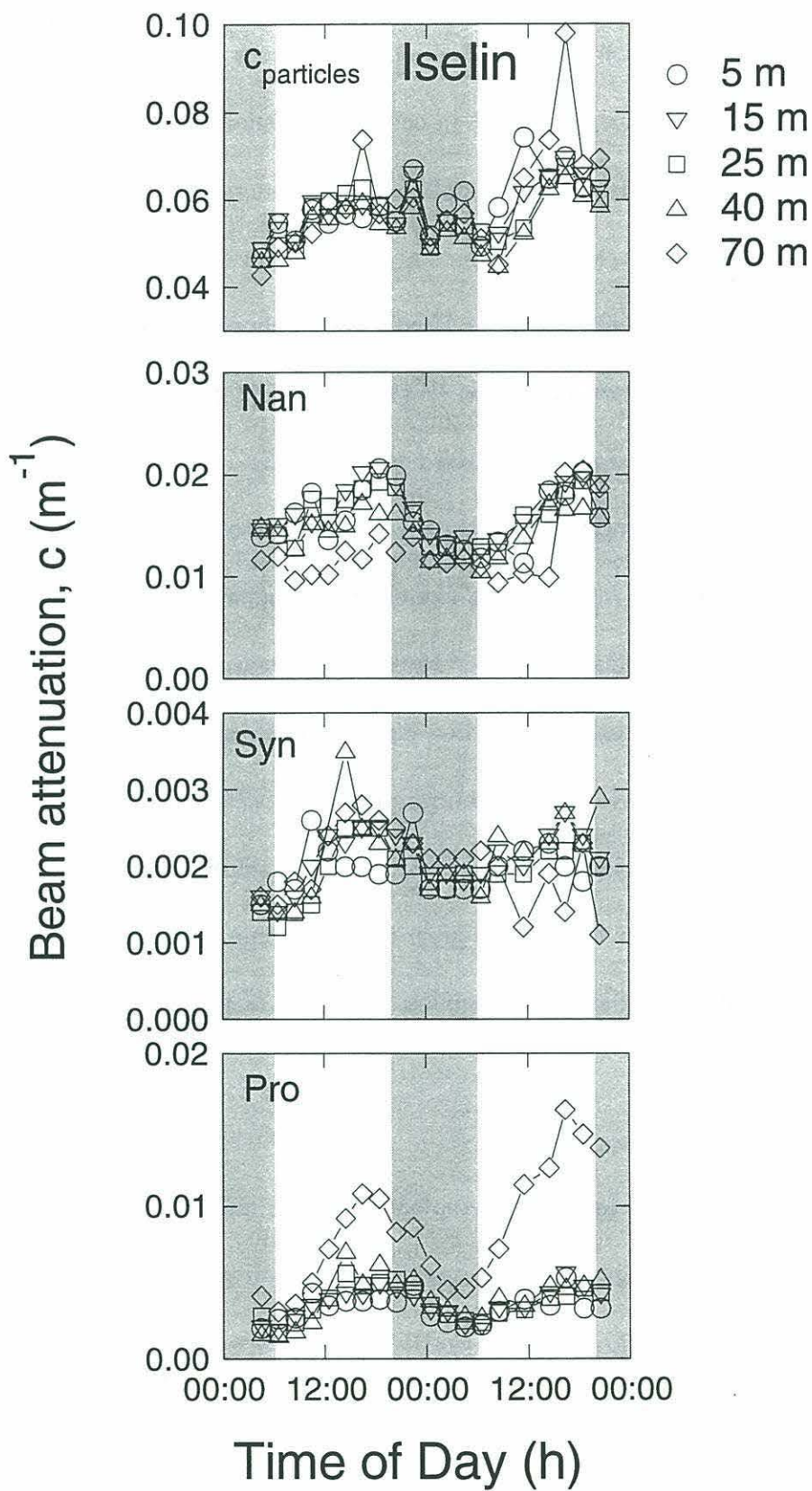


Figure 8. Time series of bulk beam attenuation due to particles (A) and calculated beam attenuation due to nanophytoplankton (B), *Synechococcus* (C), and *Prochlorococcus* (D) during a 40-hour diel sampling experiment in July 1993 at 35°N 69°W (R.V. *Columbus Iselin*, Iselin). Note that each plot is scaled independently in order to show the trends.



All tend to increase during the day and decrease during the night. For the most part, the variations in phytoplankton beam attenuation reflect those of the scattering cross section, dampened by the opposing trends in cell concentration. However, for *Prochlorococcus* during the Iselin diel sampling, the σ_b patterns are augmented by the cell concentration pattern, leading to greater variations in c_{pro} (Figs. 4, 8). For EnD2, the increase in c_{nan} which occurs at 50 and 100 m shortly after dark, is clearly caused by an increase in cell concentration (Figs. 2, 6). This increase in cell concentration was due to mixing, as discussed in Chapter 5.

For the Iselin diel sampling, the absolute magnitudes of the contributions of each phytoplankton group to beam c were examined to emphasize their relative contributions (Fig. 9, same data as Fig. 8). Most of c_{phyto} was due to the nanophytoplankton, with a smaller proportion due to *Prochlorococcus* and *Synechococcus*. At 70 m, however, the contribution of *Prochlorococcus* equals that of the nanophytoplankton at some time points during the sampling. Also, the diel variations in *Prochlorococcus* beam c are great and contribute more to the diel variation in c_p than does the nanophytoplankton beam c at that depth.

Comparison of $c_{particles}$ and c_{phyto}

Since the pattern in c for each phytoplankton group matched that of $c_{particles}$, we wished to see if the changes in $c_{particles}$ could be quantitatively accounted for by the changes in c_{phyto} ($c_{phyto} = c_{nan} [+ c_{Pro} + c_{Syn}]$). To do this, we examined the changes between successive samples in $c_{particles}$ ($\Delta c_{particles}$) and c_{phyto} (Δc_{phyto}). Thus,

positive values were obtained when c 's were increasing and negative values when decreasing. For EnD2, there was a strong correlation between $\Delta c_{\text{particles}}$ and Δc_{phyto} ($=\Delta c_{\text{nan}}$, in this case; $r^2 = 0.69$; Fig. 10). The slope of the relationship is 1.3, thus Δc_{nan} accounts for 78% ($1/1.3$, expressed as a percentage) of $\Delta c_{\text{particles}}$. For diel studies in the equatorial Pacific, we accounted for virtually all of the $\Delta c_{\text{particles}}$ with our calculated Δc_{phyto} (DuRand and Olson in press). However, the picoplankton, as well as the eukaryotic phytoplankton, were analyzed for those experiments. Here, we account for nearly 80% with the nanophytoplankton alone.

Though the general patterns in c_{phyto} are clearly similar to those in $c_{\text{particles}}$, for EnD1, Hat, and Iselin, there is enough variability in the data, that the trends do not match exactly. Thus $\Delta c_{\text{particles}}$ and Δc_{phyto} are not strongly correlated, as they are for EnD2, above. In order to reduce the jumpiness in the beam c data for these diel samplings, we smoothed the time series data with a three-point moving average.

There is some indication, at least during EnD1 and Hat, that c_{phyto} leads $c_{\text{particles}}$ by one or two time points, or about two hours (Fig. 11). It is possible that this is because the water samples were not analyzed at the exact same time the transmissometer data was taken. The Niskin bottles must be sampled and each sample analyzed with the flow cytometer. All samples are completed within two hours, however, if the cells, which are in bottles in the dark, continue to progress through the cell cycle, then this could account for the apparent time shift, since the data are plotted using the sampling (CTD) time and not the time of analysis. However, it is not clear why this would affect some diel samplings and not others (EnD2, for example). For EnD1,

Figure 9. Time series of calculated beam attenuation due to each group of phytoplankton (c_{nan} (\circ), c_{syn} (\square), and c_{pro} (\blacktriangledown)) plotted together to show their relative contributions during the Iselin diel sampling. Note that this is the same data as in the previous figure.

Beam attenuation, c (m^{-1})

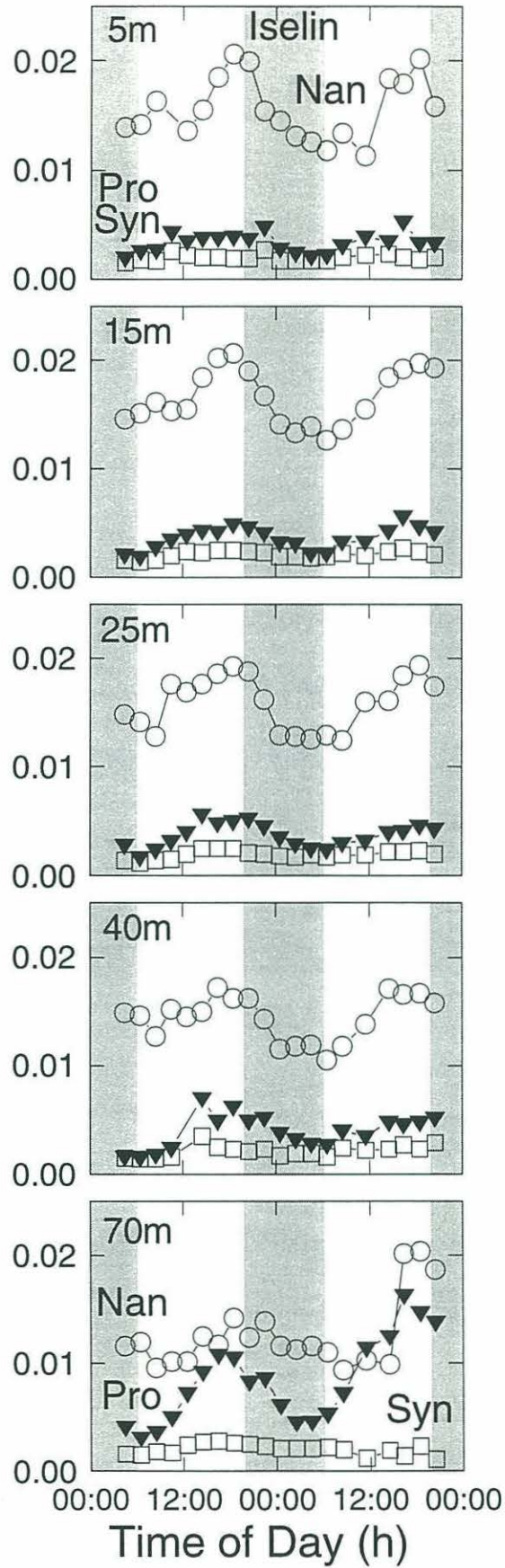


Figure 10. Relationship between $\Delta c_{\text{particles}}$ and $\Delta c_{\text{nanophytoplankton}}$ during a diel sampling experiment in January 1992 at 35°N 68°W (EnD2). Data are from 0-100 m. Solid line is linear regression ($r^2 = 0.69$, $n = 50$).

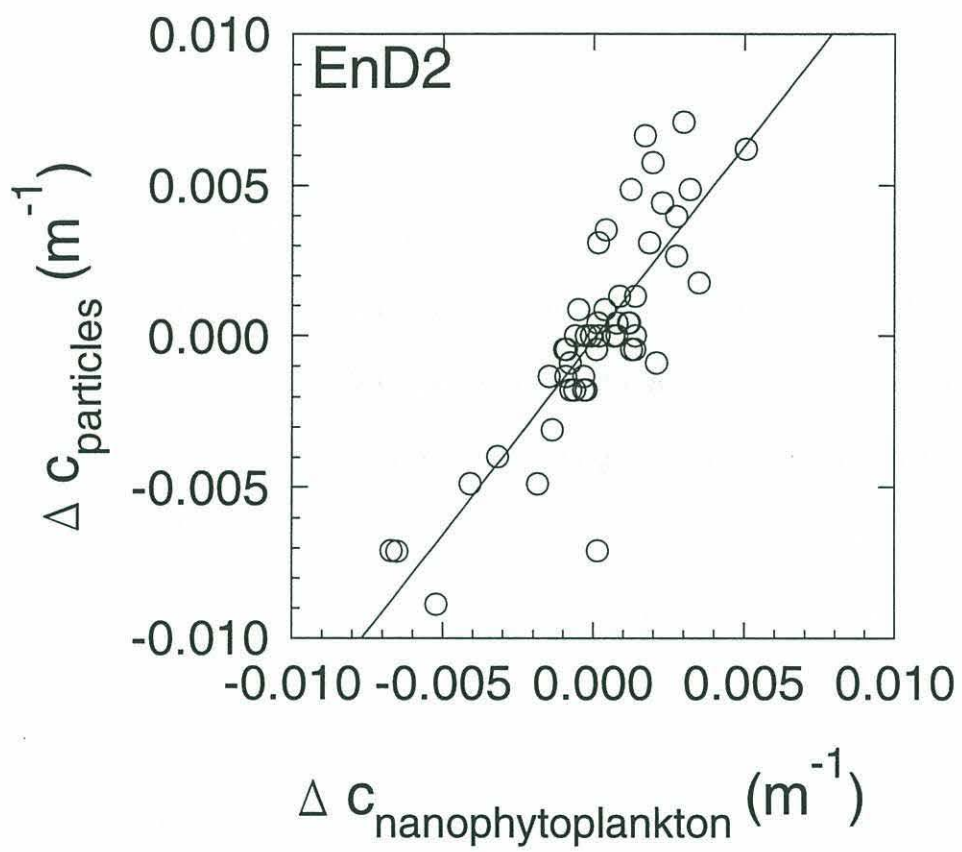
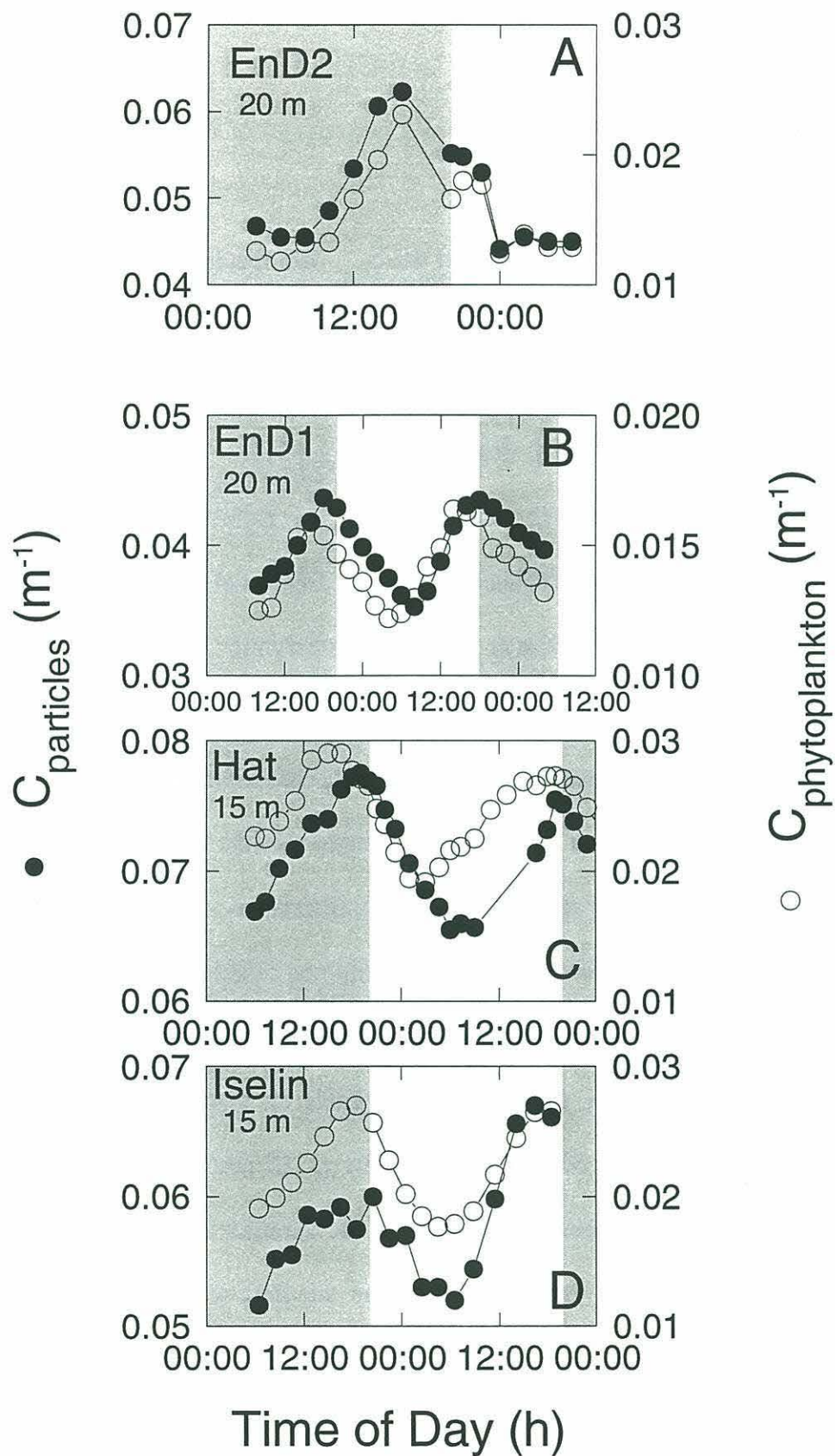


Figure 11. Comparison between $c_{\text{particles}}$ (●) and c_{phyto} (○) for A) EnD2 at 20 m, B) EnD1 at 20 m, C) Hat at 15 m, and D) Iselin at 15 m. Each axis is scaled independently in order to compare the trends. Data for EnD1, Hat, and Iselin have been smoothed with a three-point moving average. For EnD2, the patterns match well on a point-by-point basis. While the general trends match for EnD1, Hat, and Iselin, there are differences which can be seen.



the fit of $\Delta c_{\text{particles}}$ regressed on Δc_{phyto} is better if the Δc_{phyto} values are shifted by one time point ($r^2 = 0.62$ after shifting, instead of $r^2 = 0.45$). The slope of the regression is 1.53, indicating that 65% ($1/1.53$, expressed as a percentage) of the variability in $c_{\text{particles}}$ can be attributed to variations in c_{phyto} .

A regression of all the $c_{\text{particles}}$ data on all the c_{phyto} data results in a strong correlation ($r^2 = 0.82$; Fig. 12). It is clear, though, that each diel sampling forms a distinct group of points. The Endeavor diel samplings in January have the lowest c_p and the lowest c_{phyto} . For these two diel samplings, the c_{phyto} values are in the same range, though $c_{\text{particles}}$ is higher for EnD2 than for EnD1. The Hatteras diel sampling in May has the highest $c_{\text{particles}}$ and the highest c_{phyto} . The Iselin diel sampling in July, for which we measured the picoplankton as well as the eukaryotic nanophytoplankton, falls in the middle.

If we limit the data points to the upper waters (≤ 60 m, < 50 m for EnD2) where there are diel variations in c_p , then variation that we see (the range of the data on the plot) is due to changes over the day (and not to differences with depth). If we are accounting for all of the diel variations in c_p with our calculated c_{phyto} , then the slope should be equal to one. For Iselin, where we analyzed the eukaryotic nanoplankton and the picoplankton, the slope is 1.06 (Table 1); thus our calculated c_{phyto} accounts for nearly all of the observed variation in c_p . (If just C_{nan} is included, this raises the slope to 1.42, so approximately 70% of the variation is accounted for by the nanophytoplankton alone.) The average c_{phyto} for this data set is 37% of the average c_p ; thus the average $c_{\text{non-phyto}}$ ($c_p - c_{\text{phyto}}$) is 63% of c_p . For the other diel

samplings, where only the nanophytoplankton were analyzed, the slopes are higher (Table 1), and indicate that about 65% of the diel variations in c_p are accounted for by the nanophytoplankton. The average c_p can be partitioned into approximately 30% c_{phyto} and 70% $c_{\text{non-phyto}}$; however, for these samplings (EnD1, EnD2, and Hat), $c_{\text{non-phyto}}$ is a misnomer, since it includes the picophytoplankton (*Synechococcus* and *Prochlorococcus*) which were not analyzed.

It is interesting that EnD1 and EnD2 are well separated on the plot of c_p and c_{phyto} . EnD2 has a higher percent contribution of $c_{\text{non-phyto}}$ than does EnD1, indicating that there are more (or larger) non-phytoplankton particles (including bacteria, detritus, and microzooplankton) or picoplankton (since they were not analyzed for this sampling). Both diel samplings were performed in January, but EnD2 was north of Bermuda where there was a deep mixed layer (200 m) and 1 μM nitrate at the surface, whereas EnD1 was south of Bermuda where there was a 20-40 m surface mixed layer.

The calculations of beam attenuation due to phytoplankton presented here for diel sampling experiments in the Sargasso Sea can be compared to those for the equatorial Pacific (DuRand and Olson in press). The average beam attenuation due to particles tends to be higher in the equatorial Pacific than in the Sargasso Sea, except for the May 1994 (Hat) sampling (Table 1). The average contribution of beam attenuation due to phytoplankton to total c_p ranges from 37-58% for diel samplings where both pico- and nanophytoplankton were analyzed, with the lowest for the Sargasso Sea sampling (Iselin).

Figure 12. Relationship between beam attenuation due to particles ($c_{\text{particles}}$) and beam attenuation due to phytoplankton (c_{phyto}) for four diel samplings in the Sargasso Sea: EnD2 in January (\circ), EnD1 in January (∇), Hat in May (\triangle), and Iselin in July (\blacklozenge). A linear regression through all the data has an r^2 of 0.82.

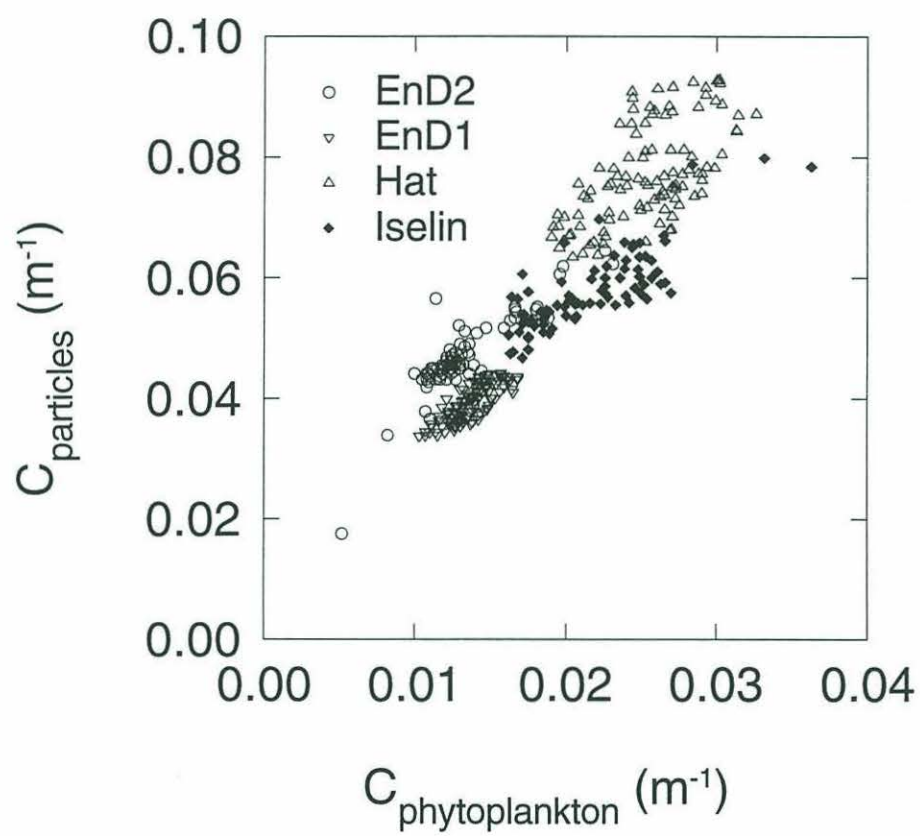


Table 1. Results of regressions of beam attenuation due to particles on the calculated beam attenuation due to the phytoplankton measured using flow cytometry (Fig. 12). For each diel sampling experiment, the following are listed: the phytoplankton populations measured with flow cytometry, the slope and fit of the regression, the average beam attenuation due to particles (m^{-1}), the average beam attenuation due to phytoplankton (expressed as percentage of c_p), and the average beam attenuation due to non-phytoplankton (expressed as percentage of c_p). Data for the Sargasso Sea sampling experiments, discussed in this chapter, and the equatorial Pacific sampling experiments (TT8D1, TT12D1, TT12D2; DuRand and Olson, in press) are compared. Asterisks indicate sampling experiments where the entire phytoplankton assemblage (<20 μm diameter) was measured.

Diel	Phyto	Slope	r^2	Avg c_p	% Avg c_{phyto}	% Avg $c_{non-phyto}$
Iselin	NanSynPro	1.06	0.48	0.057	37% *	63% *
Iselin	Nan only	1.42	0.43			
EnD1	Nan	1.51	0.52	0.040	35	65
EnD2	Nan	1.52	0.74	0.051	29	71
Hat	Nan	1.54	0.38	0.077	32	68
TT8D1	NanUltSynPro	1.09	0.62	0.066	58 *	42 *
TT12D1	NanUltSynPro	1.10	0.95	0.092	45 *	55 *
TT12D2	NanUltSynPro	1.19	0.82	0.125	42 *	58 *

CONCLUSIONS

We have shown that, during a number of diel samplings in the Sargasso Sea, diel variations in the bulk water optical property of beam attenuation (minimum near dawn and maximum near dusk) correspond to diel variations in the forward light scattering of phytoplankton populations, and not to cell concentration, which often exhibits the inverse diel pattern. These diel variations in light scattering of phytoplankton appear to be due primarily to cell growth as the cells photosynthesize and add carbon during the day, and then divide at night. The decrease in the bulk beam attenuation during the night is due to a combination of repackaging of cell material via cell division, aggregation, and grazing, as well as losses of material due to respiration and mixing. By using laboratory and theoretically derived calibrations to analyze diel variations in phytoplankton populations at sea, we were able to determine the phytoplankton contribution to beam attenuation. Beam attenuation due to particles was strongly correlated with calculated beam attenuation due to phytoplankton. During a diel sampling in July 1993, when multiple phytoplankton populations were analyzed, the majority of the c_{phyto} signal was from nanophytoplankton, with a smaller portion from the most abundant cell, *Prochlorococcus*, and even less from *Synechococcus*. At the deepest depth analyzed (70m), the *Prochlorococcus* contribution was equal to that of the nanophytoplankton at some times during the diel cycle; the *Prochlorococcus* contribution to the variations in beam attenuation was greater at that depth. Generally, the largest

contributor to the diel variation in c_{phyto} was the nanophytoplankton group. These findings emphasize the need to characterize the composition of the phytoplankton community in order to use beam attenuation as a tool for monitoring productivity.

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Chapter 5

**Analysis of phytoplankton growth in the equatorial Pacific
and the Sargasso Sea by flow cytometric measurements
of light scatter over diel cycles**

ABSTRACT

Flow cytometric measurements of phytoplankton light scattering and cell concentration over the diel light cycle were used to estimate *in situ* phytoplankton group-specific growth and loss rates in the equatorial Pacific and the Sargasso Sea. Measurements of forward light scatter were converted to cell volume and carbon using laboratory and theoretically derived calibration factors for specific groups of phytoplankton, including *Prochlorococcus*, *Synechococcus*, ultraphytoplankton, nanophytoplankton, and coccolithophores. Assuming that division was phased, specific growth rates were estimated based on volume and carbon changes between minimum and maximum values over the day. Phytoplankton group-specific loss rates, and also separate day and night loss rates, were estimated from the calculated growth rates and measured cell concentrations over time. The method used to estimate growth rates works well for *Prochlorococcus* and appears to work for small eukaryotic phytoplankton, but leads to underestimates for *Synechococcus* and larger eukaryotes. Estimated growth rates for *Prochlorococcus* reached one division per day in the upper waters of the equatorial Pacific, but were about half that in the Sargasso Sea. For the eukaryotic phytoplankton, growth rates in the equatorial Pacific were near one division per day in the upper waters; in the Sargasso Sea, the growth rates approached that, but were more often lower. In general, phytoplankton growth rates were closely matched by *in situ* loss rates over the course of a day.

INTRODUCTION

It has long been a goal in biological oceanography to measure *in situ* phytoplankton growth rates as a measure of primary productivity. Approaches to this problem include cell cycle markers, biochemical rate measurements, and cage and bottle incubations (reviewed by Furnas 1990). Some work has been done to make growth rate measurements for specific groups of phytoplankton such as *Prochlorococcus* (Goericke and Welschmeyer 1993, Vaulot et al. 1995, Liu et al. 1995), *Synechococcus* (Campbell and Carpenter 1986, Iturriaga and Marra 1988, Liu et al. 1995), and the dinoflagellate *Ceratium* (Weiler 1980).

It is particularly interesting to study high-nutrient, low-chlorophyll (HNLC) areas of the ocean, such as the equatorial Pacific, where a number of mechanisms have been invoked to explain the maintenance of an area with low phytoplankton biomass, yet excess nitrate. Hypotheses include control by grazing (Frost 1991), limitation by iron (Martin et al. 1989, 1991), and a combination of the two (the ecumenical hypothesis, Morel et al. 1991). HNLC areas can be compared with the oligotrophic Sargasso Sea, where phytoplankton biomass and nutrient levels are generally low and a seasonal cycle exists.

The equatorial Pacific is dominated by small, solitary phytoplankton less than 5- μm in diameter (Chavez 1989) consisting of *Synechococcus*, red fluorescing picoplankton (including prochlorophytes and eukaryotes such as *Micromonas*, *Chlorella*, and *Nannochloris*), prymnesiophytes, and pennate diatoms (Chavez et al.

1990). Cullen et al. (1992) estimated phytoplankton growth rates in the equatorial Pacific to be 0.6 d^{-1} or higher and concluded that phytoplankton population levels were closely controlled by grazing. In another study, growth rates estimated by both phytoplankton carbon from microscopy and ^{14}C uptake were close to 0.7 d^{-1} , or one doubling per day, and grazing rates estimated from dilution experiments were about 0.5 d^{-1} (Chavez et al. 1991). Vaultot et al. (1995) calculated *Prochlorococcus* growth rates from cell cycle analysis of DNA distributions and found them to be growing at a rate close to one doubling per day.

The Sargasso Sea north of Bermuda undergoes a seasonal variation in the phytoplankton communities, primarily due to the seasonal cycle which brings nutrients to the euphotic zone from deep mixing in the winter followed by restratification in the early spring (Siegel et al. 1990, Michaels et al. 1994).

Prochlorococcus and *Synechococcus* distributions have been examined (Olson et al. 1990a, b) and growth rate estimates for *Prochlorococcus* in the surface waters of 0.3 d^{-1} (Goericke and Welschmeyer 1993) and for *Synechococcus* of $0.42\text{-}0.86 \text{ d}^{-1}$ (Campbell and Carpenter 1986) and $0.5\text{-}1.2 \text{ d}^{-1}$ (Iturriaga and Marra 1988) reported.

From analysis of light scattering by individual phytoplankton cells in a variety of marine environments, we have seen dramatic diel patterns, which are presumably the results of cell growth during the day followed by division at night (Olson et al. 1990, 1991; DuRand and Olson in press). With calibrations using pure cultures in the laboratory, we can convert the diel variations in forward light scatter to cell volume and to cell carbon and thus estimate *in situ* growth rates of specific groups

within the phytoplankton. With data on cell concentrations over time, we can also estimate phytoplankton loss rates over the course of the diel experiments. Here we estimate *in situ* growth and loss rates in the equatorial Pacific and the Sargasso Sea for phytoplankton populations which can be distinguished using flow cytometry, including *Prochlorococcus*, *Synechococcus*, ultraphytoplankton, nanophytoplankton, and coccolithophores. This method is particularly attractive because it does not require incubation or manipulation (such as labeling) of cells, and could thus be performed using an *in situ* flow cytometer.

MATERIALS AND METHODS

Field Studies

Equatorial Pacific - Diel sampling was performed on three occasions during the two JGOFS Time Series cruises in the equatorial Pacific in 1992 (at 0°N, 140° W; Table 1). During R.V. *Thomas Thompson* 008, one diel sampling was performed (TT8D1 on 1 April) and during cruise 012, two diel samplings were performed (TT12D1 on 5 October and TT12D2 on 11 October). In each case, CTD casts were made every three hours for 24 hours and samples for flow cytometric analysis were taken with Niskin bottles at 0, 15, 30, 45, 60, 75, 90, and 100 m.

Sargasso Sea - Diel samplings were performed on four occasions during three cruises in the Sargasso Sea (Table 1). During R.V. *Endeavor* 232 in January 1992, two diel samplings were performed (EnD1 for 48-h at 27°N 68°W and EnD2 for

Table 1. Summary of diel sampling experiments in the equatorial Pacific and the Sargasso Sea.

Expt.	Cruise	Date	Location	Sampling
TT8D1	Thomas Thompson 008	1 April 92	0°N 140°W	Diel 1 24 hours
TT12D1	Thomas Thompson 008	5 Oct. 92	0°N 140°W	Diel 1 24 hours
TT12D2	Thomas Thompson 012	11 Oct. 92	0°N 140°W	Diel 2 24 hours
EnD1	Endeavor 232	20-21 Jan. 92	27°N 68°W (drogue)	Diel 1 48 hours
EnD2	Endeavor 232	29 Jan. 92	35°N 68°W (drogue)	Diel 2 28 hours
Iselin	Iselin 93-06	6-7 July 93	35°N 69°W (drogue)	40 hours
Hat	Hatteras 0694	24-25 May 94	35°N 71°W (drogue)	48 hours

28-h at 35°N 68°W). During R.V. *Columbus Iselin* 93-06 (July 1993), one 40-h sampling was performed (Iselin at 35°N 69°W). During R.V. *Cape Hatteras* 0694 (May 1994), one 48-h sampling was performed (Hat at 35°N 71°W). For each sampling, a holey-sock drogue set at 25 m was launched and tracked in order to follow the same water mass during the sampling. CTD casts were made every two hours and samples for flow cytometric analyses were taken with Niskin bottles at four to six depths.

Flow cytometry

Eukaryotic phytoplankton - An EPICS flow cytometer modified to run 5-10 ml min⁻¹ (Olson et al. 1991, 1993) was used to analyze 50 ml seawater samples live during the cruises. Eukaryotic phytoplankton from 1-20 µm in diameter were counted and, for each particle, forward light scatter (FLS, 3°-19°), side scatter (LS90, 73°-107°), red or chlorophyll fluorescence (660-700 nm), and orange or phycoerythrin fluorescence (530-630 nm) were collected. For the equatorial Pacific diel samplings (TT8D1, TT12D1, and TT12D2), two populations of eukaryotic phytoplankton were discriminated: ultraphytoplankton (1-2 µm cells), and nanophytoplankton (mostly 2-3 µm cells, but including cells up to 20 µm; DuRand and Olson in press, Fig. 1). For EnD1 and Iselin diel samplings, only one population of eukaryotic phytoplankton could be discriminated (nanophytoplankton). For EnD2, a population of coccolithophores was also distinguished (Olson et al. 1989).

Prokaryotic phytoplankton - The prokaryotic picoplankton, *Synechococcus* and *Prochlorococcus*, were analyzed for TT8D1, TT12D1, and TT12D2 and for Iselin diel samplings using an EPICS flow cytometer configured for high-sensitivity operation (Olson et al. 1993). On the TT cruises, the samples were preserved and stored until analysis on the flow cytometer in the laboratory on shore (Vaulot et al. 1989, Olson et al. 1993). For the Iselin cruise, the samples were analyzed live on the ship.

Laboratory experiments

Laboratory experiments were performed on the chlorophyte *Nannochloris* sp. grown on a 14 h light:10 h dark cycle and sampled every two hours for 24 hours. These experiments are described fully elsewhere (Chapter 2). Briefly, diel sampling experiments were performed at a range of light levels: at 250 (Nan250) and 500 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ (Nan500) in batch cultures in an incubator, at 60, 120, and 330 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ (Nan60, 120, and 330) in cyclostats in an incubator, and at 1500 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ (Nan1500) outdoors under natural sunlight. Measurements were made of cell size and concentration, light attenuation and absorption, flow cytometric light scattering (in forward and side directions), and carbon content (for Nan60, 120, and 330 only cell size, concentration, and light scattering were determined). In addition, the refractive index was calculated for Nan250, 500, and 1500.

Calculation of growth and loss rates

Mean forward light scatter for populations of phytoplankton in the field was converted to cell volume or carbon using laboratory calibrations (for eukaryotic phytoplankton) and theory (for prokaryotic picoplankton). To calculate growth rates, the model used here assumes that, for a given cell, volume or carbon doubles before division takes place, and that cell division is phased to the light:dark cycle. Thus an observation of a doubling of mean cell volume during the day would indicate that all the cells were dividing (a specific growth rate, μ , of 0.69 d^{-1} , and a division rate, k , of one doubling per day). However, if the phytoplankton cells of interest are only

partially phased, then the model used here would lead to underestimates of growth rate. If there is a fraction of cells which are not phased in their division, then the minimum near dawn would contain a number of doublet cells. There could still be a peak in FLS during the diel cycle if only a portion of the population is phased; however, we would underestimate the true growth rate using the method described here. This method would also not be expected to work for diatoms, which divide at all times during the diel light cycle (Chisholm 1981).

The calculated mean cell volume and carbon over the diel cycle were used to estimate specific growth rates (μ) from the increase in volume or carbon during the day (from the minimum near dawn to the maximum near dusk) and from the decrease in volume or carbon during the night (from the maximum near dawn to the minimum near the following dawn). Thus the equation used to determine specific

growth rate (d^{-1}) is:
$$\mu = \ln \frac{\text{maximum volume or carbon}}{\text{minimum volume or carbon}}$$

From the cell number changes from dawn of one day to dawn of the next day, we calculated net growth rate (d^{-1}). Phytoplankton loss rates over the day were calculated from the difference between this net growth rate and the specific growth rate. Separate day and night loss rates were also calculated. To calculate the daytime loss rates, we assumed that no cell division occurs during the daytime hours, thus the decrease in cell concentration is a true estimate of cell losses. The increase in cell concentration during the night is the result of cell division offset by cell losses. The number of cells expected in the absence of cell losses was calculated

from the growth rate. The difference between the expected number of cells at the end of cell division (near dawn) and the actual number of cells at that time point gives the number of cells lost during the night, which is then expressed as a loss rate (h^{-1}).

RESULTS AND DISCUSSION

Diel variations in cell light scatter and concentration

In the field, the mean forward light scatter (FLS) of the distinguishable phytoplankton populations consistently was minimal near dawn and maximal near dusk (often two hours before dusk; data from TT8D1 and Iselin in Figs. 1-3, see Appendix B for data from other diel samplings). The cell concentrations generally decreased during the day and increased at night (except for *Prochlorococcus* during the Iselin diel, which increased during the day and decreased at night; Figs. 1-3; Appendix B). We interpret the increase in scatter during the day as cell growth and the decrease during the night as the result of cell division. The maximum in cell light scatter generally occurred at the same time point as the minimum in cell concentration. The cell concentrations decrease during the day due to grazing and other cell losses, and then begin to increase when the cells divide at night. The increase seen in cell concentration during the night is dampened by cell losses from grazing, mixing, and aggregation (if aggregates are formed which are larger than the cells measured by the flow cytometer). We have generally seen that the magnitude of the forward light scatter increase is dampened with depth.

Figure 1. Time series of *Prochlorococcus* A) forward light scatter (relative to 3.79 μm bead) and B) cell concentration, and *Synechococcus* C) forward light scatter and D) cell concentration in the equatorial Pacific in April 1992 (TT8D1).

Forward Light Scatter

Cells ml⁻¹

TT8D1

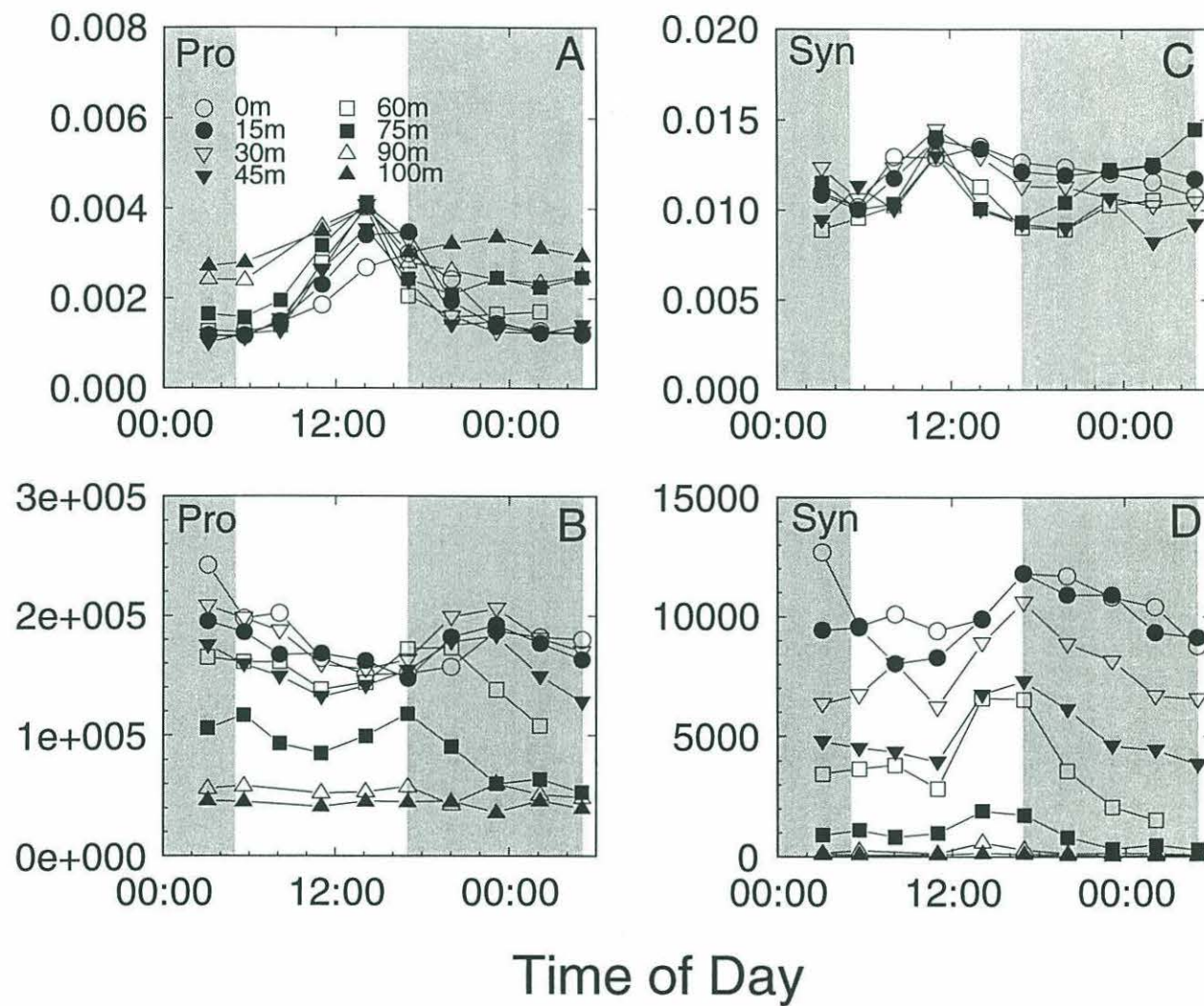
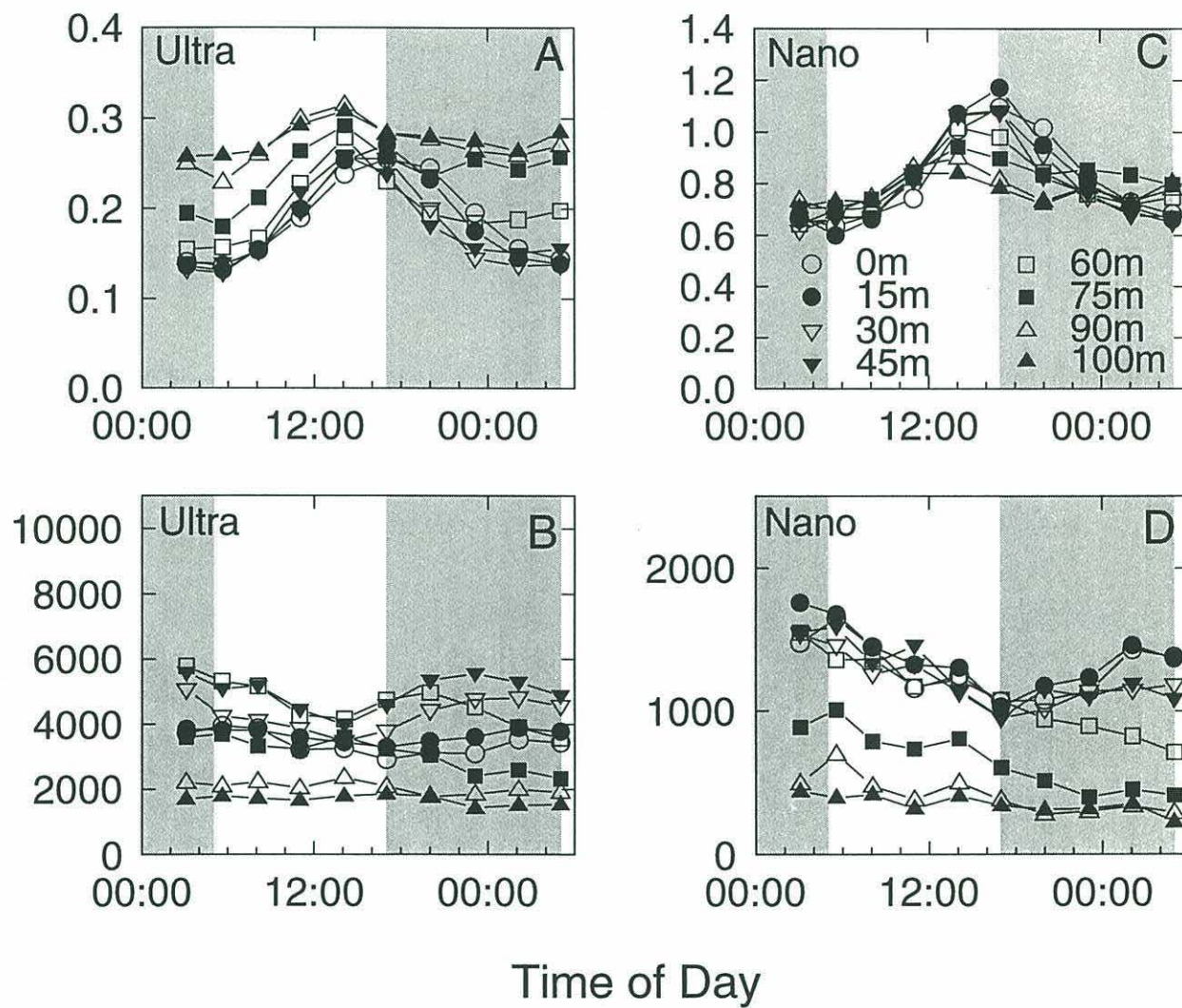


Figure 2. Time series of ultraphytoplankton A) forward light scatter (relative to 3.79 μm bead) and B) cell concentration, and nanophytoplankton C) forward light scatter and D) cell concentration in the equatorial Pacific in April 1992 (TT8D1).

Forward Light Scatter

Cells ml⁻¹

TT8D1



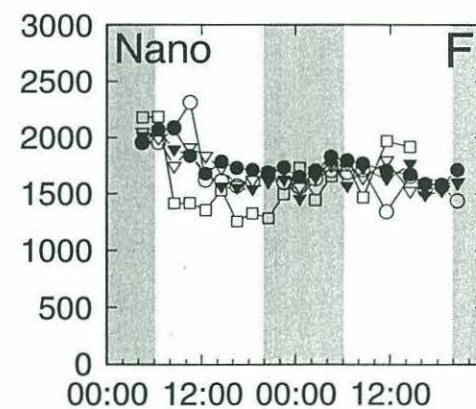
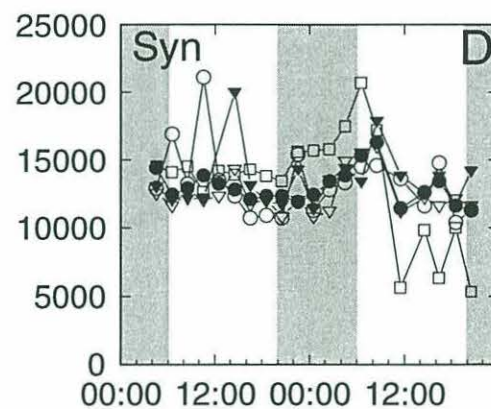
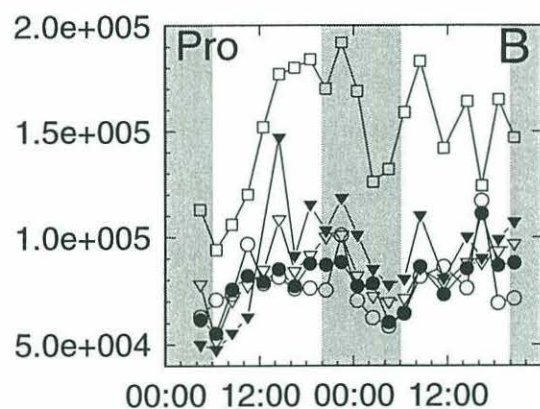
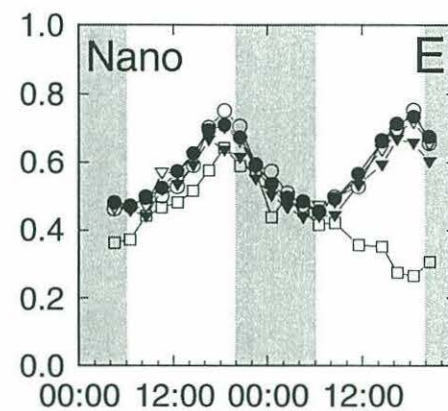
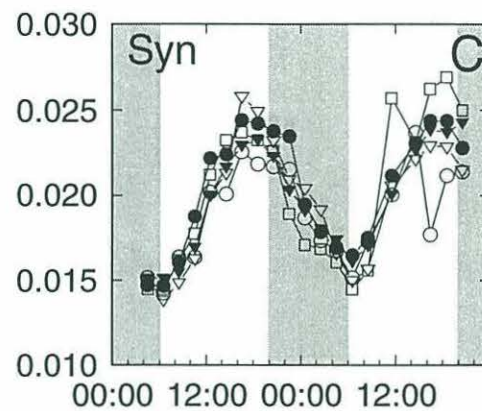
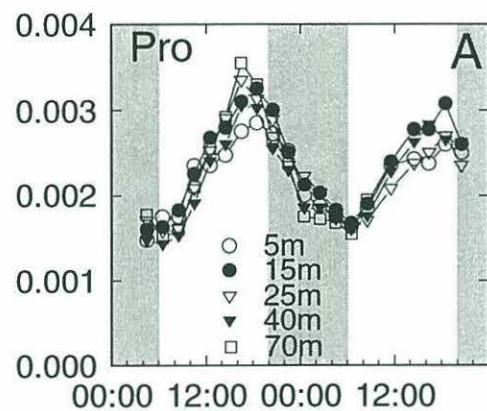
Time of Day

Figure 3. Time series of *Prochlorococcus* A) forward light scatter (relative to 3.79 μm bead) and B) cell concentration, *Synechococcus* C) forward light scatter and D) cell concentration, and nanophytoplankton E) forward light scatter and F) cell concentration during a 40-hour diel sampling experiment in July 1993 at 35°N 69°W (R.V. *Columbus Iselin*, Iselin).

Forward Light Scatter

Cells ml⁻¹

Iselin



Time of Day

During the equatorial Pacific cruises, we saw that the maximum forward light scatter (FLS) for *Prochlorococcus* occurred at the time point closest to dusk (near 1700 h) in the surface waters (Fig. 1A; 0 and 15 m for TT8D1; 0, 15, and 30 m for TT12D1; 0 m for TT12D2), but at the preceding time point (1400-1500 h) for the deeper depths (down to 100 m for all three diel samplings, with the exception of 60 and 75 m for TT12D2, where the peak was at noon). This is similar to the results of Vaultot et al. (1995) who analyzed samples from the same diel experiments for cell cycle analysis of DNA. Near 1400 h, they saw a population of cells entering the DNA synthesis (S) phase at 30 m and below, while the cells in the surface layer (0 and 15 m) did not enter S phase until 1700 h (dusk). Then, by 2300 h, they saw that the cells had divided; this corresponds to the time when we see that FLS is again near minimum (and remains there until dawn of the next day). Thus the peak that we see in *Prochlorococcus* FLS appears to correspond with the time when the cells are in the S phase, and the minimum in FLS corresponds with the time when division is complete.

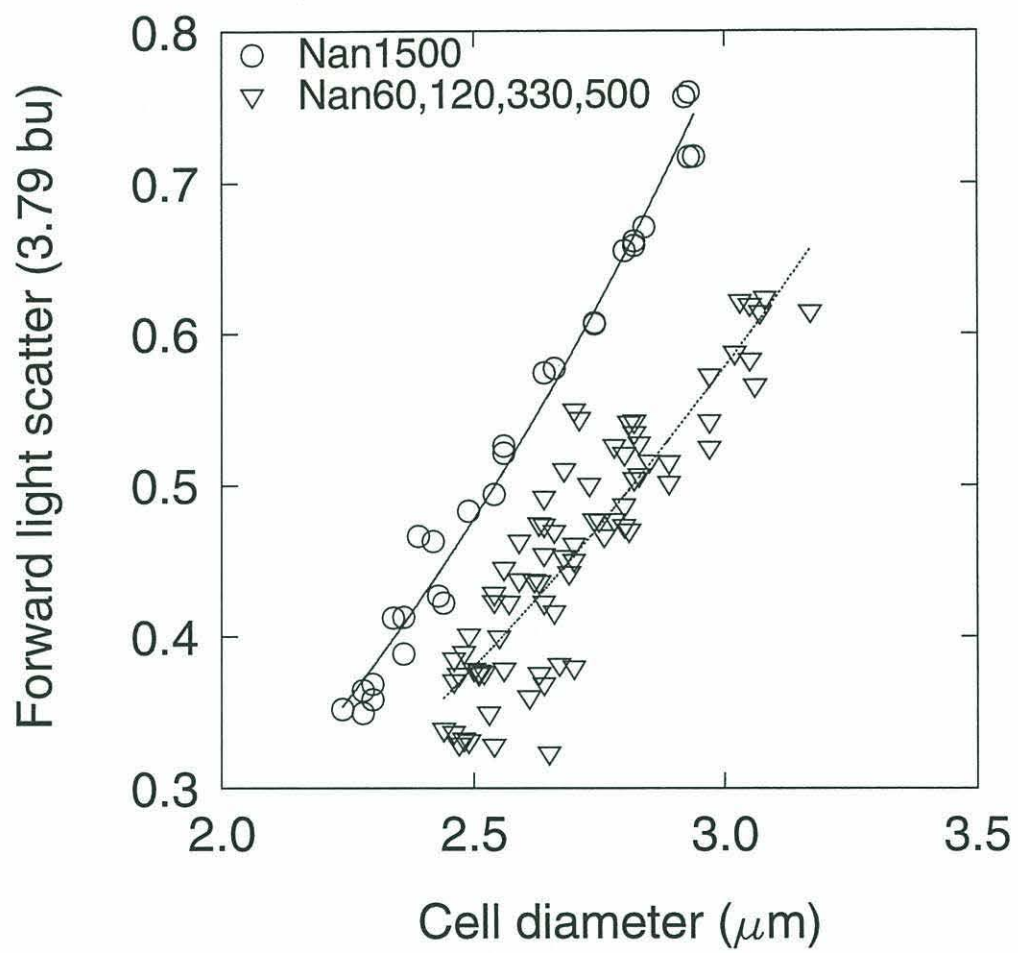
The maximum forward light scatter for *Synechococcus* in the equatorial Pacific occurs earlier in the day than for the other phytoplankton groups examined (Fig. 1C). The maximum usually occurs near noon (and occasionally, 3 out of 17 times, at the following sampling point). The *Synechococcus* concentration is at a minimum near the same time point and then increases dramatically shortly thereafter.

Laboratory calibrations

In order to convert FLS measurements made on field populations to cell size and carbon, we made measurements on *Nannochloris* cultures over the diel cycle (see Chapter 2). According to Mie theory, the forward light scatter for cells in the size range of *Nannochloris* and refractive indices typical of phytoplankton should be proportional to diameter to the power of three ($FLS \propto \text{diameter}^3$; FLS determined using Mie scattering program of Ackleson and Spinrad 1988). We first determined a simple linear regression of FLS vs. cell size for each laboratory diel experiment on *Nannochloris* separately (see Chapter 2) and performed an analysis of covariance: the hypothesis that all slopes (β) are equal was rejected ($p < 0.0005$). A multiple comparison testing of the slopes (Tukey test) revealed that they came from two separate populations, where $\beta_{\text{Nan120}} = \beta_{\text{Nan60}} = \beta_{\text{Nan500}} = \beta_{\text{Nan330}} \neq \beta_{\text{Nan1500}}$ (we cannot conclude how β_{Nan250} is related to the other population slopes). The implication is that Nan1500, the natural light experiment, is different from the others and so we will consider it separately. Since extending a linear regression between mean forward light scatter and mean cell diameter to cells of much different sizes is not appropriate, the data were then fitted with a power relationship ($FLS \propto \text{Diameter}^a$) which was used to convert forward light scatter measured in the field to cell diameter (Fig. 4; for Nan1500: $FLS = 0.0388 * \text{Diameter}^{2.74}$, $n = 28$, $r^2 = 0.98$; for Nan60,120,330,500: $FLS = 0.0463 * \text{Diameter}^{2.30}$, $n = 77$, $r^2 = 0.80$).

For picoplankton, the predicted relationship is that forward light scatter varies with diameter to the sixth power (Morel and Bricaud 1986). For measurements of

Figure 4. Relationship between forward light scatter (FLS in 3.79 μm bead units) and cell diameter (μm) for *Nannochloris* cells measured over the diel cycle in the laboratory. Data for cultures grown at 1500 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ (Nan1500) outdoors under natural sunlight were fitted with a power relationship ($\text{FLS} = 0.0388 * \text{Diameter}^{2.74}$, $n = 28$, $r^2 = 0.98$). Data for cultures grown at 60, 120, 330, and 500 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ (Nan60, 120, 330, 500) in an incubator were fitted separately ($\text{FLS} = 0.0463 * \text{Diameter}^{2.30}$, $n = 77$, $r^2 = 0.80$).



Synechococcus in the laboratory, we determined by best fit that $FLS \propto \text{Diameter}^{5.4}$; $n = 8$, $r^2 = 0.90$). We do not have an experimentally determined fit for *Prochlorococcus*. To convert *Synechococcus* and *Prochlorococcus* FLS to cell size, we will assume a power relationship of either 5.4 or 6 and compare the results.

From laboratory relationships between forward light scatter and carbon per cell, we have determined a conversion to use for field data for the eukaryotic phytoplankton. Data for cultures grown at $1500 \mu\text{mol photons m}^{-2} \text{ s}^{-1}$ (Nan1500) outdoors under natural sunlight were fitted with a power relationship ($FLS = 3.53\text{e-}4 * \text{Carbon}^{0.928}$, $n = 28$, $r^2 = 0.94$; Fig. 5). Data for cultures grown at $500 \mu\text{mol photons m}^{-2} \text{ s}^{-1}$ (Nan500) in an incubator were fitted separately ($FLS = 3.30\text{e-}3 * \text{Carbon}^{0.6350}$, $n = 25$, $r^2 = 0.89$). These two data sets were considered independently since an analysis of covariance with the hypothesis that the slopes of a linear regression for Nan250, 500, and 1500 (β) are equal was rejected ($p < 0.001$). A multiple comparison testing of the slopes (Tukey test) revealed that they came from two separate populations, where $\beta_{\text{Nan500}} \neq \beta_{\text{Nan1500}}$ (we cannot conclude how β_{Nan250} is related to the other population slopes).

Growth rates

Estimates of growth rates were calculated by converting FLS to volume or carbon using the above equations and then applying the equation $\mu = \ln (\text{Max/Min})$ to the calculated volume or carbon data, as described in the methods. The results from using the Nan1500 FLS and diameter relationship ($FLS \propto d^{2.74}$, or $\text{Volume}^{0.913}$)

were very close (within 2%) to those from using the Nan1500 FLS and carbon relationship ($\text{FLS} \propto \text{carbon}^{0.928}$). Using the Nan60,120,330,500 FLS and diameter relationship ($\text{FLS} \propto d^{2.3}$, or $\text{Volume}^{0.767}$) or the Nan500 FLS and carbon relationship led to estimates of μ 19% or 44% higher, respectively, than using the Nan1500 FLS and diameter relationship. The growth rate estimates presented here were calculated using the Nan1500 FLS and diameter relationship. For the picoplankton, estimates of growth rates using $\text{FLS} \propto d^{5.4}$ were 11% higher than those using $\text{FLS} \propto d^6$. The results presented here use $\text{FLS} \propto d^6$ for the calculations.

Equatorial Pacific growth rates

For the equatorial Pacific diel samplings, the specific growth rates of *Prochlorococcus* calculated from flow cytometric forward light scatter changes over the day compare well to those of Vaultot et al. (1995), which were determined from cell cycle analysis of DNA distributions from the same diel samplings (Fig. 6A, B, C). For each of the three diel samplings (TT8D1, TT12D1, and TT12D2), the specific growth rate is maximum at 30 m, decreases slightly toward the surface, and decreases more with depth to 100 m. The maximum growth rate is approximately equal to one doubling per day. It is encouraging that the growth rates calculated here show the same general trend as those of Vaultot et al. Linear regressions of μ calculated from FLS (separate estimates from the day increase due to cell growth and night decrease due to cell division) and growth rates from DNA measurements show strong correlations ($r^2 = 0.82$ for day, $r^2 = 0.91$ for night; Fig. 7). The estimates

Figure 5. Relationship between forward light scatter (FLS in 3.79 μm bead units) and cell carbon (fg) for *Nannochloris* cells measured over the diel cycle in the laboratory. Data for cultures grown at 1500 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ (Nan1500) outdoors under natural sunlight were fitted with a power relationship ($\text{FLS} = 3.53\text{e-}4 * \text{Carbon}^{0.928}$, $n = 28$, $r^2 = 0.94$). Data for cultures grown at 500 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ (Nan500) in an incubator were fitted separately ($\text{FLS} = 3.30\text{e-}3 * \text{Carbon}^{0.6350}$, $n = 25$, $r^2 = 0.89$).

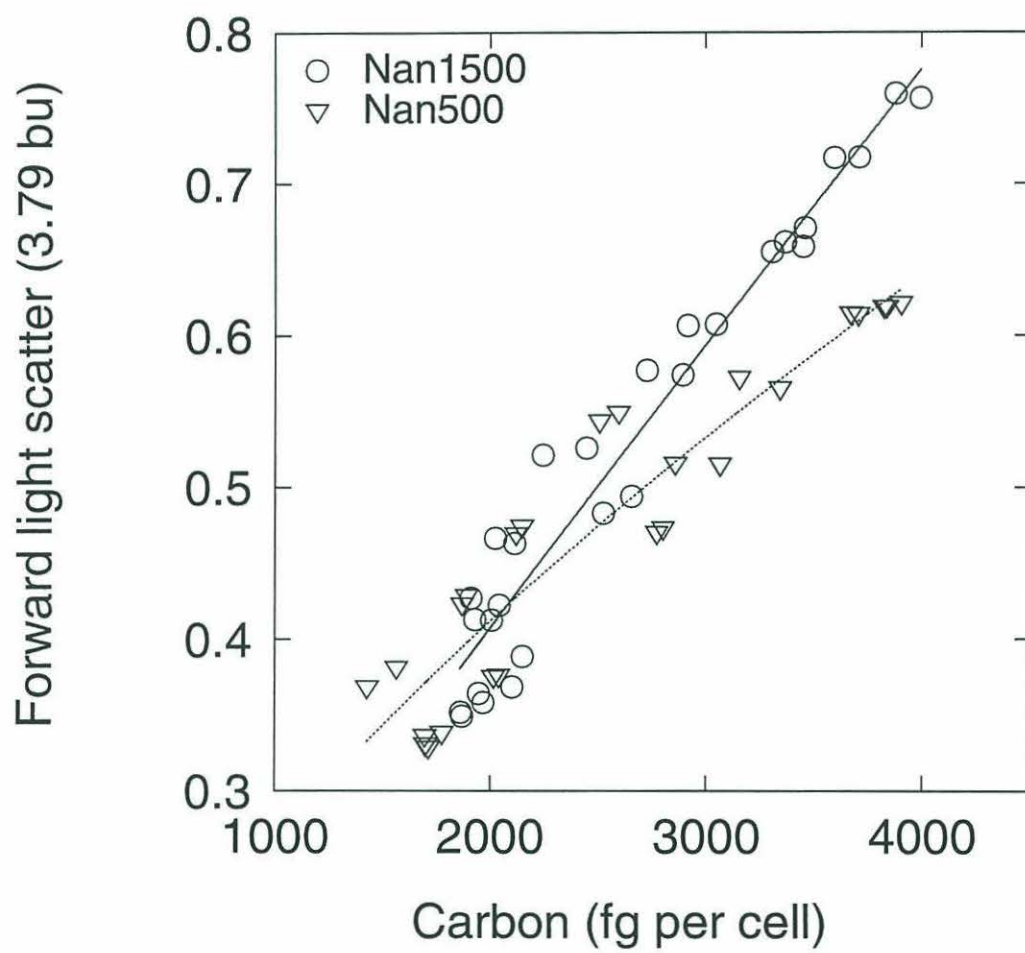


Figure 6. Depth profiles of specific growth rates (μ , d^{-1}) for *Prochlorococcus* (A, B, C) and *Synechococcus* (D, E, F) in the equatorial Pacific during one diel sampling in April (TT8D1) and two in October 1992 (TT12D1 and TT12D2). Growth rates calculated from the daytime increase in forward light scatter FLS, the nighttime decrease in FLS, or the DNA distributions (for *Prochlorococcus*, μ from DNA from Vaultot et al. 1995). Surface light intensity data from R.T. Barber (<http://www1.whoi.edu>).

Specific Growth Rate (μ , d^{-1})

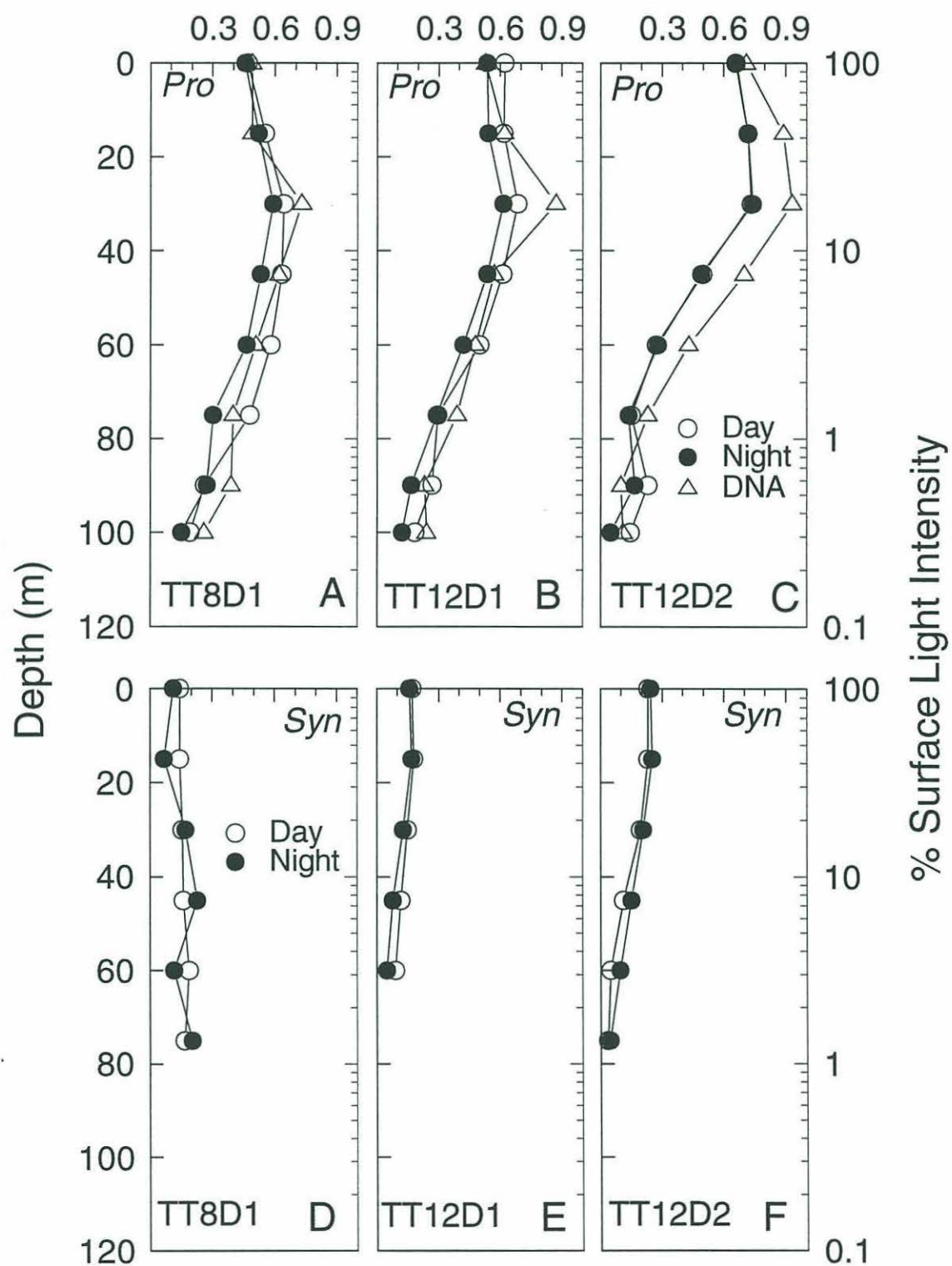
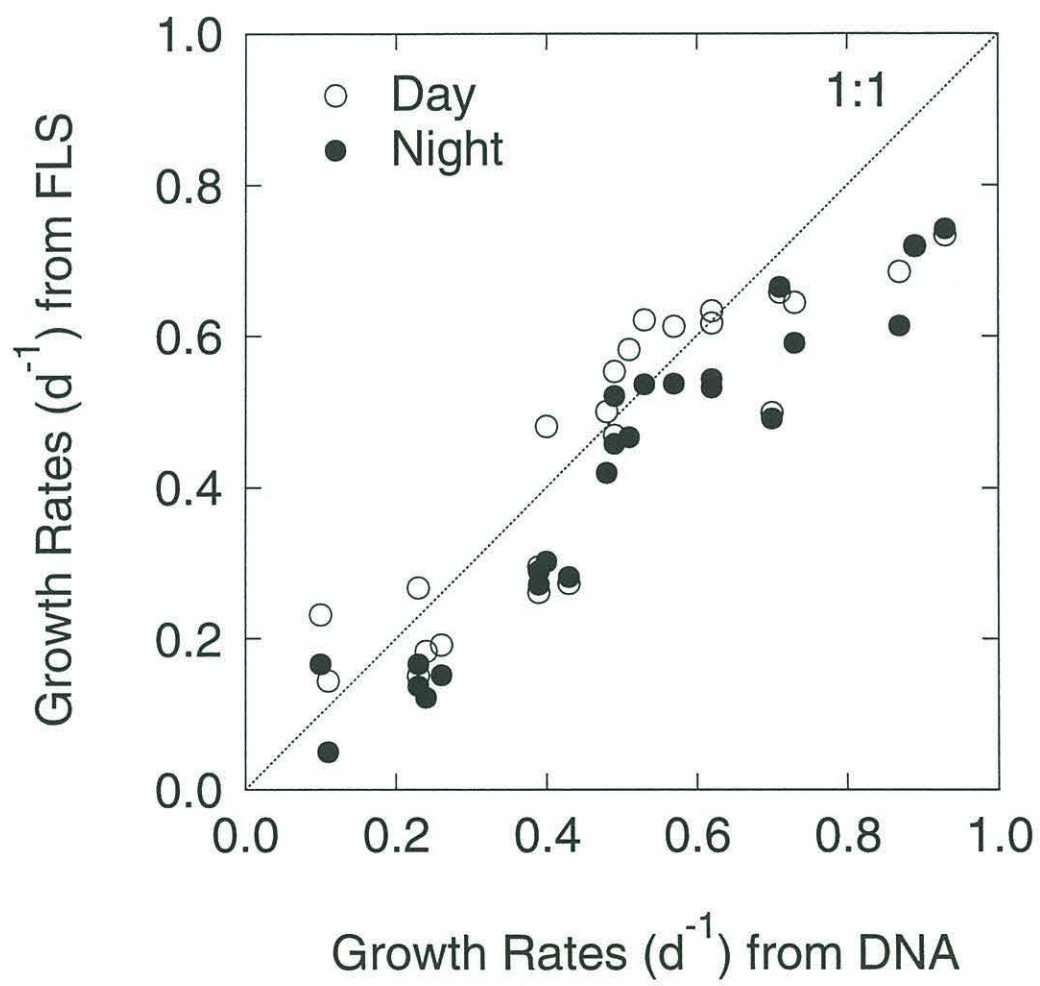


Figure 7. Comparison of *Prochlorococcus* growth rates estimated from FLS (daytime increase or nighttime decrease) and DNA distributions (Vaulot et al. 1995). The dotted line shows a 1:1 relationship.



from FLS tend to be lower than those from DNA at the highest growth rates.

However, Vaultot et al. state that their highest measurements (up to 0.93 d^{-1}) are overestimates of a doubling time of one day ($\mu = 0.69 \text{ d}^{-1}$); since there was tight synchrony and only one maximum in the synthesis phase cells, the maximum must have been one division per day. Our highest estimate is 0.74 d^{-1} , close to the maximum expected rate of 0.69 d^{-1} .

Estimated growth rates for *Synechococcus* in the equatorial Pacific reach only 0.25 d^{-1} (doubling time of once every ~ 3 days; Fig. 6D, E, F). For TT8D1, μ is about the same down to 75 m; for TT12D1 and TT12D2, μ is maximum at the surface and 15 m and decreases slightly with depth. Growth rates calculated from cell concentrations measured in the field (thus a minimum growth rate; i.e., growth minus grazing) were three-fold higher (on average in the upper 60 m) than growth rates estimated from FLS. Thus the growth rates for *Synechococcus* using the FLS method are almost certainly underestimates. We believe this underestimation is due to incomplete phasing of cell division to the light:dark cycle. This interpretation is supported by previous observations of patterns in the frequency of dividing cells (FDC) in *Synechococcus*, which would be expected to be similar to patterns in FLS. Waterbury et al. (1986) saw the frequency of dividing cells increase from 8% to only 32% in a culture with a growth rate of 0.85 d^{-1} . Campbell and Carpenter (1986), in studies of cultures at different light levels and temperatures, determined growth rates from 0.10 to 1.02 d^{-1} , yet the FDC at the highest growth rate only reached 22%. At the fastest growth rates, they observed that FDC was high all day and after the light

period ended; for growth rates less than 0.69 d^{-1} , the peak in FDC was sharper, occurring over only 2 to 4 hours. Armbrust et al. (1989) found that *Synechococcus* has two light-dependent gaps in DNA synthesis, thus cells can become blocked in the doublet stage during the night and not released until the light period. The model used here to estimate growth rates from FLS, assumes tight phasing and a maximum of one division per day; thus it does not appear that *Synechococcus* populations are amenable to growth rate analysis based on FLS patterns.

For the eukaryotic phytoplankton (subdivided into two distinguishable populations: ultra- and nanoplankton) in the equatorial Pacific, the specific growth rate was maximum in the upper waters and decreased with depth (Fig. 8). Growth rates for the ultraphytoplankton were higher during TT12 than TT8. The nanophytoplankton reached a maximum μ of about one doubling per day. According to these estimates, the ultraphytoplankton were growing faster than one doubling per day, which is impossible given the assumptions of the model. The assumptions are that a single peak in FLS during the day can, at the most, translate into each cell doubling in size before division. Thus the maximal detectable growth rate should be one doubling per day ($\mu = 0.69\text{ d}^{-1}$). Since the ultraphytoplankton cells are considerably smaller than the *Nannochloris* cells used in the laboratory, it is quite possible that our lab calibration is inappropriate for these smaller cells. If we assume that FLS varies with diameter to the fourth power rather than to the 2.74 power, then we obtain estimates of a maximum μ of 0.73, close to one doubling per day.

Figure 8. Depth profiles of specific growth rates (μ , d^{-1}) for ultraphytoplankton (A, B, C) and nanophytoplankton (D, E, F) in the equatorial Pacific during one diel sampling in April (TT8D1) and two in October 1992 (TT12D1 and TT12D2). Growth rates calculated from the daytime increase in forward light scatter FLS or the nighttime decrease in FLS. Surface light intensity data from R.T. Barber (<http://www1.who.edu>).

Sargasso Sea growth rates

During a January 1992 cruise to the Sargasso Sea, two diel sampling experiments were performed. The first diel (EnD1) was about 350 nautical miles S-SW of Bermuda, where there was a surface mixed layer of 20-40 m. The estimated growth rates of the nanophytoplankton population were maximum in the upper water ($0.4\text{--}0.45\text{ d}^{-1}$ at 0 and 20 m) and decreased with depth (Fig. 9A, B). They were roughly the same for the first and second days of the sampling. The second diel (EnD2) was about 200 nautical miles NW of Bermuda, where the water column was mixed down to 200 m and there was $1\text{ }\mu\text{M}$ nitrate at the surface. The day was cloudy (about $100\text{ }\mu\text{mol photons m}^{-2}\text{ s}^{-1}$ just below the surface at noon) and it rained heavily in the morning. The nanophytoplankton were growing at a rate of about one doubling per day near the surface, slightly lower at 20m, then dramatically lower at 50 m ($\sim 0.1\text{ d}^{-1}$), and near zero down to 200 m (Fig. 9C). Growth rates of the coccolithophore population, which was distinguishable during this diel, had the same trend with depth (Fig. 9D). However, the values calculated are likely to be underestimates, since the coccolithophore cells are considerably larger than the *Nannochloris* in the laboratory which were used for the calibration.

During a July 1993 cruise to the Sargasso Sea, a diel sampling (Iselin) was performed about 200 nautical miles N-NW of Bermuda where the mixed layer was about 20 m deep. *Prochlorococcus* growth rates were about 0.3 d^{-1} in the upper 40 m and increased with depth (Fig. 10A, B). These rates in the upper water are similar to those determined by Goericke and Welschmeyer (1993) for *Prochlorococcus* in the

Sargasso Sea from measuring the incorporation of ^{14}C into divinyl-chlorophyll *a*. The pattern seen here with depth (increasing growth rate) was only seen during the March 1985 sampling (out of 9 growth rate profiles from March 1985 - September 1986) by Goericke and Welschmeyer. It is possible, however, that we were not tracking the same water mass at the deepest depths (the drogue was set at 25 m). Our estimated growth rates for *Synechococcus* were at a maximum of $0.2\text{-}0.3\text{ d}^{-1}$ at 25 m and decreased both toward the surface and with depth, though these rates are most likely an underestimate due to incomplete phasing, as discussed above (Fig. 9C, D). For both *Prochlorococcus* and *Synechococcus*, the growth rates the second day of the diel were slightly lower than the first day. Growth rates of the eukaryotic nanophytoplankton were $0.5\text{-}0.6\text{ d}^{-1}$ in the upper waters and decreased only slightly with depth during both days of the sampling (Fig. 10E, F).

During a May 1994 cruise to the NW Sargasso Sea, a diel sampling experiment (Hat) was performed about 350 nautical miles NW of Bermuda, where the upper 20-30 m was mixed. The 48-h sampling was performed the day after a storm with 40 knot winds. For this diel, only the eukaryotic nanophytoplankton were analyzed. Their growth rates were maximum (near 0.5 d^{-1}) at 15 and 25 m and decreased slightly at the surface and 35 m (the deepest depth analyzed; Fig. 11). The second day of the sampling showed nearly constant rates in the upper 25 m, with a slight decrease at 35 m. It is interesting to note that, for the first day, the estimates based on the daytime increase in FLS or the nighttime decrease in FLS match quite well, indicating that the FLS at the first dawn and the second dawn are

Figure 9. Depth profiles of specific growth rates (μ , d^{-1}) for nanophytoplankton in the Sargasso Sea during a 48-hour diel sampling experiment in January 1992 at 27°N 68°W (A and B: R.V. *Endeavor*, EnD1, Day1 and Day2) and for nanophytoplankton and coccolithophores during a 28-hour diel sampling experiment in January 1992 at 35°N 68°W (C and D: EnD2). Growth rates calculated from the daytime increase in forward light scatter FLS or the nighttime decrease in FLS. Surface light intensity from a noon light cast using a Biospherical Instruments Inc. QSP-200 4π sensor.

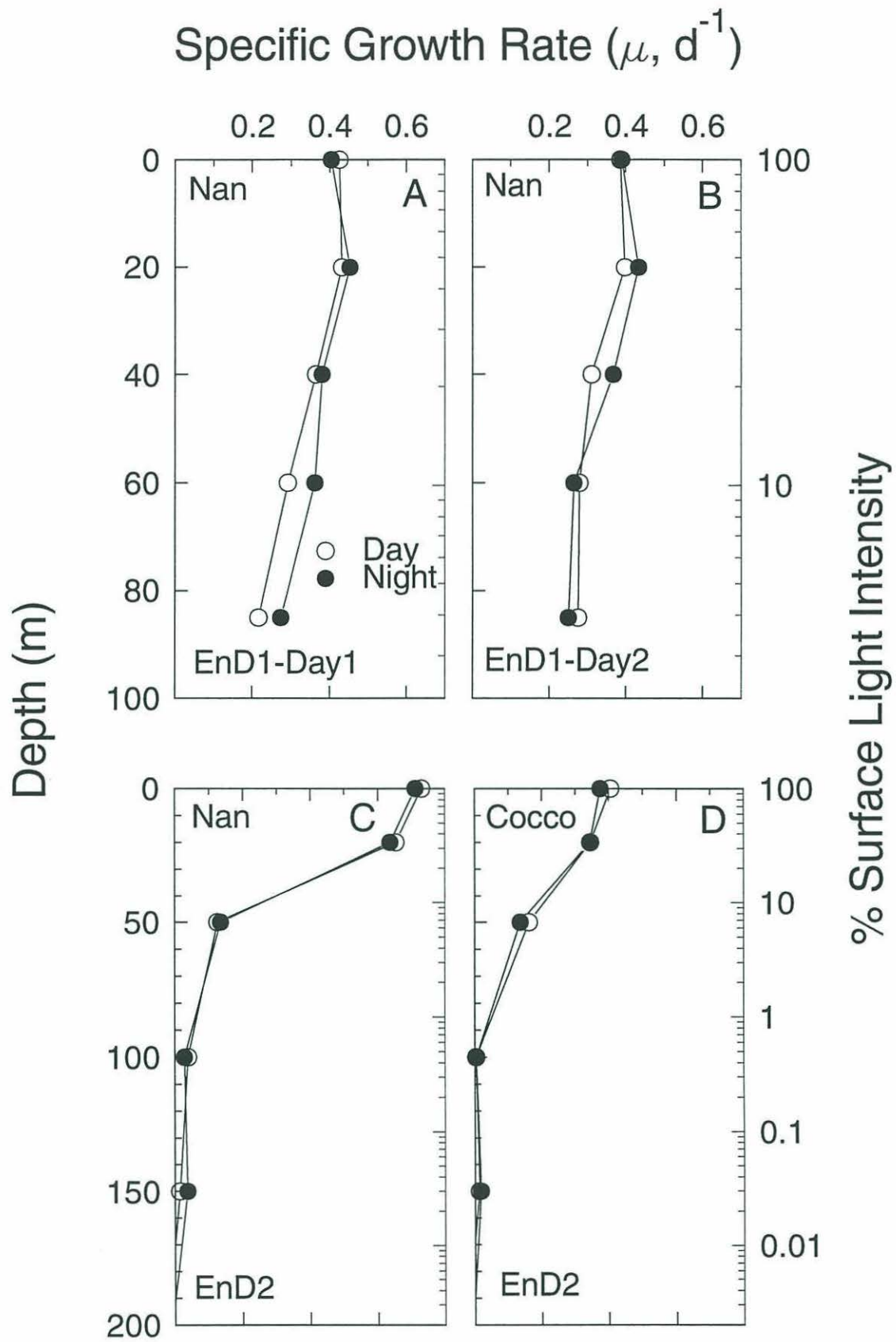
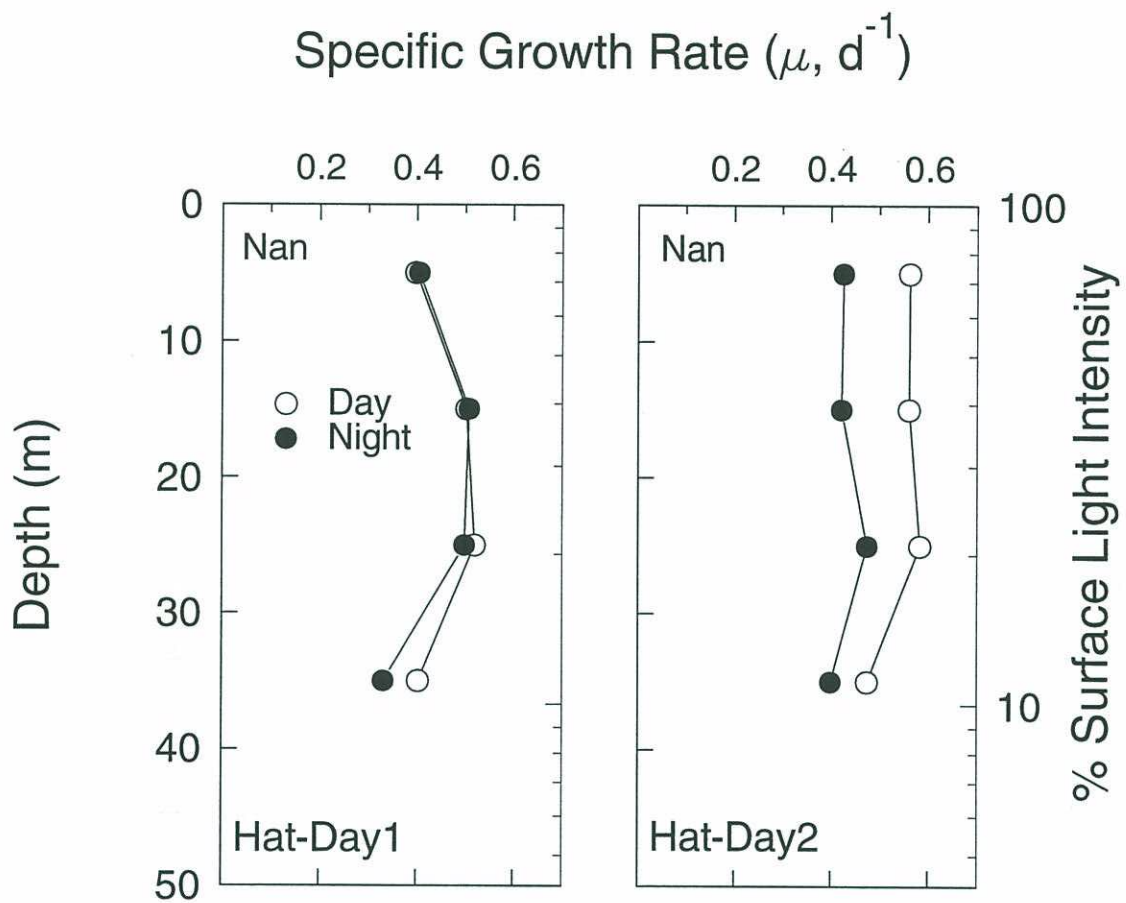


Figure 10. Depth profiles of specific growth rates (μ , d^{-1}) for *Prochlorococcus* (A, B), *Synechococcus* (C, D), and nanophytoplankton (E, F) in the Sargasso Sea during a 40-hour diel sampling experiment in July 1993 at 35°N 69°W (R.V. *Columbus Iselin*, Iselin, Day1 and Day2). Growth rates calculated from the daytime increase in forward light scatter FLS or the nighttime decrease in FLS. Surface light intensity from measurements of Secchi depth.

Figure 11. Depth profiles of specific growth rates (μ , d^{-1}) for nanophytoplankton in the Sargasso Sea during a 48-hour diel sampling experiment in May 1994 at 35°N 71°W (R.V. *Cape Hatteras*, Hat, Day1 and Day2). Growth rates calculated from the daytime increase in forward light scatter FLS or the nighttime decrease in FLS. Surface light intensity from measurements of Secchi depth.



the same. For the second day, however, these separate estimates do not match as well, since the FLS was lower for the second dawn than the third dawn, indicating that the phytoplankton were not at steady state.

Despite the uncertainties in the absolute magnitude of some of the growth rate estimates (discussed above), the overall patterns seen are encouraging. An additional problem, however, would be if the phytoplankton cells in the field are significantly changing refractive index during the day, or if cells at different depths have different refractive indices. Changes in refractive index were not taken into account for the calculations presented here. For the laboratory studies on *Nannochloris*, we saw a strong relationship between cell volume and cell carbon, and only small changes in refractive index over the course of the day; the scattering was primarily due to changes in cell size and not changes in refractive index (Chapter 2). However, we could not make measurements of refractive index in the field on specific populations of phytoplankton, thus we are assuming that the relationship seen in the lab applies. If the cells in the field are changing refractive index during the course of the day, and changes in FLS do not reflect changes in cell carbon, then the laboratory calibration may not be appropriate for these cells in the field. Others have seen changing refractive indices during laboratory studies on cultured phytoplankton (Ackleson et al. 1993, Stramski and Reynolds 1993, Stramski et al. 1995).

Loss rates

Usually, the cell concentrations at dawn of one day are about the same as the cell concentrations at dawn of the next day. This means that growth rates and loss rates approximately balance over the day for the diel samplings in the equatorial Pacific and the Sargasso Sea (Figs. 12-15). The loss rates calculated here are *in situ* loss rates, thus they may be the result of grazing, mixing, and other processes such as aggregation that could repackage cells so that they are no longer present at concentrations that would be sampled in 50 ml. In some cases, the loss rates are greater than the growth rates; this appears to occur more frequently at depth than in the upper waters. However, during Hat Day1 and 2, the loss rates exceeded the growth rates for the nanophytoplankton in the upper waters (5-35 m). The only times that the growth rates exceeded the loss rates were for *Prochlorococcus* during TT12D2, for nanophytoplankton during EnD1 Day2 (which were approximately balanced during Day1), and for *Synechococcus* during the Iselin diel sampling.

Phytoplankton growth and microzooplankton grazing rates were determined from dilution experiments during the same cruises to the equatorial Pacific (though not on the same days; Verity et al. 1994). They found that growth and grazing rates were higher in the fall (TT12) than the spring (TT8) at 15 and 60 m. In the spring, grazing exceeded growth and in the fall, growth and grazing approximately balanced. These results are similar to the ones presented here (Fig. 12).

Figure 12. Depth profiles of specific growth rates (μ from daytime increase in FLS, d^{-1}) and loss rates (d^{-1}) for *Prochlorococcus* (A, B, C), ultraphytoplankton (D, E, F), and nanophytoplankton (G, H, I) in the equatorial Pacific during one diel sampling in April (TT8D1) and two in October 1992 (TT12D1 and TT12D2).

Specific growth rate (\circ) or Loss rate (\blacksquare) d^{-1}

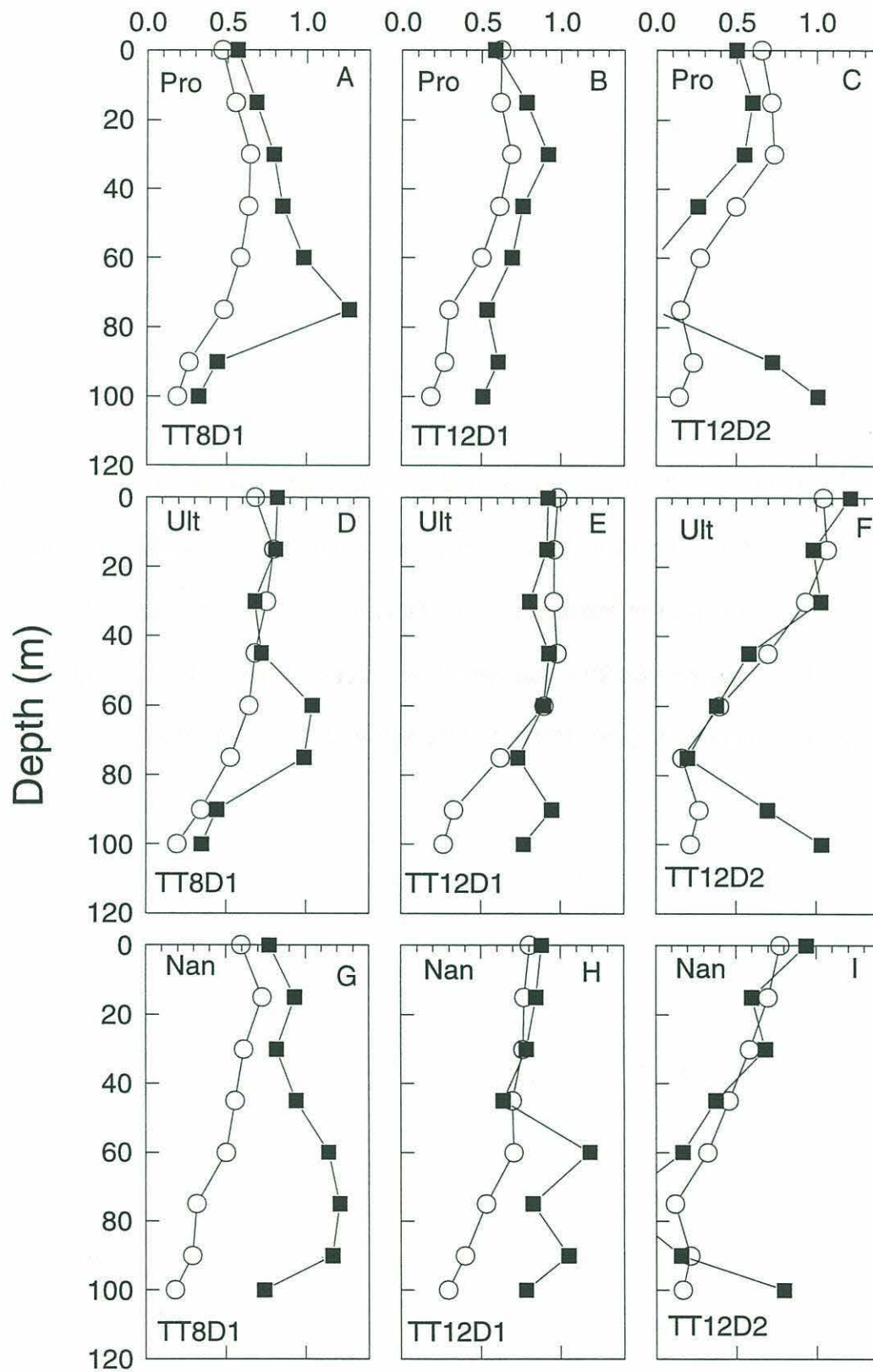


Figure 13. Depth profiles of specific growth rates (μ from daytime increase in FLS, d^{-1}) and loss rates (d^{-1}) for nanophytoplankton in the Sargasso Sea during a 48-hour diel sampling experiment in January 1992 at 27°N 68°W (A and B: EnD1, Day1 and Day2) and for nanophytoplankton and coccolithophores during a 28-hour diel sampling experiment in January 1992 at 35°N 68°W (C and D: EnD2).

Specific growth rate (○) or Loss rate (■) d^{-1}

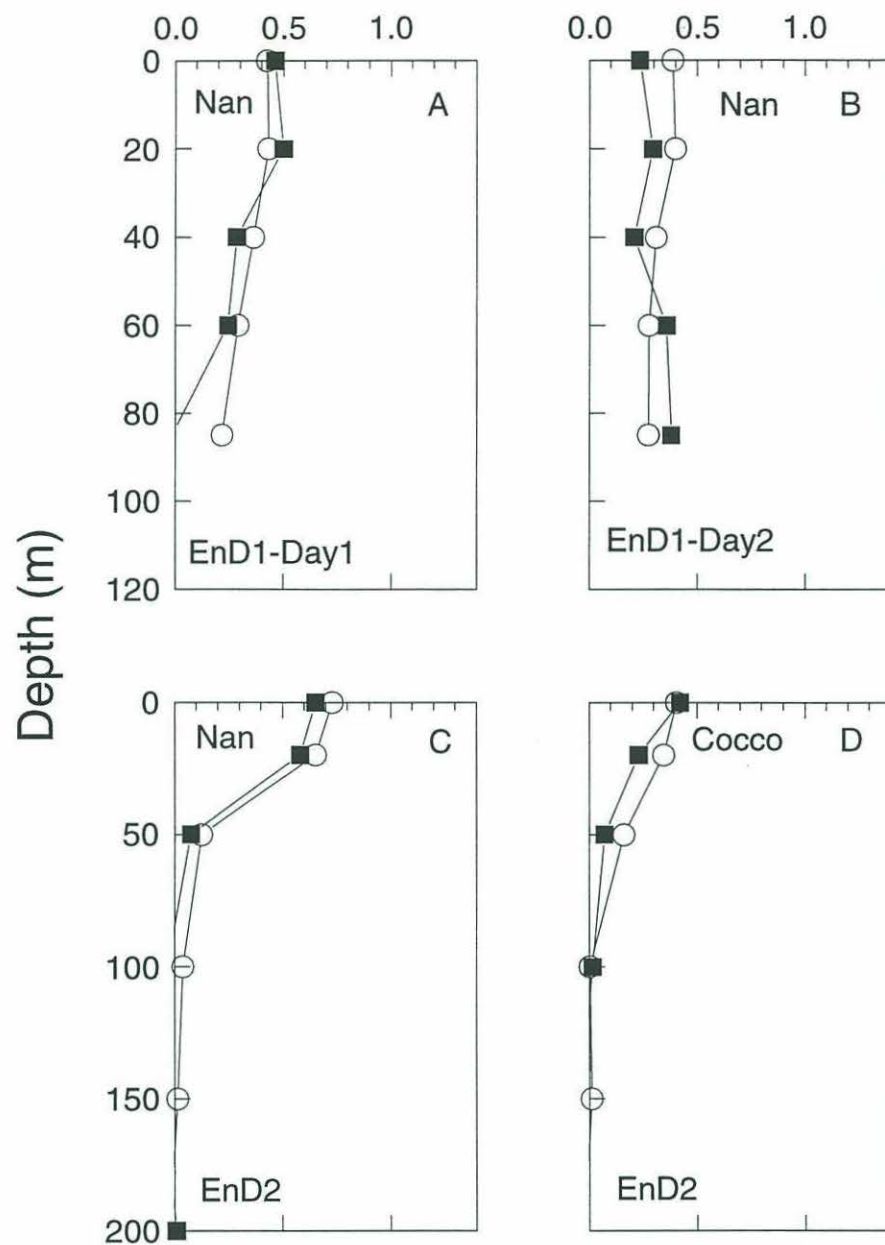


Figure 14. Depth profiles of specific growth rates (μ from daytime increase in FLS, d^{-1}) and loss rates (d^{-1}) for *Synechococcus* (A), and nanophytoplankton (B) in the Sargasso Sea during a 40-hour diel sampling experiment in July 1993 at 35°N 69°W (R.V. *Columbus Iselin*, Iselin).

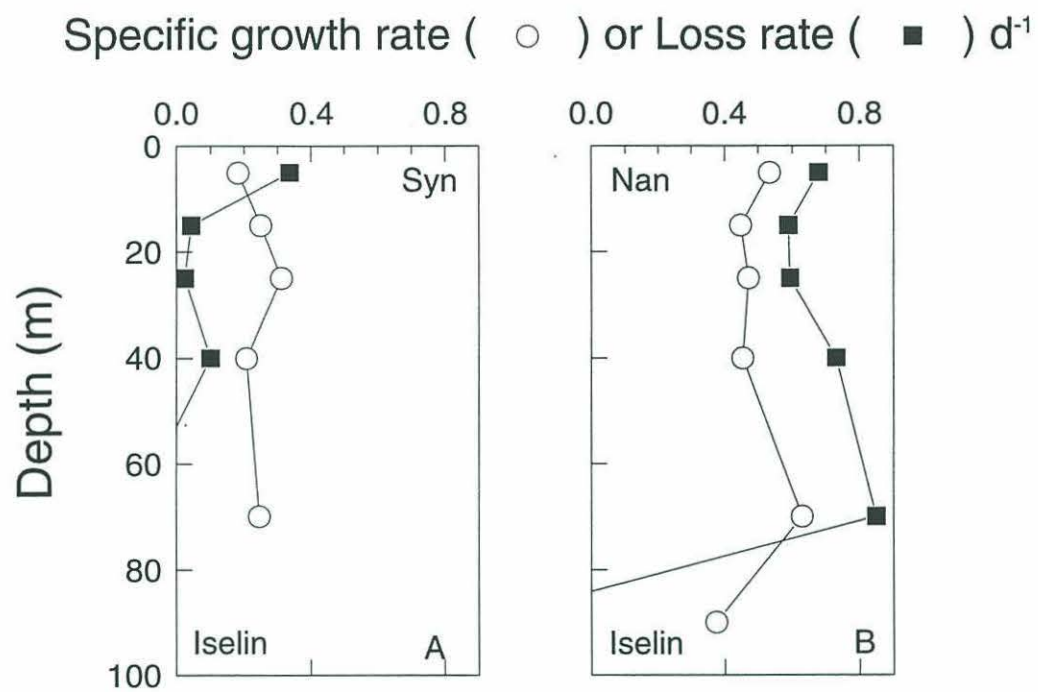


Figure 15. Depth profiles of specific growth rates (μ from daytime increase in FLS, d^{-1}) and loss rates (d^{-1}) for nanophytoplankton in the Sargasso Sea during a 48-hour diel sampling experiment in May 1994 at 35°N 71°W (Hat, Day1 and Day2).

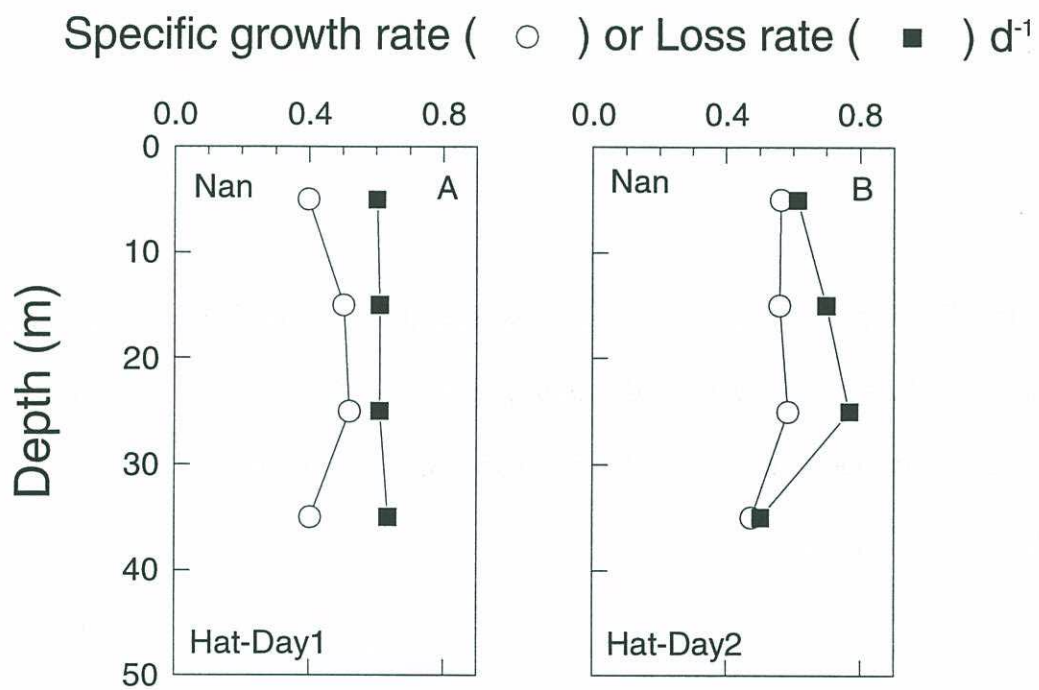


Figure 16. Depth profiles of day or night loss rates (h^{-1}) for *Prochlorococcus* (A, B, C), ultraphytoplankton (D, E, F), and nanophytoplankton (G, H, I) in the equatorial Pacific during one diel sampling in April (TT8D1) and two in October 1992 (TT12D1 and TT12D2).

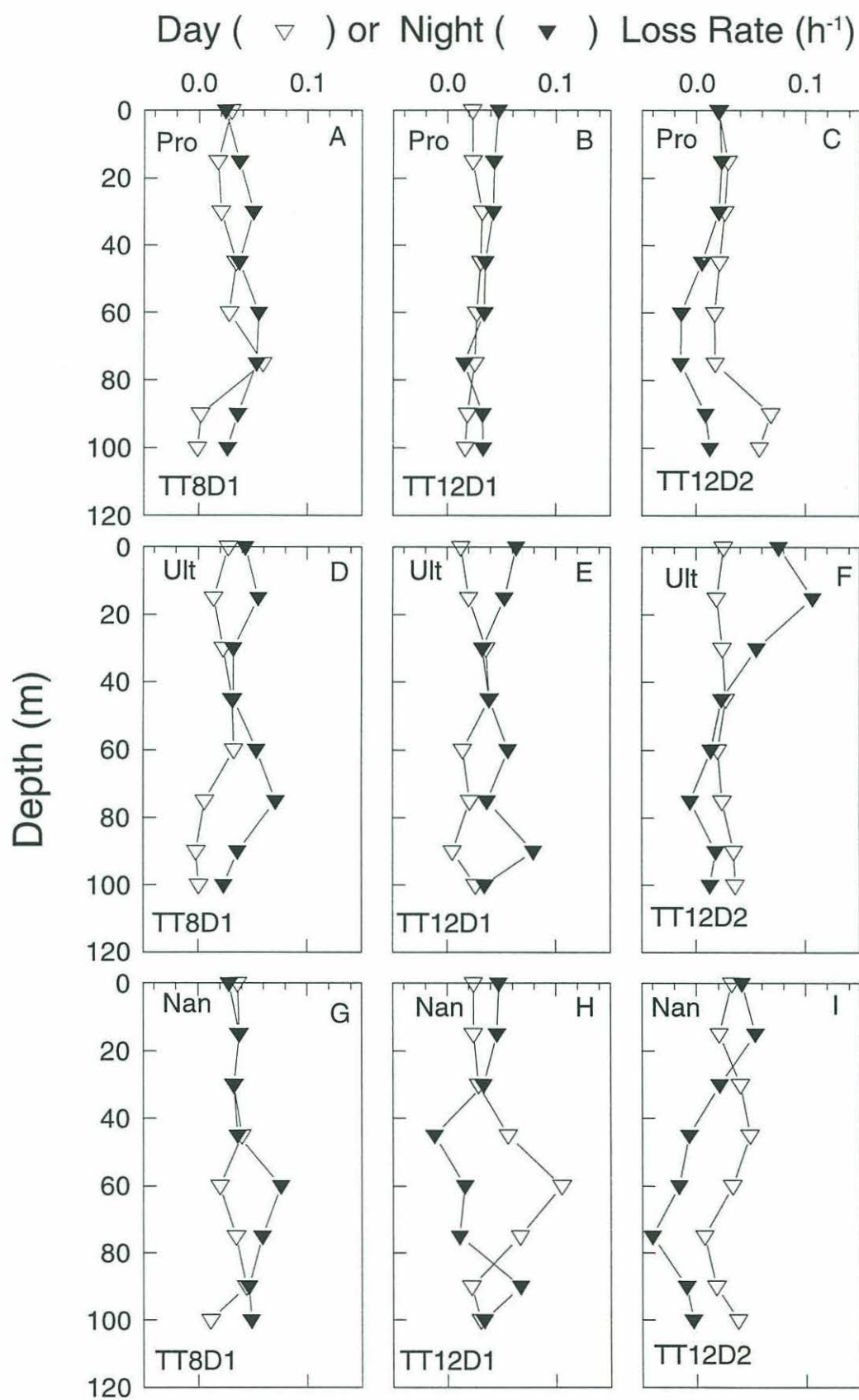


Figure 17. Depth profiles of day or night loss rates (h^{-1}) for nanophytoplankton in the Sargasso Sea during a 48-hour diel sampling experiment in January 1992 at 27°N 68°W (A and B: EnD1, Day1 and Day2) and for nanophytoplankton and coccolithophores during a 28-hour diel sampling experiment in January 1992 at 35°N 68°W (C and D: EnD2).

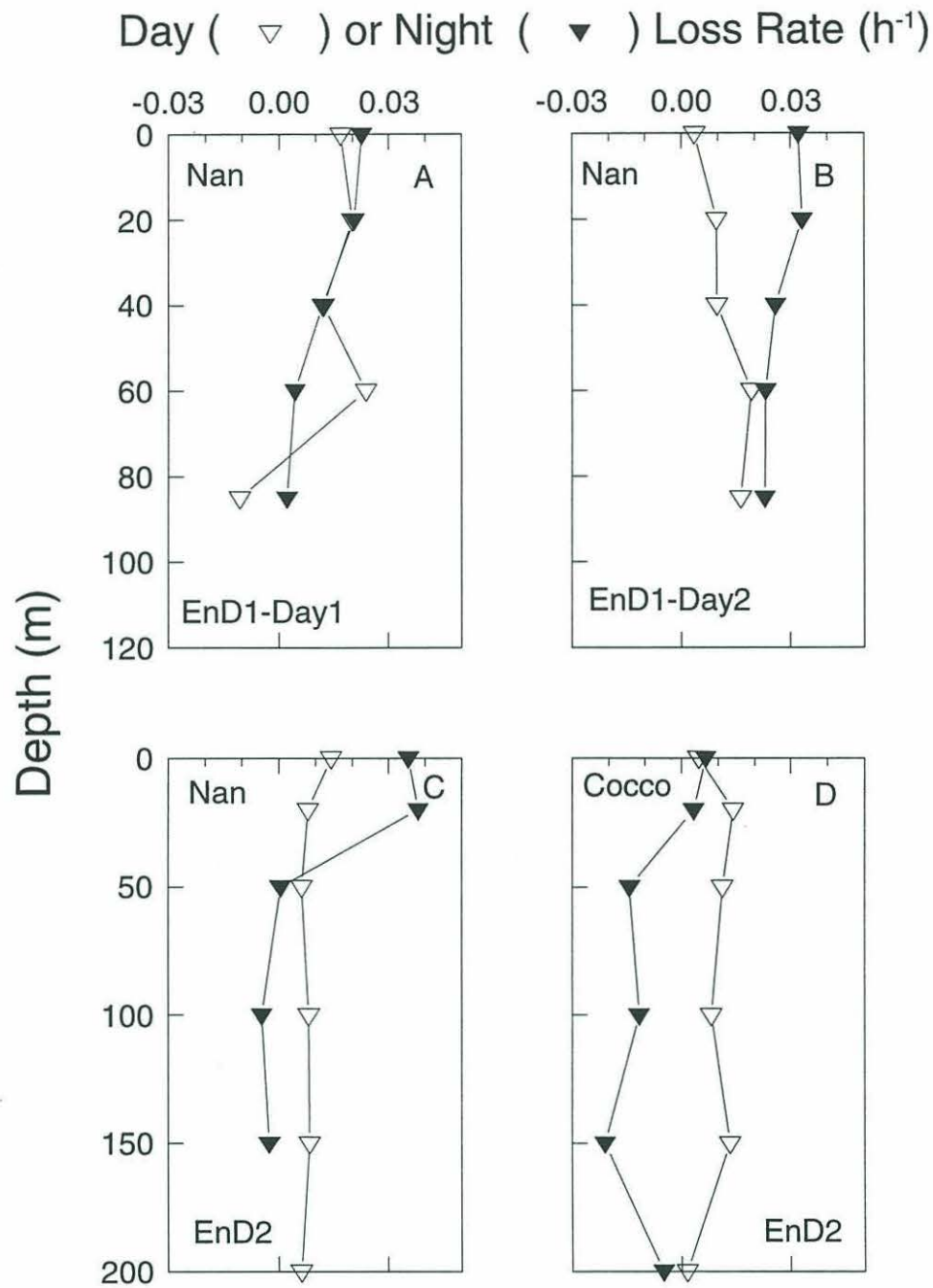


Figure 18. Depth profiles of day or night loss rates (h^{-1}) for *Synechococcus* (A), and nanophytoplankton (B) in the Sargasso Sea during a 40-hour diel sampling experiment in July 1993 at 35°N 69°W (R.V. *Columbus Iselin*, Iselin).

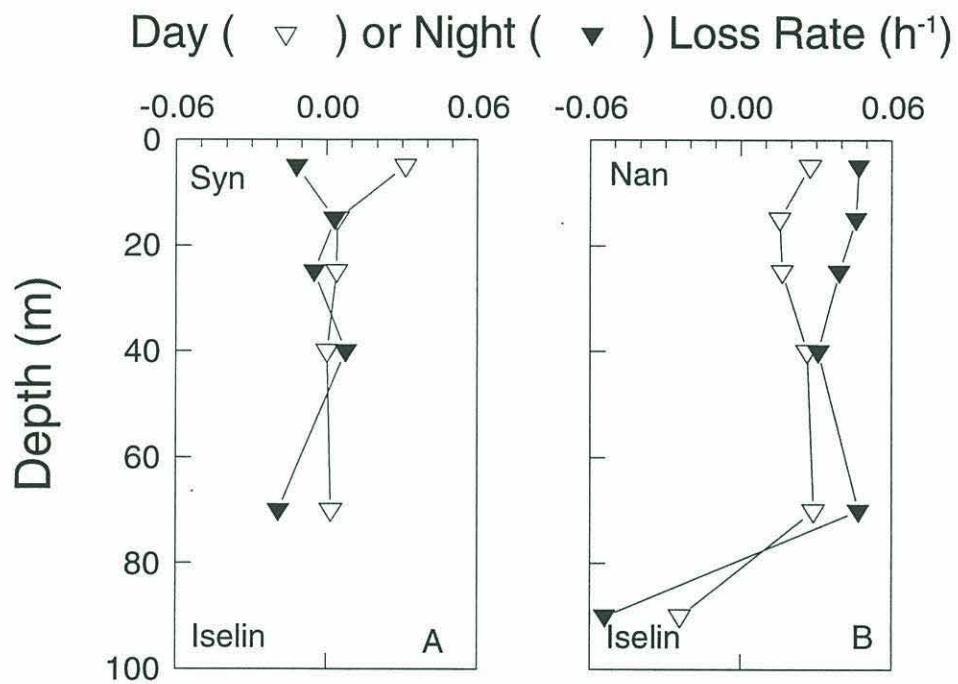


Figure 19. Depth profiles of day or night loss rates (h^{-1}) for nanophytoplankton in the Sargasso Sea during a 48-hour diel sampling experiment in May 1994 at 35°N 71°W (Hat, Day1 and Day2).

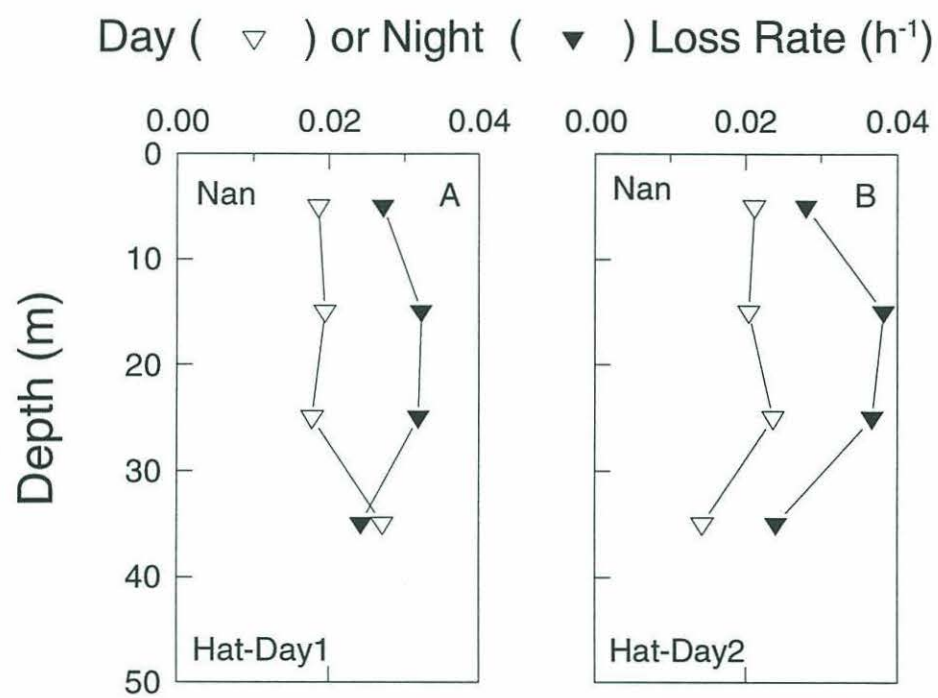
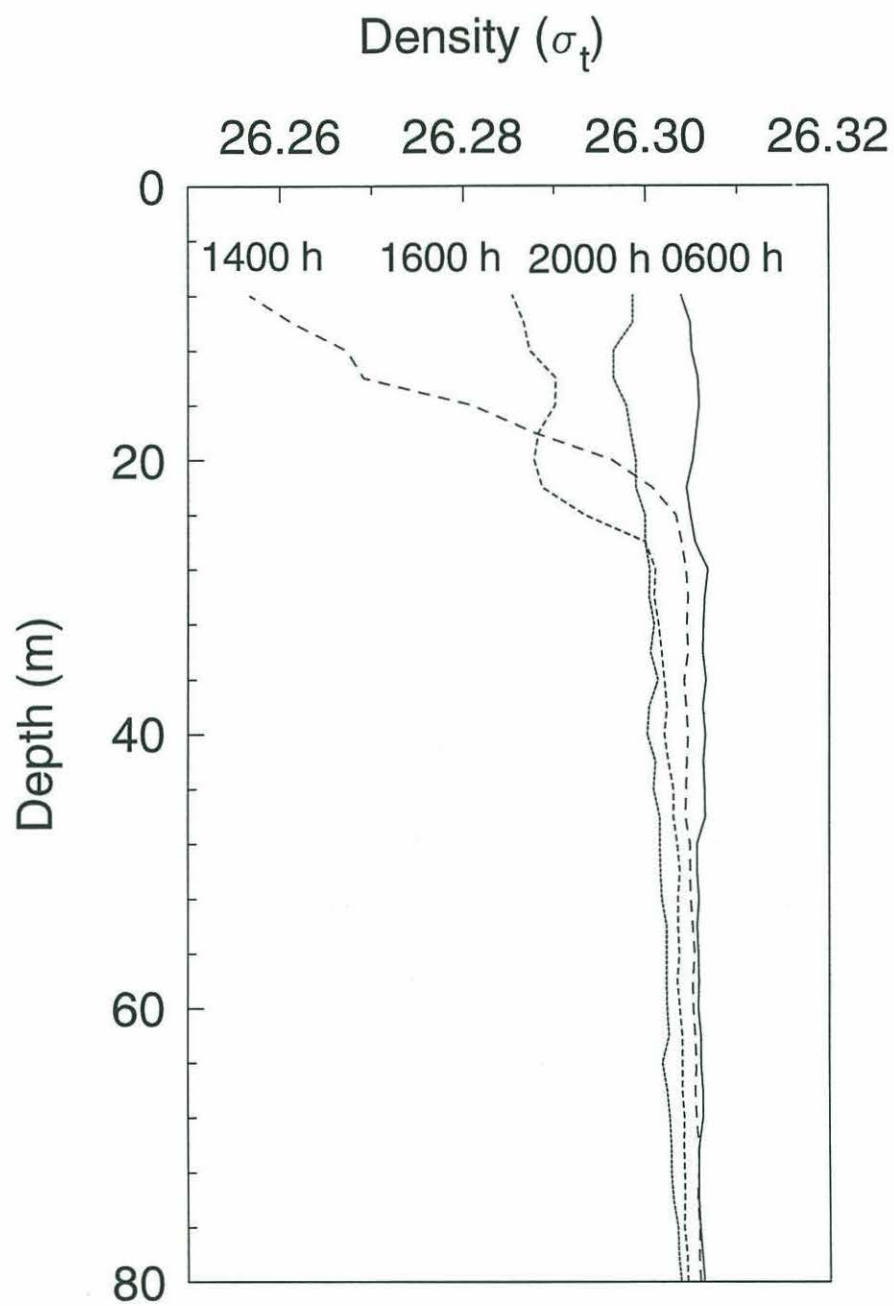


Figure 20. Depth profile of density (σ_t) during a 28-hour diel sampling experiment in January 1992 at 35°N 68°W (R.V. *Endeavor*, EnD2) at selected times (0600, 1400, 1600, and 2000 h). At 0600 h, the water column was deeply mixed. There was a rainstorm in the morning (heaviest around 1000 h), which produces less dense water near the surface in the 1400 h profile. At 1600 h, this less dense water had begun to mix downward. By 2000 h the water column again appeared to be deeply mixed.



Separate day and night loss rates were calculated as described in the methods. In general, night loss rates tend to be higher than day loss rates (with a few exceptions; Figs. 16-19). It is interesting to note that the high night loss rates for the nanophytoplankton at 0 and 20 m during EnD2 (Fig. 17C) can be attributed primarily to mixing. This is evident from an examination of depth profiles of density (Fig. 20). There was a nocturnal deepening of the surface layer, with cells from the surface waters being mixed downward after dark. Less dense water (from a heavy rainstorm in the morning) was trapped near the surface during the day and appears to have first mixed to 50 m (and deeper) sometime between 1600 h and 2000 h. Thus cells that were produced in the upper 20 m (see growth rates, Fig. 13C) were mixed deeper in the evening, causing the high nighttime loss rates in the upper waters. Since daytime shoaling followed by nocturnal deepening of the mixed layer is not uncommon and was seen during the equatorial Pacific cruises (Gardner et al. 1995), the nighttime loss rates are more likely to be a combination of losses due to grazing and to mixing in the upper waters. The nighttime loss rate in the deeper waters is sometimes less than zero (thus a gain rate) since the cells in the upper, more productive waters are mixed downward (Figs. 16C, I; 17C, D).

CONCLUSIONS

Phytoplankton group-specific growth rate estimates from diel variations in flow cytometric light scattering are comparable to more conventional determinations of growth rates for *Prochlorococcus* and small eukaryotes. *Prochlorococcus* had a higher growth rate in the equatorial Pacific than in the Sargasso Sea during these cruises. *Synechococcus* growth rates were similar between the two areas, though the rates presented here are most likely underestimates due to incomplete phasing. Growth rates for the eukaryotic phytoplankton (ultra and nano) in the equatorial Pacific were near one doubling per day in the upper waters. In the Sargasso Sea, the nanophytoplankton rates approached one doubling per day, but were more often lower than that. Generally, phytoplankton growth rates were closely matched by *in situ* loss rates over the course of a day. Nighttime loss rates were more likely to be higher than daytime loss rates, at least in part due to increased mixing due to nocturnal deepening of the mixed layer.

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Chapter 6

Conclusions

In this thesis, I utilize flow cytometric measurements of phytoplankton light scattering and cell concentrations made over the diel cycle in the field in two ways. The first is to relate them to the bulk water optical property beam attenuation in order to explain the diel variations observed. The second is to relate them to growth processes of phytoplankton groups and thus estimate *in situ* phytoplankton group-specific growth and loss rates in the oceans. In order to accomplish these intents, I have collected field data from three diel sampling experiments in the equatorial Pacific and four in the Sargasso Sea. In addition, I have performed laboratory experiments to investigate the variations over the diel cycle in optical properties of a cultured marine nanophytoplankton, *Nannochloris* sp. Calibrations from the laboratory experiments, along with theoretical calculations, are then applied to the field data to interpret the patterns seen.

The laboratory experiments on *Nannochloris*, grown at a range of light levels and under both artificial and natural light, showed pronounced diel patterns, with a minimum near dawn and a maximum near dusk, in cell size and cell-specific attenuation, absorption, flow cytometric light scatter, and carbon. The diel variations in the attenuation cross section were found to be primarily influenced by the changes in cell size due to growth and division, while changes in refractive index had only a small effect. These results are important, since, though systematic studies on variability in optical properties over the diel cycle have been performed on a marine diatom (Stramski and Reynolds 1993) and *Synechococcus* (Stramski et al. 1995),

these two groups have division patterns unlike those of most other phytoplankton. This is the first diel study on the optical properties of a more typically dividing marine phytoplankton which is of a size expected to be an important contributor to optical properties in the oceans.

Field observations of diel variations in bulk beam attenuation, with a minimum near dawn and a maximum near dusk, have been presumed to reflect particle concentration changes over the course of the day and thus used to estimate particle production rates (Siegel et al. 1989). From flow cytometric measurements of individual phytoplankton cells in the field, it is clear that the diel variations in mean forward light scatter for phytoplankton populations are similar to those in beam attenuation, whereas cell concentrations generally have the opposite pattern. From laboratory studies, it has been suggested that a portion of diel variations in cell optical properties may be carbon-independent (increases in cell size without increases in cell carbon, Ackleson et al. 1993). Diel sampling experiments on cultures of *Nannochloris* have revealed increasing cell size during the day accompanied by increasing cell carbon as the cells photosynthesize and add carbon (grow) during the day, followed by division (and corresponding decreases in cell size and carbon) during the night. Using a combination of empirical calibrations relating beam attenuation to flow cytometric measurements of pure cultures of phytoplankton in the laboratory, and Mie theory, the contributions of different groups of phytoplankton to the diel variations in beam attenuation in the field were estimated. The results indicate that the phytoplankton assemblage measured by flow cytometry can account

for essentially all of the diel variation in the beam attenuation signal. These diel variations are equated with growth processes of the cells over the diel cycle.

Diel variations in beam attenuation have been used to estimate primary production by applying a constant value for carbon-specific beam attenuation (Siegel et al. 1989, Cullen et al. 1992). However, if the carbon-specific beam attenuation (c_c^*) is different for different species (Ackleson et al. 1993, Stramski et al. 1995) or is not constant over the diel cycle (Stramski et al. 1995), then this approach may not be appropriate. In addition, the c_c^* utilized was obtained from previous studies relating beam attenuation to suspended particulate material (SPM) and assuming that 25% of SPM is due to particulate organic carbon (POC). This c_c^* was then applied to the diel variation seen in beam attenuation (Δc). The results presented in this thesis indicate that 37-58% of the total beam attenuation due to particles is due to the phytoplankton. However, I found that nearly all of the diel variation in beam attenuation can be accounted for by phytoplankton. Thus Δc should be directly converted to production using a c_c^* determined for phytoplankton. However, there are indications that c_c^* is not constant over the diel cycle; it increased by up to 25% from dawn to dusk for cultures of *Nannochloris*. In addition, *Synechococcus* appears to have a lower c_c^* than *Nannochloris*, which could be important. If different phytoplankton groups are not contributing to production in the same manner that they are contributing to changes in optical properties, then estimates of primary production cannot be made from bulk measurements of beam attenuation without knowing the composition of the phytoplankton community.

Since diel variations in mean light scattering of phytoplankton populations were equated with growth processes, flow cytometric measurements of phytoplankton light scattering over the diel light cycle were used to estimate *in situ* phytoplankton group-specific growth rates in the equatorial Pacific and the Sargasso Sea. Though there are a number of assumptions inherent in this approach, it yielded promising results for some groups. The method used to estimate growth rates works well for *Prochlorococcus* and appears to work for small eukaryotic phytoplankton, but leads to underestimates for *Synechococcus* and larger eukaryotes. Estimated growth rates for *Prochlorococcus* reached one division per day in the upper waters of the equatorial Pacific, but were about half that in the Sargasso Sea, consistent with other methods (Vaulot et al. 1995, Goericke and Welschmeyer 1993). For the eukaryotic phytoplankton, growth rates in the equatorial Pacific were near one division per day in the upper waters; in the Sargasso Sea, the growth rates approached that, but were more often lower. From the concomitant measurements of cell concentrations in the field, loss rates were also estimated. In general, phytoplankton growth rates were closely matched by *in situ* loss rates over the course of a day in both the equatorial Pacific and the Sargasso Sea. Estimating growth rates from flow cytometric measurements of phytoplankton light scatter over the diel cycle is a particularly attractive method since *in situ* processes are sampled without disturbance from incubations or manipulation, different phytoplankton groups can be distinguished, and the method could potentially be used with an *in situ* flow cytometer.

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Appendix A

Calibration of forward light scatter and cell volume

A laboratory calibration experiment was performed to determine the relationship between cell volume and flow cytometric forward light scatter (FLS) for a range of phytoplankton sizes. This calibration may be useful for conversion of flow cytometric light scatter to cell volume, which can then be converted to cell carbon (Strathman 1967, Chisholm 1992), to estimate oceanic distributions of phytoplankton volume or biomass.

Culturing - Each of the thirteen phytoplankton cultures (listed in Table 1) was grown in f/2 medium at 22° C in a continuous light incubator at approximately 90 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$. When the cells were expected to be in exponential phase, each tube was sampled and the following measurements were made.

Coulter Multisizer - Each culture was diluted with 0.22 μm filtered seawater to obtain a coincidence rate < 5% when analyzed on a Coulter Multisizer. The orifice size used depended on the cell size (30 μm orifice for the smallest two, 50 μm orifice for the next smallest three, and 70 μm orifice for the largest eight; see Table 1). Volumes are averages of at least three replicate samples analyzed for each culture.

Flow cytometer - Each culture was analyzed at approximately the same time using two different flow cytometers. In each case, replicate samples were analyzed live (after dilution with 0.22 μm filtered seawater), and then replicate preserved and frozen samples were analyzed (see below for preservation protocol). Both the flow cytometers were modified to analyze 50 ml seawater samples at 5-10 ml min^{-1} (Olson et al. 1991, 1993) and had a Profile flow cell (Coulter, EPICS) instead of a jet-in-air

Table 1. Phytoplankton cultures used, arranged in order of increasing cell volume. Listed are the culture name (identifying information or abbreviation used in the Olson stock culture collection), average Coulter volume (μm^3), and average forward light scatter (FLS) for the culture analyzed live on Epics #1, preserved on Epics #1, live on Epics #2, and preserved on Epics #2. All FLS data are expressed relative to 3.79 μm beads.

Culture	Vol. μm^3	FLS (E#1 Live)	FLS (E#1 Pres)	FLS (E#2 Live)	FLS (E#2 Pres)
<i>Pycnococcus provasoli</i> (Ω 48-23)	9	0.637	0.627	0.612	0.603
<i>Nannochloris</i> sp. (Nanno)	10	0.626	0.523	0.629	0.513
<i>Minutocellus polymorphus</i> (Hudson sort)	20	0.697	0.738	0.680	0.627
<i>Emiliana huxleyi</i> (Eh 12-1)	35	1.042	0.869	0.927	0.783
<i>Phaeodactylum tricornutum</i> (Phaeo)	38	0.311	0.235	0.384	0.281
<i>Cryptomonas</i> sp. (Eel Pond)	113	1.055	1.060	1.301	1.390
<i>Cylindrotheca fusiformis</i> (Cf)	172	0.816	0.828	0.579	0.668
<i>Dunaliella tertiolecta</i> (Dun)	200	1.811	1.528	1.356	1.259
<i>Hymenomonas carterae</i> (NCO)	356	2.891	2.251	2.258	1.990
<i>Amphidinium carteri</i> (Amphi)	380	3.524	3.233	2.311	2.494
<i>Hymenomonas carterae</i> (COII)	442	4.802	3.814	3.175	2.990
<i>Thalassiosira weissflogii</i> (Tw)	915	3.826	4.042	2.703	3.223
<i>Syracosphaera elongata</i> (SE62)	2095	5.015	4.076	4.061	3.804

as used for a previous calibration (Olson et al. 1989). One flow cytometer (referred to as E#1) was an Epics V with a standard photodiode (with wide obscuration bar) used to collect the forward light scatter signal. The second flow cytometer (E#2) was an EPICS 753 with a PMT to collect the forward light scatter signal. The window of the PMT was positioned 192 mm from the center of the flow cell. Part of the PMT was covered with black tape so that only a portion of the light reaching

the PMT was collected (a rectangle 7.5 mm high and 11 mm wide was unmasked). The estimated angles of collection are 3°-19°.

Preservation - For each phytoplankton culture, 14 1-ml aliquots were dispensed into cryovials, 5 μ L 25% glutaraldehyde (final concentration 0.125%) was added, the vials were mixed and left in the dark for 10 minutes before being frozen in liquid nitrogen. One set of these preserved samples were analyzed on the flow cytometers (as described above) and the rest remain in liquid nitrogen for calibrating future configurations.

The relationship between flow cytometric forward light scatter and cell volume (Fig. 1) is strongly correlated for the phytoplankton cultures tested (excluding the pennate diatoms, *Phaeodactylum tricornutum* and *Cylindrotheca fusiformis*, which have lower FLS than would be expected from their cell volume, Olson et al. 1989). The regression equations are as follows (FLS is relative to 3.79 μ m beads and volume is in μ m³):

$$\log (\text{FLS, E\#1 Live}) = 0.43 * \log (\text{volume}) - 0.66 \quad r^2 = 0.92$$

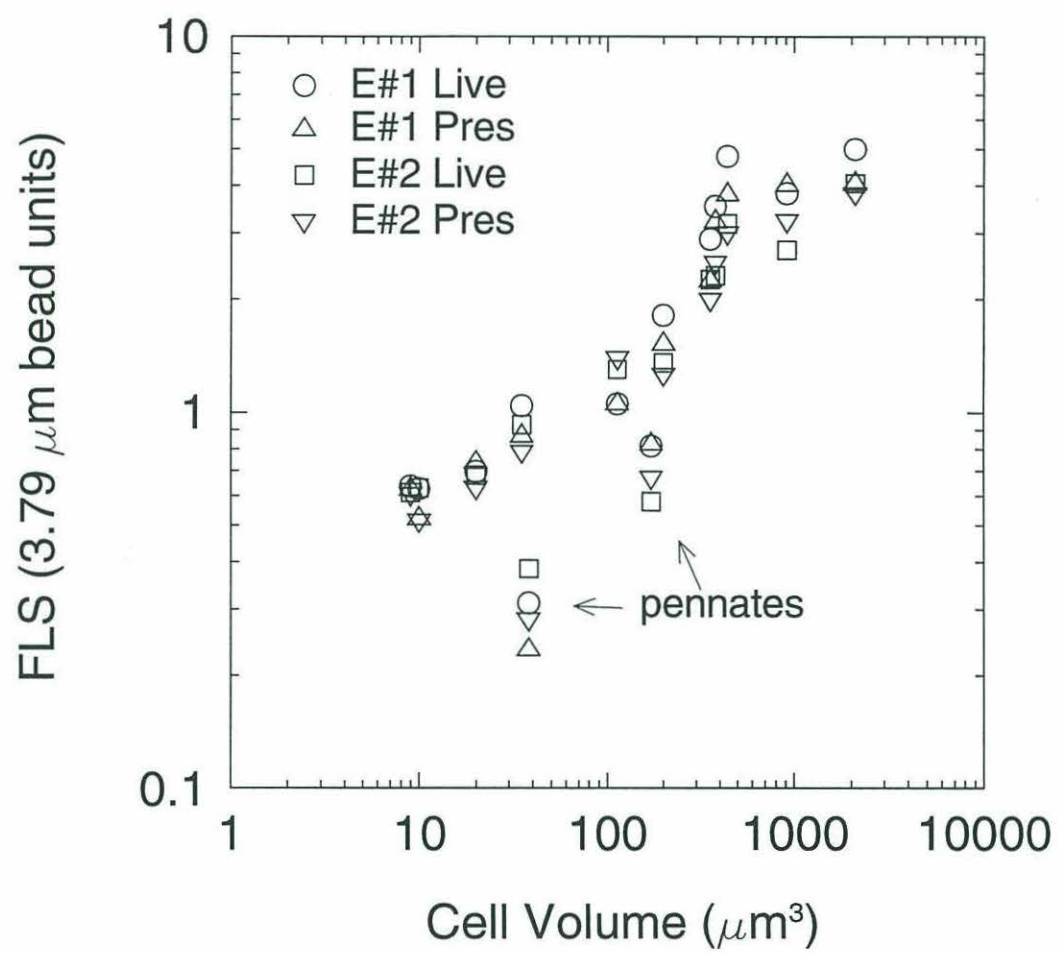
$$\log (\text{FLS, E\#1 Pres}) = 0.41 * \log (\text{volume}) - 0.67 \quad r^2 = 0.93$$

$$\log (\text{FLS, E\#2 Live}) = 0.36 * \log (\text{volume}) - 0.59 \quad r^2 = 0.96$$

$$\log (\text{FLS, E\#2 Pres}) = 0.39 * \log (\text{volume}) - 0.67 \quad r^2 = 0.96$$

Statistical analysis to compare the slopes and elevations of the equations for E#1 Live and E#2 Live indicate that the lines are not significantly different. Data from these two different configurations for FLS, each of which was used in the laboratory and field experiments described in this thesis, are considered to be comparable.

Figure 1. Relationship between flow cytometric forward light scatter (FLS expressed relative to 3.79 μm beads) and cell volume (μm^3) for Epics#1 analyzed live (E#1 Live) or preserved (E#1 Pres) and Epics#2 analyzed live (E#2 Live) or preserved (E#2 Pres). The thirteen phytoplankton cultures used are listed in Table 1.



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Appendix B

Ancillary data for Chapter 5

Presented here are time series data of forward light scatter and cell concentrations from diel sampling experiments in the equatorial Pacific and the Sargasso Sea (described below). Data from two selected experiments (TT8D1 in the equatorial Pacific and Iselin in the Sargasso Sea) are included in Chapter 5 (Figs. 1-3). The rest of the data (which are used to estimate the growth and loss rates presented in Chapter 5) are included in this appendix.

Field Studies

Equatorial Pacific - Diel sampling was performed on three occasions during the two JGOFS Time Series cruises in the equatorial Pacific in 1992 (at 0°N, 140° W). During R.V. *Thomas Thompson* 008, one diel sampling was performed (TT8D1 on 1 April, see Chapter 5, Figs. 1, 2) and during cruise 012, two diel samplings were performed (TT12D1 on 5 October and TT12D2 on 11 October; Figs. 1-4). In each case, CTD casts were made every three hours for 24 hours and samples for flow cytometric analysis were taken with Niskin bottles at 0, 15, 30, 45, 60, 75, 90, and 100 m.

Sargasso Sea - Diel samplings were performed on four occasions during three cruises in the Sargasso Sea. During R.V. *Endeavor* 232 in January 1992, two diel samplings were performed (EnD1 for 48-h at 27°N 68°W and EnD2 for 28-h at 35°N 68°W; Figs. 5, 6). During R.V. *Columbus Iselin* 93-06 (July 1993), one 40-h

sampling was performed (Iselin at 35°N 69°W; see Chapter 5, Fig. 3). During R.V. *Cape Hatteras* 0694 (May 1994), one 48-h sampling was performed (Hat at 35°N 71°W; Fig. 7). For each sampling, a holey-sock drogue set at 25 m was launched and tracked in order to follow the same water mass during the sampling. CTD casts were made every two hours and samples for flow cytometric analyses were taken with Niskin bottles at four to six depths (depths varied with each sampling).

Fig. 1. Time series of *Prochlorococcus* A) forward light scatter (relative to 3.79 μm bead) and B) cell concentration, and *Synechococcus* C) forward light scatter and D) cell concentration in the equatorial Pacific in October 1992 (TT12D1).

Forward Light Scatter
Cells ml⁻¹

TT12D1

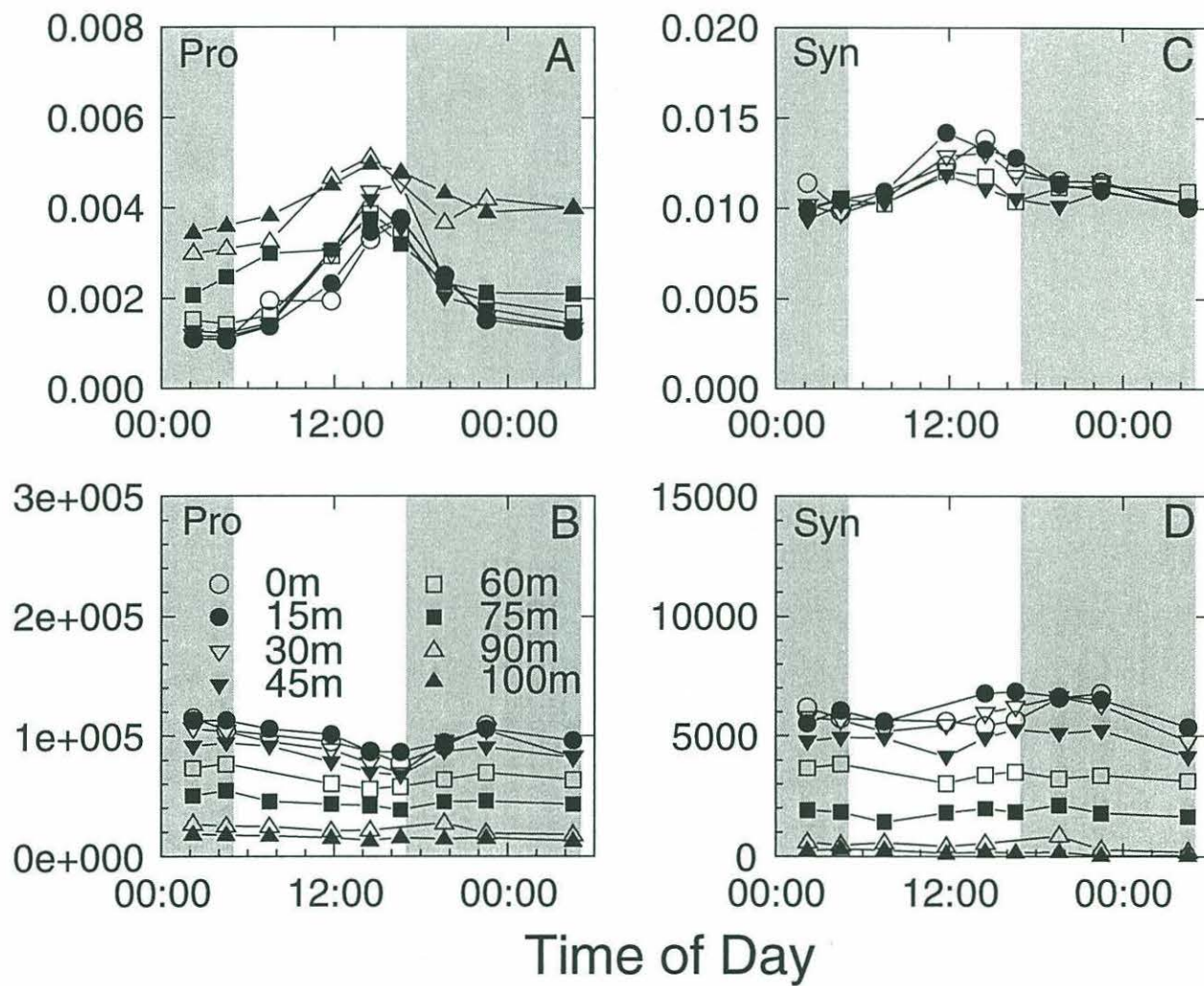


Fig. 2. Time series of ultraphytoplankton A) forward light scatter (relative to 3.79 μm bead) and B) cell concentration, and nanophytoplankton C) forward light scatter and D) cell concentration in the equatorial Pacific in October 1992 (TT12D1).

Forward Light Scatter
Cells ml⁻¹

TT12D1

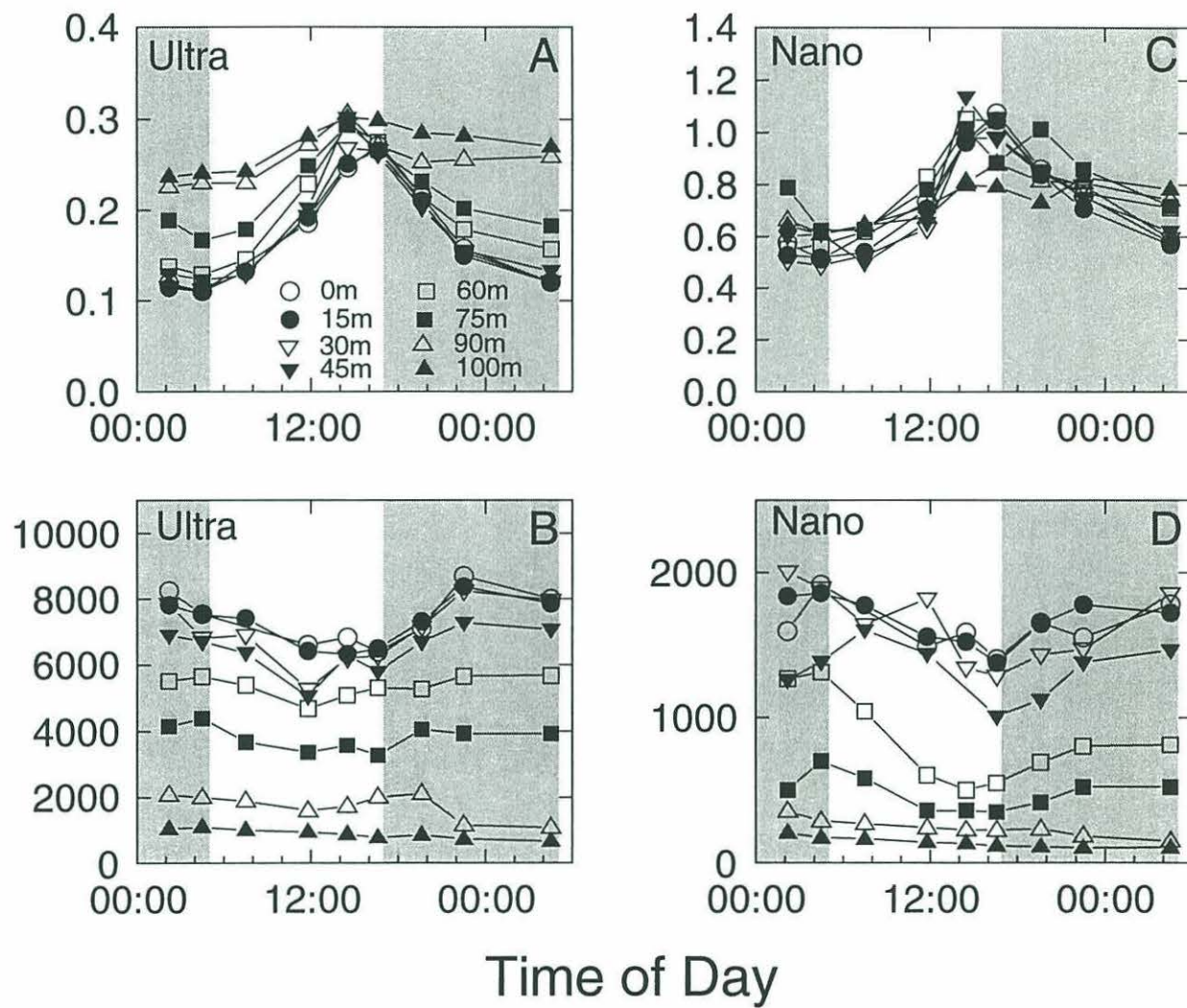


Fig. 3. Time series of *Prochlorococcus* A) forward light scatter (relative to 3.79 μm bead) and B) cell concentration, and *Synechococcus* C) forward light scatter and D) cell concentration in the equatorial Pacific in October 1992 (TT12D2).

Forward Light Scatter

Cells ml⁻¹

TT12D2

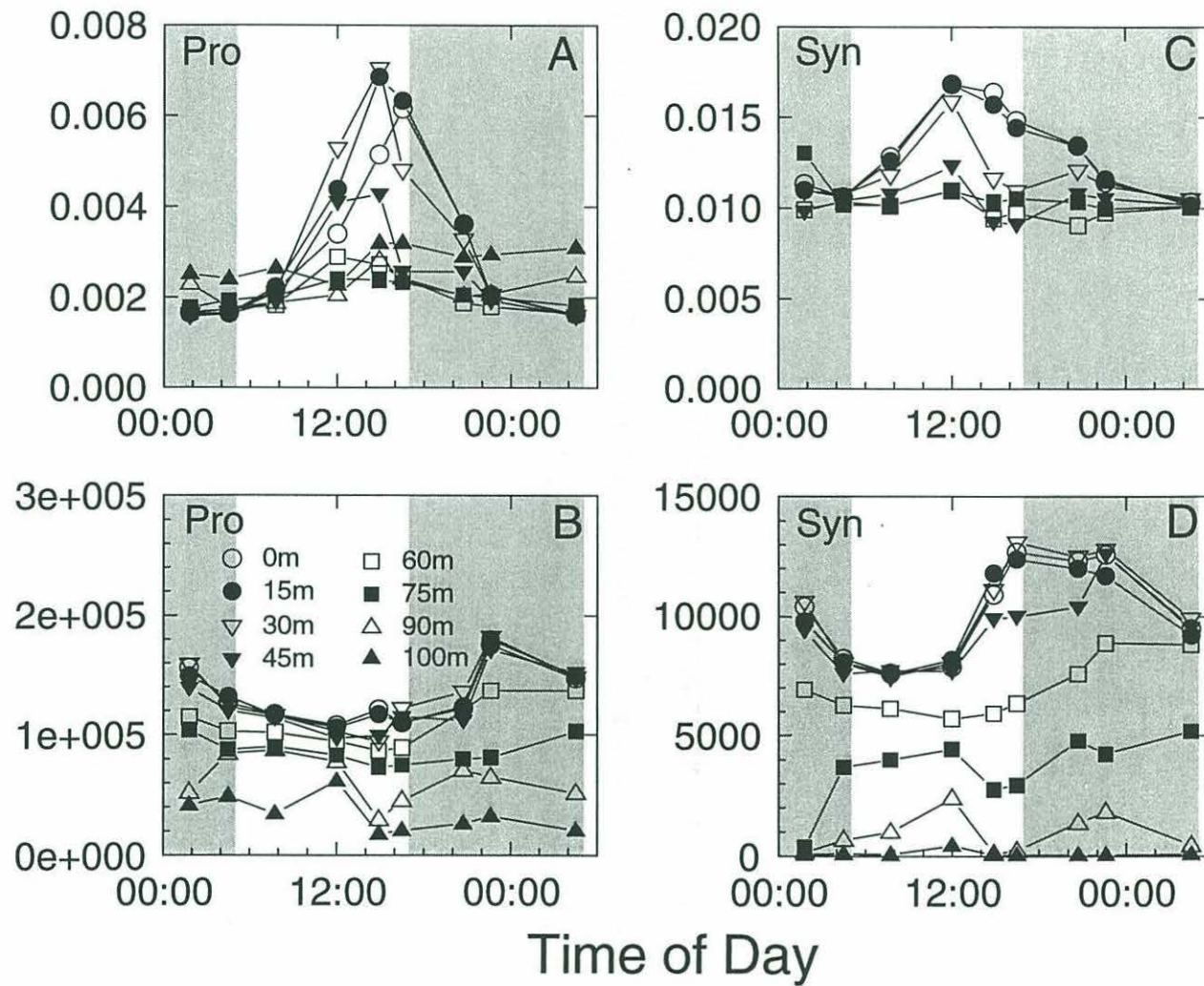


Fig. 4. Time series of ultraphytoplankton A) forward light scatter (relative to 3.79 μm bead) and B) cell concentration, and nanophytoplankton C) forward light scatter and D) cell concentration in the equatorial Pacific in October 1992 (TT12D2).

TT12D2

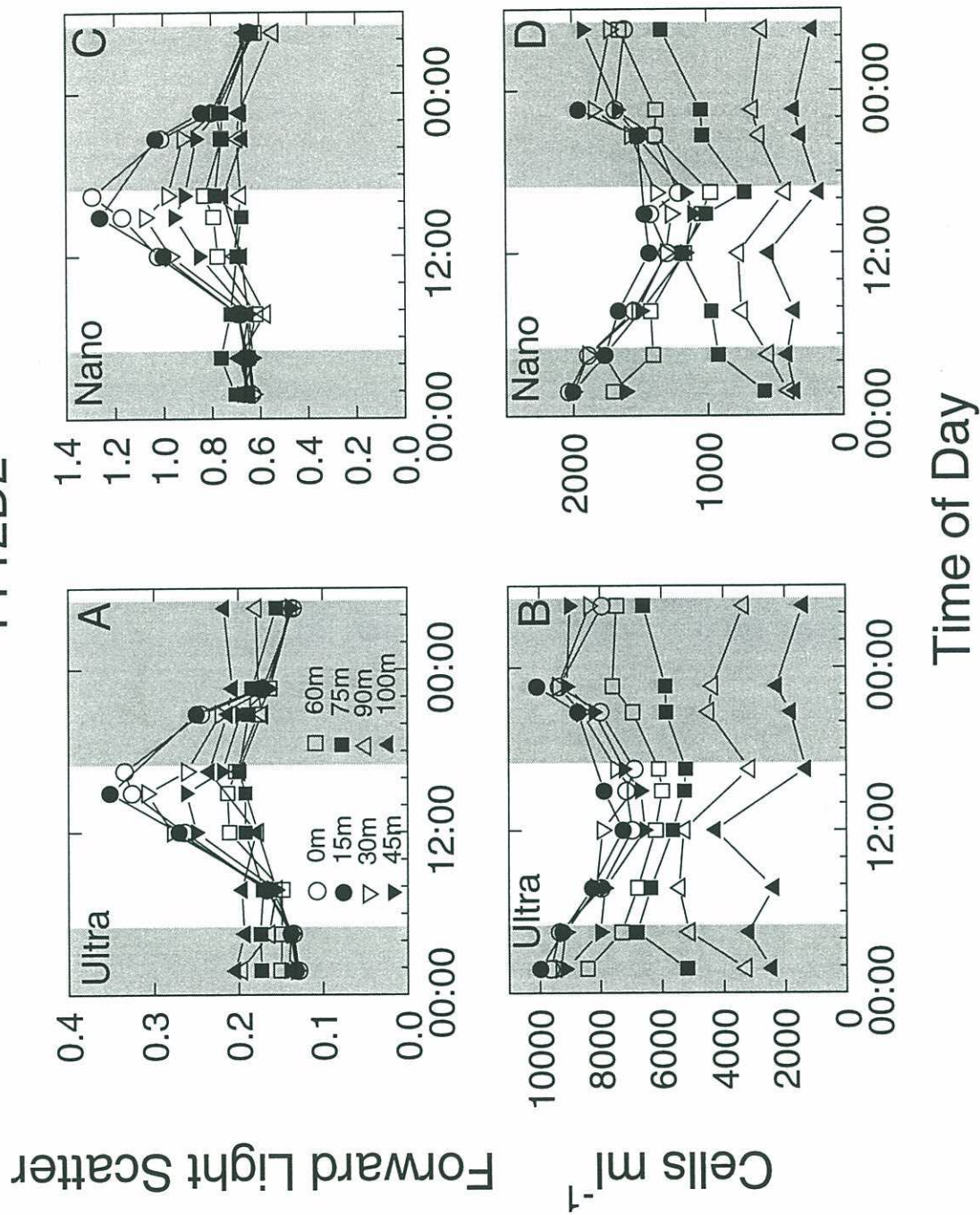


Fig. 5. Time series of nanophytoplankton A) forward light scatter (relative to 3.79 μm beads) and B) cell concentration during a 48-hour diel sampling experiment in January 1992 in the Sargasso Sea at 27°N 68°W (R.V. *Endeavor*, EnD1).

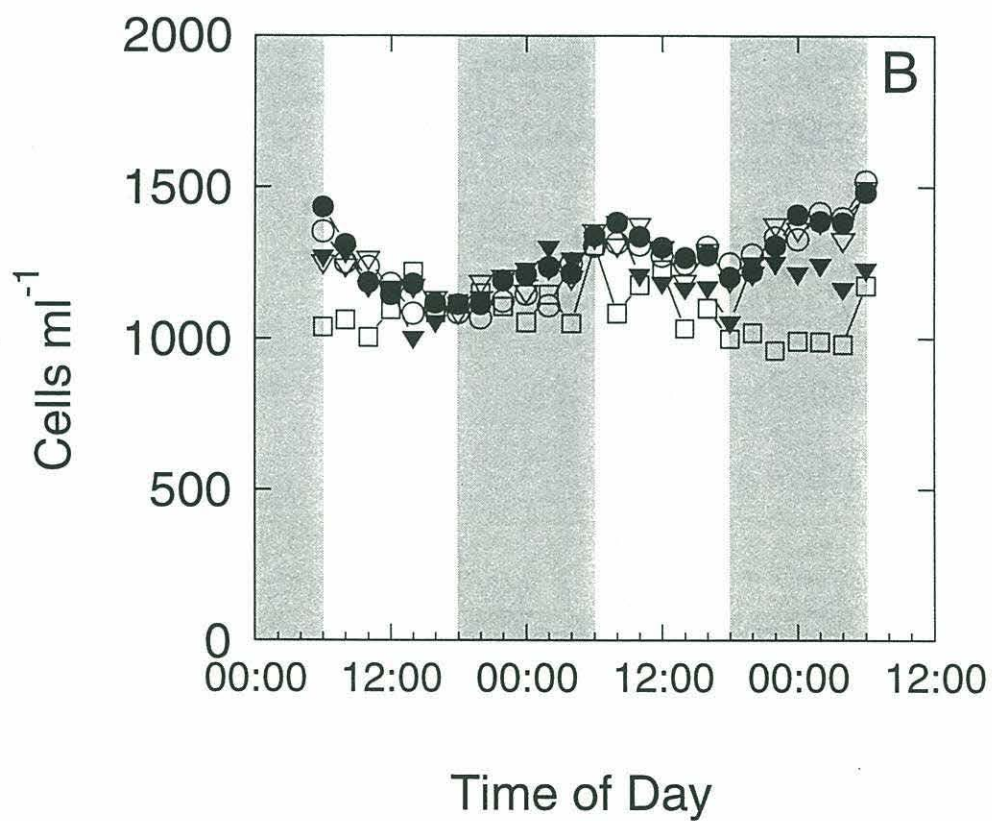
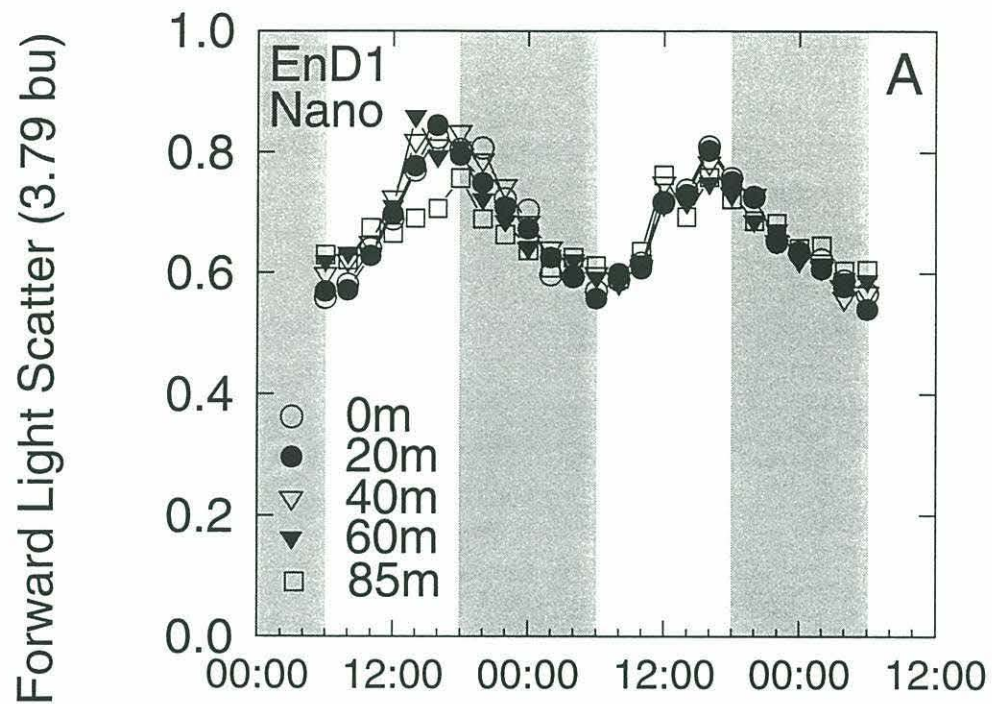
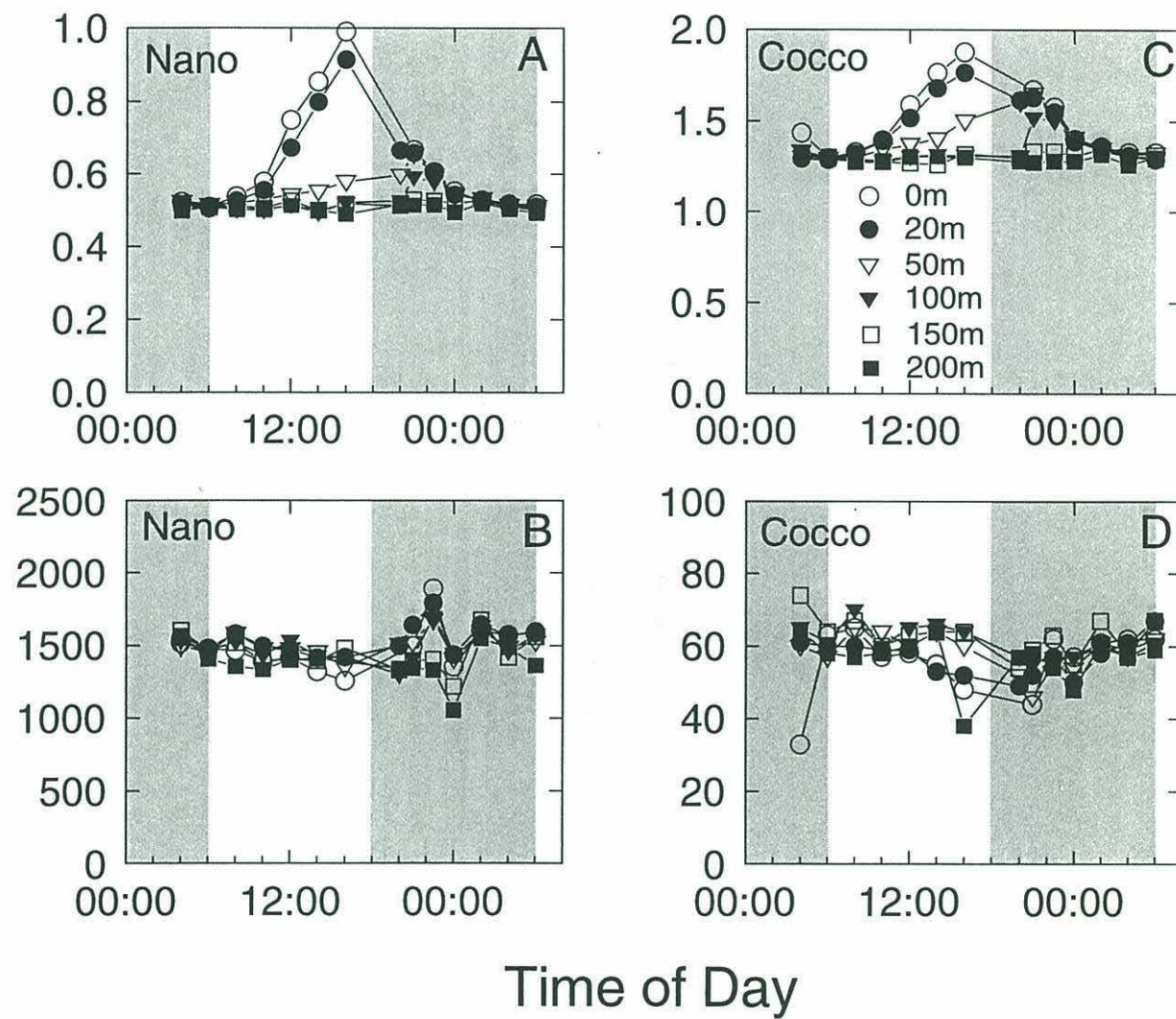


Fig. 6. Time series of nanophytoplankton A) forward light scatter (relative to 3.79 μm bead) and B) cell concentration, and coccolithophore C) forward light scatter and D) cell concentration during a 28-hour diel sampling experiment in January 1992 in the Sargasso Sea at 35°N 68°W (R.V. *Endeavor*, EnD2).

Forward Light Scatter
Cells ml⁻¹

EnD2



Time of Day

Fig. 7. Time series of nanophytoplankton A) forward light scatter (relative to 3.79 μm bead) and B) cell concentration during a 48-hour diel sampling experiment in May 1994 in the Sargasso Sea at 35°N 71°W (R.V. *Cape Hatteras*, Hat).

